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(71) Applicants (for all designated States except US): OREGON

(71) Applicants (for all designated States except US): OREGON HEALTH SCIENCES UNIVERSITY [US/US]; 3181 S.W. Sam Jackson Park Road, Portland, OR 97201 (US). DANA-FARBER CANCER INSTITUTE [US/US]; 44 Binney Street, Boston, M A 02115 (US).

(72) Inventors; and

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(75) Inventors/Applicants (for US only): BAKER, Sean, M. [GB/US]; 2520 S.W. Beaverton Highway, Portland, OR 97201 (US). BOLLAG, Roni, J. [US/US]; 231 Watervale Road, Martinez, GA 30907 (US). KOLODNER, Richard, D. [US/US]; 241 Perkins Street, Jamaica Plain, MA 02130 (US). BRONNER, C., Eric [US/US]; Apartment 110, 3211 S.W. Tenth, Portland, OR 97201 (US). LISKAY, Robert, M. [US/US]; 1110 Terrace Drive, Lake Oswego, OR 97034 (US).

(74) Agent: VAN RYSSELBERGHE, Pietre; Kolisch, Hartwell, Dickinson, McCormack & Heuser, Suite 200, 520 S.W. Yambill, Portland, OR 97204 (US).

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(54) Title: COMPOSITIONS AND METHODS RELATING TO DNA MISMATCH REPAIR GENES

(57) Abstract

Genomic sequences of human mismatch repair genes are described, as are methods of detecting mutations and/or polymorphisms in those genes. Also described are methods of diagnosing cancer susceptibility in a subject, and methods of identifying and classifying mismatch-repair-defective numors. In particular, sequences and methods relating to human mutl. homologs, hMLH1 and hPMS1 genes are provided.

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COMPOSITIONS AND METHODS RELATING TO DNA MISMATCH REPAIR GENES

This invention was made with government support under Agreement No. GM 32741 and Agreement No. HG00395/GM50006 awarded by the National Institute of Health in the General Sciences Division. The government has certain rights in the invention.

This application is a continuation-in-part from U.S. Patent Application Serial No. 08/209,521, titled: MAMMALIAN DNA MISMATCH REPAIR GENES PMS1 AND MLH1, filed on March 8, 1994, which is a continuation-in-part from U.S. Patent Application Serial No. 08/168,877, filed on December 17, 1993. All of the above patent applications are incorporated by reference.

Field of the Invention

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The present invention involves DNA mismatch repair genes. In particular, the invention relates to identification of mutations and polymorphisms in DNA mismatch repair genes, to identification and characterization of DNA mismatch-repair-defective tumors, and to detection of genetic susceptibility to cancer.

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Background

In recent years, with the development of powerful cloning and amplification techniques such as the polymerase chain reaction (PCR), in combination with a rapidly accumulating body of information concerning the structure and location of numerous human genes and markers, it has become practical and advisable to collect and analyze samples of DNA or RNA from individuals who are members of families which are identified as exhibiting a high frequency of certain genetically transmitted disorders. For example, screening procedures are routinely used to screen for genes involved in sickle cell anemia, cystic fibrosis, fragile X chromosome syndrome and multiple sclerosis. For some types of disorders, early diagnosis can greatly improve the person's long-term prognosis by, for example, adopting an aggressive diagnostic routine, and/or by

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making life style changes if appropriate to either prevent or prepare for an anticipated problem.

Once a particular human gene mutation is identified and linked to a disease, development of screening procedures to identify high-risk individuals can be relatively straight forward. For example, after the structure and abnormal phenotypic role of the mutant gene are understood, it is possible to design primers for use in PCR to obtain amplified quantities of the gene from individuals for testing. However, initial discovery of a mutant gene, i.e., its structure, location and linkage with a known inherited health problem, requires substantial experimental effort and creative research strategies.

One approach to discovering the role of a mutant gene in causing a disease begins with clinical studies on individuals who are in families which exhibit a high frequency of the disease. In these studies, the approximate location of the disease-causing locus is determined indirectly by searching for a chromosome marker which tends to segregate with the locus. A principal limitation of this approach is that, although the approximate genomic location of the gene can be determined, it does not generally allow actual isolation or sequencing of the gene. For example, Lindblom et al.3 reported results of linkage analysis studies performed with SSLP (simple sequence length polymorphism) markers on individuals from a family known to exhibit a high incidence of hereditary non-polyposis colon cancer (HNPCC). Lindblom et al. found a "tight linkage" between a polymorphic marker on the short arm of human chromosome 3 (3p21-23) and a disease locus apparently responsible for increasing an individual's risk of developing colon cancer. Even though 3p21-23 is a fairly specific location relative to the entire genome, it represents a huge DNA region relative to the probable size of the mutant gene. The mutant gene could be separated from the markers identifying the locus by millions of bases. At best, such linkage studies have only limited utility for screening purposes because in order to predict one person's risk, genetic analysis must be performed with tightly linked genetic markers on a number of related individuals in the family. It is often impossible to obtain such information, particularly if affected family members are deceased. Also, informative markers may not exist in the family

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under analysis. Without knowing the gene's structure, it is not possible to sample, amplify, sequence and determine directly whether an individual carries the mutant gene.

Another approach to discovering a disease-causing mutant gene begins with design and trial of PCR primers, based on known information about the disease, for example, theories for disease state mechanisms, related protein structures and function, possible analogous genes in humans or other species, etc. The objective is to isolate and sequence candidate normal genes which are believed to sometimes occur in mutant forms rendering an individual disease prone. This approach is highly dependent on how much is known about the disease at the molecular level, and on the investigator's ability to construct strategies and methods for finding candidate genes. Association of a mutation in a candidate gene with a disease must ultimately be demonstrated by performing tests on members of a family which exhibits a high incidence of the disease. The most direct and definitive way to confirm such linkage in family studies is to use PCR primers which are designed to amplify portions of the candidate gene in samples collected from the family members. The amplified gene products are then sequenced and compared to the normal gene structure for the purpose of finding and characterizing mutations. A given mutation is ultimately implicated by showing that affected individuals have it while unaffected individuals do not, and that the mutation causes a change in protein function which is not simply a polymorphism.

Another way to show a high probability of linkage between a candidate gene mutation and disease is by determining the chromosome location of the gene, then comparing the gene's map location to known regions of disease-linked loci such as the one identified by Lindblom et al. Coincident map location of a candidate gene in the region of a previously identified disease-linked locus may strongly implicate an association between a mutation in the candidate gene and the disease.

There are other ways to show that mutations in a gene candidate may be linked to the disease. For example, artificially produced mutant forms of the gene can be introduced into animals. Incidence of the disease in animals

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carrying the mutant gene can then be compared to animals with the normal genotype. Significantly elevated incidence of disease in animals with the mutant genotype, relative to animals with the wild-type gene, may support the theory that mutations in the candidate gene are sometimes responsible for occurrence of the disease.

One type of disease which has recently received much attention because of the discovery of disease-linked gene mutations is Hereditary Nonpolyposis Colon Cancer (HNPCC).^{1,2} Members of HNPCC families also display increased susceptibility to other cancers including endometrial, ovarian, gastric and breast. Approximately 10% of colorectal cancers are believed to be HNPCC. Tumors from HNPCC patients display an unusual genetic defect in which short, repeated DNA sequences, such as the dinucleotide repeat sequences found in human chromosomal DNA ("microsatellite DNA"), appear to be unstable. This genomic instability of short, repeated DNA sequences, sometimes called the "RER+" phenotype, is also observed in a significant proportion of a wide variety of sporadic tumors, suggesting that many sporadic tumors may have acquired mutations that are similar (or identical) to mutations that are inherited in HNPCC.

Genetic linkage studies have identified two HNPCC loci thought to account for as much as 90% of HNPCC. The loci map to human chromosome 2p15-16 (2p21) and 3p21-23. Subsequent studies have identified human DNA mismatch repair gene hMSH2 as being the gene on chromosome 2p21, in which mutations account for a significant fraction of HNPCC cancers. hMSH2 is one of several genes whose normal function is to identify and correct DNA mispairs including those that follow each round of chromosome replication.

The best defined mismatch repair pathway is the *E.coli* MutHLS pathway that promotes a long-patch (approximately 3Kb) excision repair reaction which is dependent on the *mutH*, *mutL*, *mutS* and *mutU* (uvrD) gene products. The MutHLS pathway appears to be the most active mismatch repair pathway in *E.coli* and is known to both increase the fidelity of DNA replication and to act on recombination intermediates containing mispaired bases. The system has been reconstituted *in vitro*, and requires the *mutH*, *mutL*, *mutS* and uvrD (helicase II)

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proteins along with DNA polymerase III holoenzyme, DNA ligase, single-stranded DNA binding protein (SSB) and one of the single-stranded DNA exonucleases, Exo I, Exo VII or RecJ. hMSH2 is homologous to the bacterial *mutS* gene. A similar pathway in yeast includes the yeast *MSH2* gene and two *mutL*-like genes referred to as *PMS1* and *MLH1*.

With the knowledge that mutations in a human *mutS* type gene (*hMSH2*) sometimes cause cancer, and the discovery that HNPCC tumors exhibit microsatellite DNA instability, interest in other DNA mismatch repair genes and gene products, and their possible roles in HNPCC and/or other cancers, has intensified. It is estimated that as many as 1 in 200 individuals carry a mutation in either the *hMSH2* gene or other related genes which encode for other proteins in the same DNA mismatch repair pathway.

An important objective of our work has been to identify human genes which are useful for screening and identifying individuals who are at elevated risk of developing cancer. Other objects are: to determine the sequences of exons and flanking intron structures in such genes; to use the structural information to design testing procedures for the purpose of finding and characterizing mutations which result in an absence of or defect in a gene product which confers cancer susceptibility; and to distinguish such mutations from "harmless" polymorphic variations. Another object is to use the structural information relating to exon and flanking intron sequences of a cancer-linked gene, to diagnose tumor types and prescribe appropriate therapy. Another object is to use the structural information relating to a cancer-linked gene to identify other related candidate human genes for study.

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Summary of the Invention

Based on our knowledge of DNA mismatch repair mechanisms in bacteria and yeast including conservation of mismatch repair genes, we reasoned that human DNA mismatch repair homologs should exist, and that mutations in such homologs affecting protein function, would be likely to cause genetic instability, possibly leading to an increased risk of developing certain forms of human cancer.

We have isolated and sequenced two human genes, hPMS1 and hMLH1 each of which encodes for a protein involved in DNA mismatch repair. hPMS1 and hMLH1 are homologous to mutL genes found in E.coli. Our studies strongly support an association between mutations in DNA mismatch repair genes and susceptibility to HNPCC. Thus, DNA mismatch repair gene sequence information of the present invention, namely, cDNA and genomic structures relating to hMLH1 and hPMS1, make possible a number of useful methods relating to cancer risk determination and diagnosis. The invention also encompasses a large number of nucleotide and protein structures which are useful in such methods.

We mapped the location of *hMLH1* to human chromosome 3p21-23. This is a region of the human genome that, based upon family studies, harbors a locus that predisposes individuals to HNPCC. Additionally, we have found a mutation in a conserved region of the *hMLH1* cDNA in HNPCC-affected individuals from a Swedish family. The mutation is not found in unaffected individuals from the same family, nor is it a simple polymorphism. We have also found that a homologous mutation in yeast results in a defective DNA mismatch repair protein. We have also found a frameshift mutation in *hMLH1* of affected individuals from an English family. Our discovery of a cancer-linked mutations in *hMLH1*, combined with the gene's map position which is coincident with a previously identified HNPCC-linked locus, plus the likely role of the *hMLH1* gene in mutation avoidance makes the *hMLH1* gene a prime candidate for underlying one form of common inherited human cancer, and a prime candidate to screen and identify individuals who have an elevated risk of developing cancer.

hMLH1 has 19 exons and 18 introns. We have determined the location of each of the 18 introns relative to hMLH1 cDNA. We have also determined the structure of all intron/exon boundary regions of hMLH1. Knowledge of the intron/exon boundary structures makes possible efficient screening regimes to locate mutations which negatively affect the structure and function of gene products. Further, we have designed complete sets of oligonucleotide primer pairs which can be used in PCR to amplify individual complete exons together with surrounding intron boundary structures.

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We mapped the location of hPMS1 to human chromosome 7. Subsequent studies by others³⁹ have confirmed our prediction that mutations in this gene are linked to HNPCC.

The most immediate use of the present invention will be in screening tests on human individuals who are members of families which exhibit an unusually high frequency of early onset cancer, for example HNPCC. Accordingly, one aspect of the invention comprises a method of diagnosing cancer susceptibility in a subject by detecting a mutation in a mismatch repair gene or gene product in a tissue from the subject, wherein the mutation is indicative of the subject's susceptibility to cancer. In a preferred embodiment of the invention, the step of detecting comprises detecting a mutation in a human *mutL* homolog gene, for example, *hMLH1* of *hPMS1*.

The method of diagnosing preferably comprises the steps of: 1) amplifying a segment of the mismatch repair gene or gene product from an isolated nucleic acid; 2) comparing the amplified segment with an analogous segment of a wild-type allele of the mismatch repair gene or gene product; and 3) detecting a difference between the amplified segment and the analogous segment, the difference being indicative of a mutation in the mismatch repair gene or gene product which confers cancer susceptibility.

Ariother aspect of the invention provides methods of determining whether the difference between the amplified segment and the analogous wild-type segment causes an affected phenotype, i.e., does the sequence alteration affect the individual's ability to repair DNA mispairs.

The method of diagnosing may include the steps of: 1) reverse transcribing all or a portion of an RNA copy of a DNA mismatch repair gene; and 2) amplifying a segment of the DNA produced by reverse transcription. An amplifying step in the present invention may comprise: selecting a pair of oligonucleotide primers capable of hybridizing to opposite strands of the mismatch repair gene, in an opposite orientation; and performing a polymerase chain reaction utilizing the oligonucleotide primers such that nucleic acid of the mismatch repair chain intervening between the primers is amplified to become the amplified segment.

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In preferred embodiments of the methods summarized above, the DNA mismatch repair gene is *hMLH1* or *hPMS1*. The segment of DNA corresponds to a unique portion of a nucleotide sequence selected from the group consisting of SEQ ID NOS: 6-24. "First stage" oligonucleotide primers selected from the group consisting of SEQ ID NOS: 44-82 are used in PCR to amplify the DNA segment are. The invention also provides a method of using "second stage" nested primers (SEQ ID NOS: 83-122), for use with the first stage primers to allow more specific amplification and conservation of template DNA.

Another aspect of the present invention provides a method of identifying and classifying a DNA mismatch repair defective tumor comprising detecting in a tumor a mutation in a mismatch repair gene or gene product, preferably a mutL homolog (hMLH1 or hPMS1), the mutation being indicative of a defect in a mismatch repair system of the tumor.

The present invention also provides useful nucleotide and protein compositions. One such composition is an isolated nucleotide or protein structure including a segment sequentially corresponding to a unique portion of a human mutL homolog gene or gene product, preferably derived from either hMLH1 or hPMS1.

Other composition aspects of the invention comprise oligonucleotide primers capable of being used together in a polymerase chain reaction to amplify specifically a unique segment of a human *mutL* homolog gene, preferably *hMLH1* or *hPMS1*.

Another aspect of the present invention provides a probe including a nucleotide sequence capable of binding specifically by Watson/Crick pairing to complementary bases in a portion of a human *mutL* homolog gene; and a label-moiety attached to the sequence, wherein the label-moiety has a property selected from the group consisting of fluorescent, radioactive and chemiluminescent.

We have also isolated and sequenced mouse *MLH1* (*mMLH1*) and *PMS1* (*mPMS1*) genes. We have used our knowledge of mouse mismatch repair genes to construct animal models for studying cancer. The models will be useful to identify additional oncogenes and to study environmental effects on mutagenesis.

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We have produced polyclonal antibodies directed to a portion of the protein encoded by *mPMS1* cDNA. The antibodies also react with hPMS1 protein and are useful for detecting the presence of the protein encoded by a normal *hPMS1* gene. We are also producing monoclonal antibodies directed to *hMLH1* and *hPMS1*.

In addition to diagnostic and therapeutic uses for the genes, our knowledge of *hMLH1* and *hPMS1* can be used to search for other genes of related function which are candidates for playing a role in certain forms of human cancer.

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Description of the Figures

Figure 1 is a flow chart showing an overview of the sequence of experimental steps we used to isolate, characterize and use human and mouse *PMS1* and *MLH1* genes.

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Figure 2 is an alignment of protein sequences for *mutL* homologs (SEQ ID NOS: 1-3) showing two highly-conserved regions (underlined) which we used to create degenerate PCR oligonucleotides for isolating additional *mutL* homologs.

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Figure 3 shows the entire cDNA nucleotide sequence (SEQ ID NO: 4) for the human *MLH1* gene, and the corresponding predicted amino acid sequence (SEQ ID NO: 5) for the human MLH1 protein. The underlined DNA sequences are the regions of cDNA that correspond to the degenerate PCR primers that were originally used to amplify a portion of the *MLH1* gene (nucleotides 118-135 and 343-359).

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Figure 4A shows the nucleotide sequences of the 19 exons which collectively correspond to the entire hMLH1 cDNA structure. The exons are flanked by intron boundary structures. Primer sites are underlined. The exons with their flanking intron structures correspond to SEQ ID NOS: 6-24. The exons, shown in non-underlined small case letters, corespond to SEQ ID NOS: 25-43.

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Figure 4B shows nucleotide sequences of primer pairs which have been used in PCR to amplify the individual exons. The "second stage"

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amplification primers (SEQ ID NOS: 83-122) are "nested" primers which are used to amplify target exons from the amplification product obtained with corresponding "first stage" amplification primers (SEQ ID NOS: 44-82). The structures in Figure 4B correspond to the structures in Tables 2 and 3.

Figure 5 is an alignment of the predicted amino acid sequences for human and yeast (SEQ ID NOS: 5 and 123, respectively) MLH1 proteins. Amino acid identities are indicated by boxes and gaps are indicated by dashes.

Figure 6 is a phylogenetic tree of MutL-related proteins.

Figure 7 is a two-panel photograph. The first panel (A) is a metaphase spread showing hybridization of the *hMLH1* gene of chromosome 3. The second panel (B) is a composite of chromosome 3 from multiple metaphase spreads aligned with a human chromosome 3 ideogram. The region of hybridization is indicated in the ideogram by a vertical bar.

Figure 8 is a comparison of sequence chromatograms from affected and unaffected individuals showing identification of a C to T transition mutation that produces a non-conservative amino acid substitution at position 44 of the hMLH1 protein.

Figure 9 is an amino acid sequence alignment (SEQ ID NOS: 124-131) of the highly-conserved region of the MLH family of proteins surrounding the site of the predicted amino acid substitution. Bold type indicates the position of the predicted serine to phenylalanine amino acid substitution in affected individuals. Also highlighted are the serine or alanine residues conserved at this position in MutL-like proteins. Bullets indicate positions of highest amino acid conservation. For the MLH1 protein, the dots indicate that the sequence has not been obtained. Sequences were aligned as described below in reference to the phylogenetic tree of Figure 6.

Figure 10 shows the entire nucleotide sequence for hPMS1 (SEQ ID NO: 132).

Figure 11 is an alignment of the predicted amino acid sequences for human and yeast PMS1 proteins (SEQ ID NOS: 133 and 134, respectively). Amino acid identities are indicated by boxes and gaps are indicated by dashes.

Figure 12 is a partial nucleotide sequence of mouse MLH1 mMLH1) cDNA (SEQ ID NO: 135).

Figure 13 is a comparison of the predicted amino acid sequence for mMLH1 and hMLH1 proteins (SEQ ID NOS: 136 and 5, respectively).

Figure 14 shows the cDNA nucleotide sequence for mouse PMS1 (mPMS1) (SEQ ID NO: 137).

Figure 15 is a comparison of the predicted amino acid sequences for mPMS1 and hPMS1 proteins (SEQ ID NOS: 138 and 133, respectively).

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Definitions

gene - "Gene" means a nucleotide sequence that contains a complete coding sequence. Generally, "genes" also include nucleotide sequences found upstream (e.g. promoter sequences, enhancers, etc.) or downstream (e.g. transcription termination signals, polyadenylation sites, etc.) of the coding sequence that affect the expression of the encoded polypeptide.

gene product - A "gene product" is either a DNA or RNA (mRNA) copy of a portion of a gene, or a corresponding amino acid sequence translated from

mRNA.

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wild-type - The term "wild-type", when applied to nucleic acids and proteins of the present invention, means a version of a nucleic acid or protein that functions in a manner indistinguishable from a naturally-occurring, normal version of that nucleic acid or protein (i.e. a nucleic acid or protein with wild-type activity). For example, a "wild-type" allele of a mismatch repair gene is capable of functionally replacing a normal, endogenous copy of the same gene within a host cell without detectably altering mismatch repair in that cell. Different wild-type versions of the same nucleic acid or protein may or may not differ structurally from each other.

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non-wild-type - The term "non-wild-type" when applied to nucleic acids and proteins of the present invention, means a version of a nucleic acid or protein that

functions in a manner distinguishable from a naturally-occurring, normal version of that nucleic acid or protein. Non-wild-type alleles of a nucleic acid of the invention may differ structurally from wild-type alleles of the same nucleic acid in any of a variety of ways including, but not limited to, differences in the amino acid sequence of an encoded polypeptide and/or differences in expression levels of an encoded nucleotide transcript of polypeptide product.

For example, the nucleotide sequence of a non-wild-type allele of a nucleic acid of the invention may differ from that of a wild-type allele by, for example, addition, deletion, substitution, and/or rearrangement of nucleotides. Similarly, the amino acid sequence of a non-wild-type mismatch repair protein may differ from that of a wild-type mismatch repair protein by, for example, addition, substitution, and/or rearrangement of amino acids.

Particular non-wild-type nucleic acids or proteins that, when introduced into a normal host cell, interfere with the endogenous mismatch repair pathway, are termed "dominant negative" nucleic acids or proteins.

homologous - The term "homologous" refers to nucleic acids or polypeptides that are highly related at the level of nucleotide or amino acid sequence. Nucleic acids or polypeptides the are homologous to each other are termed "homologues".

The term "homologous" necessarily refers to a comparison between two sequences. In accordance with the invention, two nucleotide sequences are considered to be homologous if the polypeptides they encode are at least about 50-60% identical, preferably about 70% identical, for at least one stretch of at least 20 amino acids. Preferably, homologous nucleotide sequences are also characterized by the ability to encode a stretch of at least 4-5 uniquely specified amino acids. Both the identity and the approximate spacing of these amino acids relative to one another must be considered for nucleotide sequences to be considered to be homologous. For nucleotide sequences less than 60 nucleotides in length, homology is determined by the ability to encode a stretch of at least 4-5 uniquely specified amino acids.

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upstream/downstream - The terms "upstream" and "downstream" are artunderstood terms referring to the position of an element of nucleotide sequence. "Upstream" signifies an element that is more 5' than the reference element. "Downstream" refers to an element that is more 3' than a reference element.

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intron/exon - The terms "exon" and "intron" are art-understood terms referring to various portions of genomic gene sequences. "Exons" are those portions of a genomic gene sequence that encode protein. "Introns" are sequences of nucleotides found between exons in genomic gene sequences.

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affected - The term "affected", as used herein, refers to those members of a kindred that either have developed a characteristic cancer (e.g. colon cancer in an HNPCC lineage) and/or are predicted, on the basis of, for example, genetic studies, to carry an inherited mutation that confers susceptibility to cancer.

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unique - A "unique" segment, fragment or portion of a gene or protein means a portion of a gene or protein which is different sequentially from any other gene or protein segment in an individual's genome. As a practical matter, a unique segment or fragment of a gene will typically be a nucleotide of at least about 13 bases in length and will be sufficiently different from other gene segments so that oligonucleotide primers may be designed and used to selectively and specifically amplify the segment. A unique segment of a protein is typically an amino acid sequence which can be translated from a unique segment of a gene.

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Description of the Invention

We have discovered mammalian genes which are involved in DNA mismatch repair. One of the genes, hPMSI, encodes a protein which is homologous to the yeast DNA mismatch repair protein PMS1. We have mapped the locations of hPMSI to human chromosome 7 and the mouse PMSI gene to mouse chromosome 5, band G. Another gene, hMLHI (MutL Homolog) encodes a protein which is homologous to the yeast DNA mismatch repair protein hALHI. We have mapped the locations of hMLHI to human chromosome 3p21. 23 and to mouse chromosome 9, band E.

Studies^{1,2} have demonstrated involvement of a human DNA mismatch repair gene homolog, hMSH2, on chromosome 2p in HNPCC. Based upon linkage data, a second HNPCC locus has been assigned to chromosome 3p21-23.³ Examination of tumor DNA from the chromosome 3-linked kindreds revealed dinucleotide repeat instability similar to that observed for other HNPCC families⁶ and several types of sporadic tumors.⁷⁻¹⁰ Because dinucleotide repeat instability is characteristic of a defect in DNA mismatch repair, ^{5, 11, 12} we reasoned that HNPCC linked to chromosome 3p21-23 could result from a mutation in a second DNA mismatch repair gene.

Repair of mismatched DNA in Escherichia coli requires a number of genes including mutS, mutL and mutH, defects in any one of which result in

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elevated spontaneous mutation rates.¹³ Genetic analysis in the yeast Saccharomyces cerevisiae has identified three DNA mismatch repair genes: a mutS homolog, MSH2, ¹⁴ and two mutL homologs, PMS1¹⁶ and MLH1.⁴ Each of these three genes play an indispensable role in DNA replication fidelity, including the stabilization of dinucleotide repeats.⁵

We believe that *hMLH1* is the HNPCC gene previously linked to chromosome 3p based upon the similarity of the *hMLH1* gene product to the yeast DNA mismatch repair protein, MLH1,⁴ the coincident location of the *hMLH1* gene and the HNPCC locus on chromosome 3, and *hMLH1* missense mutations which we found in affected individuals from chromosome 3-linked HNPCC families.

Our knowledge of the human and mouse *MLH1* and *PMS1* gene structures has many important uses. The gene sequence information can be used to screen individuals for cancer risk. Knowledge of the gene structures makes it possible to easily design PCR primers which can be used to selectively amplify portions of *hMLH1* and *hPMS1* genes for subsequent comparison to the normal sequence and cancer risk analysis. This type of testing also makes it possible to search for and characterize *hMLH1* and *hPMS1* cancer-linked mutations for the purpose of eventually focusing the cancer screening effort on specific gene loci. Specific characterization of cancer-linked mutations in *hMLH1* and *hPMS1* makes possible the production of other valuable diagnostic tools such as allele specific probes which may be used in screening tests to determine the presence or absence of specific gene mutations.

Additionally, the gene sequence information for hMLHI and/or hPMSI can be used, for example, in a two hybrid system, to search for other genes of related function which are candidates for cancer involvement.

The hMLH1 and hPMS1 gene structures are useful for making proteins which are used to develop antibodies directed to specific portions or the complete hMLH1 and hPMS1 proteins. Such antibodies can then be used to isolate the corresponding protein and possibly related proteins for research and diagnostic purposes.

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The mouse MLH1 and PMS1 gene sequences are useful for producing mice that have mutations in the respective gene. The mutant mice are useful for studying the gene's function, particularly its relationship to cancer.

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Methods for Isolating and Characterizing Mammalian MLHI and PMSI Genes

human MLH1 (hMLH1), human PMS1 (hPMS1), mouse MLH1 (mPMS1) and mouse PMS1 (mPMS1). Due to the structural similarity between these genes, the

methods we have employed to isolate and characterize them are generally the same. Figure 1 shows in broad terms, the experimental approach which we used to isolate and characterize the four genes. The following discussion refers to the

step-by-step procedure shown in Figure 1.

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Step 1

Design of degenerate oligonucleotide pools for PCR

We have isolated and characterized four mammalian genes, i.e.,

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Earlier reports indicated that portions of three MutL-like proteins, two from bacteria, MutL and HexB, and one from yeast, PMS1 are highly conserved. 16,18,19 After inspection of the amino acid sequences of HexB, MutL and PMS1 proteins, as shown in Figure 2, we designed pools of degenerate oligonucleotide pairs corresponding to two highly-conserved regions, KELVEN and GFRGEA, of the MutL-like proteins. The sequences (SEQ ID NOS: 139 and 140, respectively) of the degenerate oligonucleotides which we used to isolate the four genes are:

5'-CTTGATTCTAGAGC(T/C)TCNCCNC(T/G)(A/G)AANCC-3' and 5'-AGGTCGGAGCTCAA(A/G)GA(A/G)(T/C)TNGTNGANAA-3'.

The underlined sequences within the primers are XbaI and SacI restriction endonuclease sites respectively. They were introduced in order to facilitate the cloning of the PCR-amplified fragments. In the design of the oligonucleotides, we took into account the fact that a given amino acid can be coded for by more than one DNA triplet (codon). The degeneracy within these sequences are indicated by multiple nucleotides within parentheses or N, for the presence of any base at that position.

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Step 2 Reverse transcription and PCR on poly A+ selected mRNA isolated from human cells

We isolated messenger (poly A+ enriched) RNA from cultured human cells, synthesized double-stranded cDNA from the mRNA, and performed PCR with the degenerate oligonucleotides.⁴ After trying a number of different PCR conditions, for example, adjusting the annealing temperature, we successfully amplified a DNA of the size predicted (~210bp) for a MutL-like protein.

Step 3 Cloning and sequencing of PCR-generated fragments; identification of two gene fragments representing human PMS1 and MLH1

We isolated the PCR amplified material (~210bp) from an agarose gel and cloned this material into a plasmid (pUC19). We determined the DNA sequence of several different clones. The amino acid sequence inferred from the DNA sequence of two clones showed strong similarity to other known MutL-like proteins. The predicted amino acid sequence for one of the clones was most similar to the yeast PMS1 protein. Therefore we named it hPMS1, for human PMS1. The second clone was found to encode a polypeptide that most closely resembles yeast MLH1 protein and was named, hMLH1, for human MLH1.

Step 4 Isolation of complete human and mouse *PMS1* and *MLH1* cDNA clones using the PCR fragments as probes

We used the 210bp PCR-generated fragments of the *hMLH1* and *hPMS1* cDNAs, as probes to screen both human and mouse cDNA libraries (from Stratagene, or as described in reference 30). A number of cDNAs were isolated that corresponded to these two genes. Many of the cDNAs were truncated at the 5' end. Where necessary, PCR techniques ³¹ were used to obtain the 5' -end of the gene in addition to further screening of cDNA libraries. Complete composite cDNA sequences were used to predict the amino acid sequence of the human and mouse, MLH1 and PMS1 proteins.

Step 5 Isolation of human and mouse, PMS1 and MLH1 genomic clones

Information on genomic and cDNA structure of the human *MLH1* and *PMS1* genes are necessary in order to thoroughly screen for mutations in cancer prone families. We have used human cDNA sequences as probes to isolate the genomic sequences of human *PMS1* and *MLH1*. We have isolated four cosmids and two P1 clones for *hPMS1*, that together are likely to contain most, if not all, of the cDNA (exon) sequence. For *hMLH1* we have isolated four overlapping λ-phage clones containing 5'-*MLH1* genomic sequences and four P1 clones (two full length clones and two which include the 5' coding end plus portions of the promoter region) P1 clone. PCR analysis using pairs of oligonucleotides specific to the 5' and 3' ends of the *hMLH1* cDNA, clearly indicates that the P1 clone contains the complete *hMLH1* cDNA information. Similarly, genomic clones for mouse *PMS1* and *MLH1* genes have been isolated and partially characterized (described in Step 8).

Step 6 Chromosome positional mapping of the human and mouse,

PMS1 and MLH1 genes by fluorescence in situ hybridization

We used genomic clones isolated from human and mouse *PMS1* and *MLH1* for chromosomal localization by fluorescence *in situ* hybridization (FISH).^{20,21} We mapped the human *MLH1* gene to chromosome 3p21.3-23, shown in Figure 7 as discussed in more detail below. We mapped the mouse *MLH1* gene to chromosome 9 band E, a region of synteny between mouse and human.²² In addition to FISH techniques, we used PCR with a pair of *hMLH1*-specific oligonucleotides to analyze DNA from a rodent/human somatic cell hybrid mapping panel (Coriell Institute for Medial Research, Camden, N.J.). Our PCR results with the panel clearly indicate that *hMLH1* maps to chromosome 3. The position of *hMLH1* 3p21.3-23 is coincident to a region known to harbor a second locus for HNPCC based upon linkage data.

We mapped the hPMSI gene, as shown in Figure 12, to the long (q) arm of chromosome 7 (either 7q11 or 7q22) and the mouse PMSI to chromosome 5 band G, two regions of synteny between the human and the mouse.²² We performed PCR using oligonucleotides specific to hPMSI on DNA from a

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rodent/human cell panel. In agreement with the FISH data, the location of hPMSI was confirmed to be on chromosome 7. These observations assure us that our human map position for hPMSI to chromosome 7 is correct. The physical localization of hPMSI is useful for the purpose of identifying families which may potentially have a cancer linked mutation in hPMSI.

Step 7 Using genomic and cDNA sequences to identify mutations in hPMS1 and hMLH1 genes from HNPCC Families

We have analyzed samples collected from individuals in HNPCC families for the purpose of identifying mutations in hPMS1 or hMLH1 genes. Our approach is to design PCR primers based on our knowledge of the gene structures, to obtain exon/intron segments which we can compare to the known normal sequences. We refer to this approach as an "exon-screening".

Using cDNA sequence information we have designed and are continuing to design hPMS1 and hMLH1 specific oligonucleotides to delineate exon/intron boundaries within genomic sequences. The hPMS1 and hMLH1 specific oligonucleotides were used to probe genomic clones for the presence of exons containing that sequence. Oligonucleotides that hybridized were used as primers for DNA sequencing from the genomic clones. Exon-intron junctions were identified by comparing genomic with cDNA sequences.

Amplification of specific exons from genomic DNA by PCR and sequencing of the products is one method to screen HNPCC families for mutations.^{1,2} We have identified genomic clones containing *hMLH1* cDNA information and have determined the structures of all intron/exon boundary regions which flanks the 19 exons of hMCH1.

We have used the exon-screening approach to examine the MLH1 gene of individuals from HNPCC families showing linkage to chromosome $3.^3$ As will be discussed in more detail below, we identified a mutation in the MLH1 gene of one such family, consisting of a C to T substitution. We predict that the C to T mutation causes a serine to phenylalanine substitution in a highly-conserved region of the protein. We are continuing to identify HNPCC families from whom we can obtain samples in order to find additional mutations in hMLH1 and hPMS1 genes.

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We are also using a second approach to identify mutations in hPMS1 and hMLH1. The approach is to design hPMS1 or hMLH1 specific oligonucleotide primers to produce first-strand cDNA by reverse transcription off RNA. PCR using gene-specific primers will allow us to amplify specific regions from these genes. DNA sequencing of the amplified fragments will allow us to detect mutations.

Step 8 Design targeting vectors to disrupt mouse PMS1 and MLH1 genes in ES cells; study mice deficient in mismatch repair.

We constructed a gene targeting vector based on our knowledge of the genomic mouse *PMS1* DNA structure. We used the vector to disrupt the *PMS1* gene in mouse embryonic stem cells.³⁶ The cells were injected into mouse blastocysts which developed into mice that are chimeric (mixtures) for cells carrying the *PMS1* mutation. The chimeric animals will be used to breed mice that are heterozygous and homozygous for the *PMS1* mutation. These mice will be useful for studying the role of the *PMS1* gene in the whole organism.

Human MLH1

The following discussion is a more detailed explanation of our experimental work relating to hMLH1. As mentioned above, to clone mammalian MLH 3-nes, we used PCR techniques like those used to identify the yeast MSH1, MSH2 and MLH1 genes and the human MSH2 gene. 1. 2. 4. 14 As template in the PCR, we used double-stranded cDNA synthesized from poly (A+) enriched RNA prepared from cultured primary human fibroblasts. The degenerate oligonucleotides were targeted at the N-terminal amino acid sequences KELVEN and GFRGEA (see Figure 3), two of the most conserved regions of the MutL family of proteins previously described for bacteria and yeast. 16,18,19 Two PCR products of the predicted size were identified, cloned and shown to encode a predicted amino acid sequence with homology to MutL-like proteins. These two fragments generated by PCR were used to isolate human cDNA and genomic DNA clones.

The oligonucleotide primers which we used to amplify human MutL-related sequences were 5' -

CTTGATTCTAGAGC(T/C)TCNCCNC(T/G)(A/G)AANCC-3' (SEQ ID NO: 139) and 5' - AGGTCGGAGCTCAA(A/G)GA(A/G)(T/C)TNGTNGANAA-3' (SEQ ID NO: 140). PCR was carried out in 50 µL reactions containing cDNA template, 1.0 µM each primer, 5 IU of Taq polymerase (C) 50 mM KCl, 10 mM Tris buffer pH 7.5 and 1.5 mM MgCl. PCR was carried out for 35 cycles of 1 minute at 94 °C, 1 minute at 43 °C and 1.5 minutes at 62 °C. Fragments of the expected size, approximately 212 bp, were cloned into pUC19 and sequenced. The cloned MLH1 PCR products were labeled with a random primer labeling kit (RadPrime, Gibco BRL) and used to probe human cDNA and genomic cosmid libraries by standard procedures. DNA sequencing of double-stranded plasmid DNAs was performed as previously described.

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The *hMLH1* cDNA nucleotide sequence as shown in Figure 3 encodes an open reading frame of 2268 bp. Also shown in Figure 3 is the predicted protein sequence encoded for by the *hMLH1* cDNA. The underlined DNA sequences are the regions of cDNA that correspond to the degenerate PCR primers that were originally used to amplify a portion of the *MLH1* gene (nucleotides 118-135 and 343-359).

Figure 4A shows 19 nucleotide sequences corresponding to portions of hMLH1. Each sequence includes one of the 19 exons, in its entirety, surrounded by flanking intron sequences. Target PCR primer cites are underlined. More details relating to the derivation and uses of the sequences shown in Figure 4A, are set forth below.

As shown in Figure 5, the hMLH1 protein is comprised of 756 amino acids and shares 41% identity with the protein product of the yeast DNA mismatch repair gene, MLH1.⁴ The regions of the hMLH1 protein most similar to yeast MLH1 correspond to amino acids 11 through 317, showing 55% identity, and the last 13 amino acids which are identical between the two proteins. Figure 5 shows an alignment of the predicted human MLH1 and S. cerevisiae MLH1 protein sequences. Amino acid identities are indicated by boxes, and gaps are indicated by dashes. The pair wise protein sequence alignment was performed with DNAStar MegAlign using the clustal method.²⁷ Pair wise alignment parameters were a ktuple of 1, gap penalty of 3, window of 5 and diagonals of 5.

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Furthermore, as shown in Figure 13, the predicted amino acid sequences of the human and mouse MLH1 proteins show at least 74% identity.

Figure 6 shows a phylogenetic tree of MutL-related proteins. The phylogenetic tree was constructed using the predicted amino acid sequences of 7 MutL-related proteins: human MLH1; mouse MLH1; S. cerevisiae MLH1; S. cerevisiae PMS1; E. coli; MutL; S. typhimurium MutL and S. pneumoniae HexB. Required sequences were obtained from GenBank release 7.3. The phylogenetic tree was generated with the PILEUP program of the Genetics Computer Group software using a gap penalty of 3 and a length penalty of 0.1. The recorded DNA sequences of hMLH1 and hPMS1 have been submitted to GenBank.

hMLH1 Intron Location and Intron/Exon Boundary Structures

In our previous U.S. Patent Application No. 08/209,521, we described the nucleotide sequence of a complimentary DNA (cDNA) clone of a human gene, hMLH1. The cDNA sequence of hMLH1 (SEQ ID NO: 4) is presented in this application in Figure 3. We note that there may be some variability between individuals hMLH1 cDNA structures, resulting from polymorphisms within the human population, and the degeneracy of the genetic code.

In the present application, we report the results of our genomic

sequencing studies. Specifically, we have cloned the human genomic region that includes the *hMLH1* gene, with specific focus on individual exons and surrounding intron/exon boundary structures. Toward the ultimate goal of designing a comprehensive and efficient approach to identify and characterize mutations which confer susceptibility to cancer, we believe it is important to know the wild-type sequences of intron structures which flank exons in the *hMLH1* gene. One advantage of knowing the sequence of introns near the exon boundaries, is that it makes it possible to design primer pairs for selectively amplifying entire individual exons. More importantly, it is also possible that a mutation in an intron region, which, for example, may cause a mRNA splicing error, could result

in a defective gene product, i.e., susceptibility to cancer, without showing any abnormality in an exon region of the gene. We believe a comprehensive

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screening approach requires searching for mutations, not only in the exon or cDNA, but also in the intron structures which flank the exon boundaries.

We have cloned the human genomic region that includes hMLH1 using approaches which are known in the art, and other known approaches could have been used. We used PCR to screen a P1 human genomic library for the hMLH1 gene. We obtained four clones, two that contained the whole gene and two which lacked the C-terminus. We characterized one of the full length clones by cycle sequencing, which resulted in our definition of all intron/exon junction sequences for both sides of the 19 hMLH1 exons. We then designed multiple sets of PCR primers to amplify each individual exon (first stage primers) and verified the sequence of each exon and flanking intron sequence by amplifying several different genomic DNA samples and sequencing the resulting fragments using an ABI 373 sequencer. In addition, we have determined the sizes of each hMLH1 exon using PCR methods. Finally, we devised a set of nested PCR primers (second stage primers) for reamplification of individual exons. We have used the second stage primers in a multi-plex method for analyzing HNPCC families and tumors for hMLHI mutations. Generally, in the nested PCR primer approach, we perform a first multi-plex amplification with four to eight sets of "first stage" primers, each directed to a different exon. We then reamplify individual exons from the product of the first amplification step, using a single set of second stage primers. Examples and further details relating to our use of the first and second stage primers are set forth below.

Through our genomic sequencing studies, we have identified all nineteen exons within the *hMLH1* gene, and have mapped the intron/exon boundaries. One aspect of the invention, therefore, is the individual exons of the *hMLH1* gene. Table 1 presents the nucleotide coordinates (i.e., the point of insertion of each intron within the coding region of the gene) of the *hMLH1* exons (SEQ ID NOS: 25-43). The presented coordinates are based on the *hMLH1* cDNA sequence, assigning position "1" to the "A" of the start "ATG" (which A is nucleotide 1 in SEQ ID NO: 4.

Table 1

Intron Number	cDNA Sequence Coordinates
intron 1	116 & 117
intron 2	207 & 208
intron 3	306 & 307
intron 4	380 & 381
intron 5	453 & 454
intron 6	545 & 546
intron 7	592 & 593
intron 7	677 & 678
intron 9	790 & 791
intron 10	884 & 885
intron 11	1038 & 1039
intron 12	1409 & 1410
intron 13	1558 & 1559
intron 14	1667 & 1668
intron 15	1731 & 1732
intron 16	1896 & 1897
intron 17	1989 & 1990
intron 18	2103 & 2104

We have also determined the nucleotide sequence of intron regions which flank exons of the *hMLH1* gene. SEQ ID NOS: 6-24 are individual exon sequences bounded by their respective upstream and downstream intron

sequences. The same nucleotide structures are shown in Fig. 4A, where the exons are numbered from N-terminus to C-terminus with respect to the chromosomal locus. The 5-digit numbers indicate the primers used to amplify the exon. All sequences are numbered assuming the A of the ATG codon is nucleotide 1. The numbers in () are the nucleotide coordinates of the coding sequence found in the indicated exon. Uppercase is intron. Lowercase is exon or non-translated sequences found in the mRNA/cDNA clone. Lowercase and underlined sequences correspond to primers. The stop codon at 2269-2271 is in italics and underlined.

Table 2 presents the sequences of primer pairs ("first stage" primers) which we have used to amplify individual exons together with flanking intron

structures.

Table 2

Table 2					
EXON	PRIMER	PRIMER	PRIMER	PRIMER NUCLEOTIDE	
NO.	LOCATION	NO.	SEQ ID NO	SEQUENCE .	
1	upstream	18442	44	5'aggcactgaggtgattggc	
1	downstream	19109	45	5'tegtagecettaagtgage	
2	upstream	19689	46	5'aatatgtacattagagtagttg	
2	downstream	19688	47	5'cagagaaaggtcctgactc	
3	upstream	19687	48	5'agagatttggaaaatgagtaac	
3	downstream	19786	49	5'acaatgtcatcacaggagg	
4	upstream	18492	50	5'aacctttccctttggtgagg	
4	downstream	18421	51	5'gattactetgagacctagge	
5	upstream	18313	52	5'gattitetetttteeeettggg	
5	downstream	18179	53	5'caaacaaagettcaacaatttac	

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EXON NO.	PRIMER LOCATION	PRIMER NO.	PRIMER SEQ ID NO	PRIMER NUCLEOTIDE SEQUENCE
6	upstream	18318	54	5'gggttttattttcaagtacttctatg
6	downstream	18317	55	5'gctcagcaactgttcaatgtatgagc
7	upstream	19009	56	5'ctagtgtgtgtttttggc
7	downstream	19135	57	5'cataacettatetecace
8	upstream	18197	58	5'ctcagccatgagacaataaatcc
8	downstream	18924	59	5'ggttcccaaataatgtgatgg
9	upstream	18765	60	5'caaaagettcagaatete
9	downstream	18198	61	5'ctgtgggtgtttcctgtgagtgg
10	upstream	18305	62	5'catgactttgtgtgaatgtacacc
10	downstream	18306	63	5'gaggagagcctgatagaacatctg
11	upstream	18182	64	5'gggettttteteeeeeteee
11	downstream	19041	65	5'aaaatetgggeteteaeg
12	upstream	18579	66	5'aattatacctcatactagc
12	downstream	18178	67	5'gttttattacagaataaaggagg
12	downstream	19070	68	5'aagccaaagttagaaggca
13	upstream	18420	69	5'tgcaacccacaaaatttggc
13	downstream	18443	70	5'ctttctccatttccaaaacc
14	upstream	19028	71	5'tggtgtctctagttctgg
14	downstream	18897	72	5'cattgttgtagtagctctgc
15	upstream	19025	73	5'cccatttgtcccaactgg

EXON NO.	PRIMER LOCATION	PRIMER NO.	PRIMER SEQ ID NO	PRIMER NUCLEOTIDE SEQUENCE
15	downstream	18575	74	5'cggtcagttgaaatgtcag
16	upstream	18184	75	5'cattiggatgctccgttaaagc
16	downstream	18314	76	5'cacccggctggaaattttatttg
17	upstream	18429	77	5'ggaaaggcactggagaaatggg
17	downstream	18315	78	5'ccctccagcacacatgcatgtaccg
18	upstream	18444	79	5'taagtagtctgtgatctccg
18	downstream	18581	80	5'atgtatgaggtcctgtcc
19	upstream	18638	81	5'gacaccagtgtatgttgg
19	downstream	18637	82	5'gagaaagaagaacacatccc

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Additionally, we have designed a set of "second stage" amplification primers, the structures of which are shown below in Table 3. We use the second stage primers in conjunction with the first stage primers in a nested amplification protocol, as described below.

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Table 3

EXON	PRIMER	PRIMER	PRIMER	PRIMER
NO.	LOCATION	NO.	SEQ ID	NUCLEOTIDE SEQUENCE
			110	SEQUENCE
1	upstream	19295	83	5'tgtaaaacgacggccagtcact gaggtgattggctgaa
1	downstream	19446	84	*5'tagcccttaagtgagcccg
2	upstream	18685	85	5'tgtaaaacgacggccagttacat tagagtagttgcaga

EXON	PRIMER	PRIMER	PRIMER	PRIMER
NO.	LOCATION	NO.	SEQ ID	NUCLEOTIDE
			NO	SEQUENCE
2	downstream	19067	86	*5'aggtcetgactcttccatg
3	upstream	18687	87	5'tgtaaaacgacggccagtttgga aaatgagtaacatgatt
3	downstream	19068	88	*5'tgtcatcacaggaggatat
4	upstream	19294	89	5'tgtaaaacgacggccagtctttc cctttggtgaggtga
4	downstream	19077	90	*5'tactctgagacctaggccca
5	upstream	19301	91	5'tgtaaaacgacggccagttctct tttccccttgggattag
5	downstream	19046	92	*5'acaaagcttcaacaatttactc t
6	upstream	19711	93	5'tgtaaaacgacggccagtgtttt attttcaagtacttctatgaatt
б	downstream	19079	94	*5'cagcaactgttcaatgtatgag
7	upstream	19293	95	5'tgtaaaacgacggccagtgtgtg tgtttttggcaac
7	downstream	19435	96	*5'aaccttatetecaccage
8	upstream	19329	97	5'tgtaaaacgacggccagtagcc atgagacaataaatccttg
8	downstream	19450	98	*5'tcccaaataatgtgatggaatg
9	upstream	19608	99	5'tgtaaaacgacggccagtaagc ttcagaatctctttt

EXON	PRIMER	PRIMER	PRIMER	PRIMER
NO.	LOCATION	NO.	SEQ ID	NUCLEOTIDE
			NO	SEQUENCE
9	downstream	19449	100	*5'tgggtgtttcctgtgagtggatt
10	upstream	19297	101	5'tgtaaaacgacggccagtacttt gtgtgaatgtacacctgtg
10	downstream	19081	102	*5'gagagcctgatagaacatctgt
11	upstream	19486	103	5'tgtaaaacgacggccagtcttttt ctccccctcccacta
11	downstream	19455	104	*5'tctgggctctcacgtct
12	upstream	20546	105	*5'cttattctgagtctctcc
12	downstream	20002	106	5'tgtaaaacgacggccagtgtttg ctcagaggctgc
12	upstream	19829	107	*5'gatggttcgtacagattcccg
12	downstream	19385	108	5'tgtaaaacgacggccagtttatt acagaataaaggaggtag
13	upstream	19300	109	5'tgtaaaacgacggccagtaacc cacaaaatttggctaag
13	downstream	19078	110	*5'tetecattteeaaaaeettg
14	upstream	19456	111	*5'tgtctctagttctggtgc
14	downstream	19472	112	5'tgtaaaacgacggccagttgttg tagtagctctgcttg
15	upstream	19697	113	*5'atttgtcccaactggttgta

	T		Γ	I
EXON	PRIMER	PRIMER	PRIMER	PRIMER
NO.	LOCATION	NO.	SEQ ID	NUCLEOTIDE
			NO	SEQUENCE
15	downstream	19466	114	5'tgtaaaacgacggccagttcagt tgaaatgtcagaaagtg
16	upstream	19269	115	5'tgtaaaacgacggccagt
16	downstream	19047	116	*5'ccggctggaaattttatttggag
17	upstream	19298	117	5'tgtaaaacgacggccagtaggc actggagaaatgggatttg
17	downstream	19080	118	*5'tccagcacacatgcatgtaccg aaat
18	upstream	19436	119	*5'gtagtctgtgatctccgttt
18	downstream	19471	120	5'tgtaaaacgacggccagttatga ggtcctgtcctag
19	upstream	19447	121	*5'accagtgtatgttgggatg
19	· downstream -	19330	122	5'tgtaaaacgacggccagtgaaa gaagaacacatcccaca

In Table 3 an asteric (*) indicates that the 5' nucleotide is biotinylated. Exons 1-7, 10, 13 and 16-19 can be specifically amplified in PCR reactions containing either 1.5 mM or 3 mM MgCl₂. Exons 11 and 14 can only be specifically amplified in PCR reactions containing 1.5 mM MgCl₂ and exons 8, 9, 12 and 15 can only be specifically amplified in PCR reactions containing 3 mM MgCl₂. With respect to exon 12, the second stage amplification primers have been designed so that exon 12 is reamplified in two halves. The 20546 and 20002 primer set amplifies the N-terminal half. The primer set 19829 and 19835 amplifies the C-terminal half. An alternate primer for 18178 is 19070.

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The hMLH1 sequence information provided by our studies and disclosed in this application and preceding related applications, may be used to design a large number of different oligonucleotide primers for use in identifying hMLH1 mutations that correlate with cancer susceptibility and/or with tumor development in an individual, including primers that will amplify more than one exon (and/or flanking intron sequences) in a single product band.

One of ordinary skill in the art would be familiar with considerations important to the design of PCR primers for use to amplify the desired fragment or gene.³⁷ These considerations may be similar, though not necessarily identical to those involved in design of sequencing primers, as discussed above. Generally it is important that primers hybridize relatively specifically (i.e. have a T_m of greater than about 55-degrees° C, and preferably around 60-degrees° C). For most cases, primers between about 17 and 25 nucleotides in length work well. Longer primers can be useful for amplifying longer fragments. In all cases, it is desirable to avoid using primers that are complementary to more than one sequence in the human genome, so that each pair of PCR primers amplifies only a single, correct fragment. Nevertheless, it is only absolutely necessary that the correct band be distinguishable from other product bands in the PCR reaction.

The exact PCR conditions (e.g. salt concentration, number of cycles, type of DNA polymerase, etc.) can be varied as known in the art to improve, for example, yield or specificity of the reaction. In particular, we have found it valuable to use nested primers in PCR reactions in order to reduce the amount of required DNA substrate and to improve amplification specificity.

Two examples follow. The first example illustrates use of a first stage primer pair (SEQ ID NOS: 69 and 70) to amplify intron/exon segment (SEQ ID NO: 18). The second example illustrates use of second stage primers to amplify a target intron/exon segment from the product of a first PCR amplification step employing first stage primers.

EXAMPLE 1: Amplification of hMLHI genomic clones from a P1 phage library

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25ng genomic DNA (or 1ng of a P1 phage can be used) was used in PCR reactions including:

0.05mM dNTPs

50mM KCl

3mM Mg

10mM Tris-HCl pH 8.5

0.01% gelatin

5µM primers

Reactions were performed on a Perkin-Elmer Cetus model 9600 thermal cycler. Reactions were incubated at 95-degrees° C for 5 minutes, followed by 35 cycles (30 cycles from a P1 phage) of:

94-degrees° C for 30 seconds

55-degrees° C for 30 seconds

72-degrees° C for 1 minute.

A final 7 minute extension reaction was then performed at 72°-degrees C. Desirable P1 clones were those from which an approximately bp product band was produced.

EXAMPLE 2: Amplification of *hMLH1* sequences from genomic DNA using nested PCR primers

We performed two-step PCR amplification of *hMLH1* sequences from genomic DNA as follows. Typically, the first amplification was performed in a 25 microliter reaction including:

25ng of chromosomal DNA

Perkin-Elmer PCR buffer II (any suitable buffer could be used)

3mM MgCl₂

50µM each dNTP

Taq DNA polymerase

5μM primers (SEQ ID NOS: 69, 70)

and incubated at 95-degrees° C for 5 minutes, followed by 20 cycles of:

94-degrees° C for 30 seconds

55-degrees° C for 30 seconds.

The product band was typically small enough (less than an approximately 500 bp) that separate extension steps were not performed as part of each cycle. Rather, a single extension step was performed, at 72-degrees °C for 7 minutes, after the 20 cycles were completed. Reaction products were stored at 4-degrees °C.

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The second amplification reaction, usually 25 or 50 microliters in volume, included:

1 or 2 microliters (depending on the volume of the reaction) of the first amplification reaction product

Perkin-Elmer PCR buffer II (any suitable buffer could be used)

3mM or MgCl₂

50 μM each dNTP

Taq DNA polymerase

 $5\mu M$ nested primers (SEQ ID NOS: 109, 110),

and was incubated at 95-degrees° C for 5 minutes, followed by 20-25 cycles of:

94-degrees° C for 30 seconds

55-degrees° C for 30 seconds

a single extension step was performed, at 72-degrees° C for 7 minutes, after the cycles were completed. Reaction products were stored at 4-degrees° C.

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Any set of primers capable of amplifying a target hMLH1 sequence can be used in the first amplification reaction. We have used each of the primer sets presented in Table 2 to amplify an individual hMLH1 exon in the first amplification reaction. We have also used combinations of those primer sets, thereby amplifying multiple individual hMLH1 exons in the first amplification reaction.

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The nested primers used in the first amplification step were designed relative to the primers used in the first amplification reaction. That is, where a single set of primers is used in the first amplification reaction, the primers used in the second amplification reaction should be identical to the primers used in the first reaction except that the primers used in the second reaction should not include the 5'-most nucleotides of the first amplification reaction primers, and should extend sufficiently more at the 3' end that the T_m of the second amplification primers is approximately the same as the T_m of the first

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amplification reaction primers. Our second reaction primers typically lacked the 3 5'-most nucleotides of the first amplification reaction primers, and extended approximately 3-6 nucleotides farther on the 3' end. SEQ ID NOS: 109, 110 are examples of nested primer pairs that could be used in a second amplification reaction when SEQ ID NOS: 69 and 70 were used in the first amplification reaction.

We have also found that it can be valuable to include a standard sequence at the 5' end of one of the second amplification reaction primers to prime sequencing reactions. Additionally, we have found it useful to biotinylate that last nucleotide of one or both of the second amplification reaction primers so that the product band can easily be purified using magnetic beads⁴⁰ and then sequencing reactions can be performed directly on the bead-associated products.⁴¹⁻⁴⁵

For additional discussion of multiplex amplification and sequencing methods, see References by Zu et al. and Espelund et al.^{46, 47}

hMLH1 Link to Cancer

As a first step to determine whether *hMLH1* was a candidate for the HNPCC locus on human chromosome 3p21-23,³ we mapped *hMLH1* by fluorescence *in situ* hybridization (FISH).^{20,21} We used two separate genomic fragments (data not shown) of the *hMLH1* gene in FISH analysis. Examination of several metaphase chromosome spreads localized *hMLH1* to chromosome 3p21.3-23.

Panel A of Figure 7 shows hybridization of hMLH1 probes in a metaphase spread. Biotinylated hMLH1 genomic probes were hybridized to banded human metaphase chromosomes as previously described.^{20,21} Detection was performed with fluorescein isothiocyanate (FITC)-conjugated avidin (green signal); chromosomes, shown in blue, were counterstained with 4'6-diamino-2-phenylindole (DAPI). Images were obtained with a cooled CCD camera, enhanced, pseudocoloured and merged with the following programs: CCD Image Capture; NIH Image 1.4; Adobe Photoshop and Genejoin Maxpix respectively. Panel B of Figure 7 shows a composite of chromosome 3 from multiple

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metaphase spreads aligned with the human chromosome 3 ideogram. Region of hybridization (distal portion of 3p21.3-23) is indicated in the ideogram by a vertical bar.

As independent confirmation of the location of *hMLH1* on chromosome 3, we used both PCR with a pair of *hMLH1*-specific oligonucleotides and Southern blotting with a *hMLH1*-specific probe to analyze DNA from the NIGMS2 rodent/human cell panel (Coriell Inst. for Med. Res., Camden, NJ, USA). Results of both techniques indicated chromosome 3 linkage. We also mapped the mouse *MLH1* gene by FISH to chromosome 9 band E. This is a position of synteny to human chromosome 3p.²² Therefore, the *hMLH1* gene localizes to 3p21.3-23, within the genomic region implicated in chromosome 3-linked HNPCC families.³

Next, we analyzed blood samples from affected and unaffected individuals from two chromosome-3 candidate HNPCC families ³ for mutations. One family, Family 1, showed significant linkage (lod score = 3.01 at recombination fraction of 0) between HNPCC and a marker on 3p. For the second family, Family 2, the reported lod score (1.02) was below the commonly accepted level of significance, and thus only suggested linkage to the same marker on 3p. Subsequent linkage analysis of Family 2 with the microsatellite marker D3S1298 on 3p21.3 gave a more significant lod score of 1.88 at a recombination fraction of 0. Initially, we screened for mutations in two PCR-amplified exons of the hMLHI gene by direct DNA sequencing (Figure 4). We examined these two exons from three affected individuals of Family 1, and did not detect any differences from the expected sequence. In Family 2, we observed that four individuals affected with colon cancer are heterozygous for a C to T substitution in an exon encoding amino acids 41-69, which corresponds to a highly-conserved region of the protein (Figure 9). For one affected individual, we screened PCRamplified cDNA for additional sequence differences. The combined sequence information obtained from the two exons and cDNA of this one affected individual represents 95% (i.e. all but the first 116 bp) of the open reading frame. We observed no nucleotide changes other than the C to T substitution. In addition, four individuals from Family 2, predicted to be carriers based upon

linkage data, and as yet unaffected with colon cancer, were found to be heterozygous for the same C to T substitution. Two of these predicted carriers are below and two are above the mean age of onset (50 years) in this particular family. Two unaffected individuals examined from this same family, both predicted by linkage data to be non carriers, showed the expected normal sequence at this position. Linkage analysis that includes the C to T substitution in Family 2 gives a lod score of 2.23 at a recombination fraction 0. Using low stringency cancer diagnostic criteria, we calculated a lod score of 2.53. These data indicate the C to T substitution shows significant linkage to the HNPCC in Family 2.

Figure 8 shows sequence chromatograms indicating a C to T transition mutation that produces a non-conservative amino acid substitution at position 44 of the hMLH1 protein. Sequence analysis of one unaffected (top panels, plus and minus strands) and one affected individual (lower panels, plus and minus strands) is presented. The position of the heterozygous nucleotide is indicated by an arrow. Analysis of the sequence chromatographs indicates that there is sufficient T signal in the C peak and enough A signal in the G peak for the affected individuals to be heterozygous at this site.

To determine whether this C to T substitution was a polymorphism, we sequenced this same exon amplified from the genomic DNA from 48 unrelated individuals and observed only the normal sequence. We have examined an additional 26 unrelated individuals using allele specific oligonucleotide (ASO) hybridization analysis.³³ The ASO sequences (SEQ ID NOS: 141 and 142, respectively) which we used are:

5'-ACTTGTGGATTTTGC-3' and

5'-ACTTGTGAATTTTGC-3'.

Based upon direct DNA sequencing and ASO analysis, none of these 74 unrelated individuals carry the C to T substitution. Therefore, the C to T substitution observed in Family 2 individuals is not likely to be a polymorphism. As mentioned above, we did not detect this same C to T substitution in affected individuals from a second chromosome 3-linked family, Family 1.³ We are continuing to study individuals of Family 1 for mutations in hMLH1.

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Table 4 below summarizes our experimental analysis of blood samples from affected and unaffected individuals from Family 2 and unrelated individuals.

Table 4

0/2

0/74

5 Number of Individuals with C to T Mutation/ Status Number of Individuals Tested F Α Affected 4/4 M 10 I Predicted Carriers 4/4 L Y

Predicted Non-carriers

15 Unrelated Individuals

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Based on several criteria, we suggest that the observed C to T substitution in the coding region of hMLH1 represents the mutation that is the basis for HNPCC in Family 2.3 First, DNA sequence and ASO analysis did not detect the C to T substitution in 74 unrelated individuals. Thus, the C to T substitution is not simply a polymorphism. Second, the observed C to T substitution is expected to produce a serine to phenylalanine change at position 44 (See Figure 9). This amino acid substitution is a non-conservative change in a conserved region of the protein (Figures 3 and 9). Secondary structure predictions using Chou-Fasman parameters suggest a helix-turn-beta sheet structure with position 44 located in the turn. The observed Ser to Phe substitution, at position 44 lowers the prediction for this turn considerably, suggesting that the predicted amino acid substitution alters the conformation of the hMLH1 protein. The suggestion that the Ser to Phe substitution is a mutation which confers cancer susceptibility is further supported by our experiments which

show that an analogous substitution (alanine to phenylalanine) in a yeast MLH1 gene results in a nonfunctional mismatch repair protein. In bacteria and yeast, a mutation affecting DNA mismatch repair causes comparable increases in the rate of spontaneous mutation including additions and deletions within dinucleotide repeats. 4.5.11.13,14.15.16 In humans, mutation of hMSH2 is the basis of chromosome-2 HNPCC, 1.2 tumors which show microsatellite instability and an apparent defect in mismatch repair. 12 Chromosome 3-linked HNPCC is also associated with instability of dinucleotide repeats. 3 Combined with these observations, the high degree of conservation between the human MLH1 protein and the yeast DNA mismatch repair protein MLH1 suggests that hMLH1 is likely to function in DNA mismatch repair. During isolation of the hMLH1 gene, we identified the hPMS1 gene. This observation suggests that mammalian DNA mismatch repair, like that in yeast, 4 may require at least two MutL-like proteins.

It should be noted that it appears that different HNPCC families show different mutations in the MLH1 gene. As explained above, affected individuals in Family 1 showed "tight linkage" between HNPCC and a locus in the region of 3p21-23. However, affected individuals in Family 1 do not have the C to T mutation found in Family 2. It appears that the affected individuals in Family 1 have a different mutation in their MLH1 gene. Further, we have used the structure information and methods described in this application to find and characterize another hMLH1 mutation which apparently confers cancer susceptibility in heterozygous carriers of the mutant gene in a large English HNPCC family. The hMLH1 mutation in the English family is a + 1 T frameshift which is predicted to lead to the synthesis of a truncated hMLH1 protein. Unlike, for example, sickle cell anemia, in which essentially all known affected individuals have the same mutation multiple hMLH1 mutations have been discovered and linked to cancer. Therefore, knowledge of the entire cDNA sequence for hMLH1 (and probably hPMS1), as well as genomic sequences particularly those that surround exons, will be useful and important for characterizing mutations in families identified as exhibiting a high frequency of cancer.

Subsequent to our discovery of a cancer conferring mutation in hMLH1, studies by others have resulted in the characterization of at least 5

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additional mutations in *hMLH1*, each of which appears to have conferred cancer susceptibility to individuals in at least one HNPCC family. For example, Papadopoulos et al. indentified such as a mutation, characterized by an in-frame deletion of 165 base pairs between codons 578 to 632. In another family, Papadopoulos et al. observed an *hMLH1* mutation, characterized by a frame shift and substitution of new amino acids, namely, a 4 base pair deletion between codons 727 and 728. Papadopoulos et al. also reports an *hMLH1* cancer linked mutation, characterized by an extension of the COOH terminus, namely, a 4 base pair insertion between codons 755 and 756.³⁸

In summary, we have shown that DNA mismatch repair gene *hMLH1* which is likely to be the hereditary nonpolyposis colon cancer gene previously localized by linkage analysis to chromosome 3p21-23.³ Availability of the *hMLH1* gene sequence will facilitate the screening of HNPCC families for cancer-linked mutations. In addition, although loss of heterozygosity (LOH) of linked markers is not a feature of either the 2p or 3p forms of HNPCC,^{3,6} LOH involving the 3p21.3-23 region has been observed in several human cancers.²⁴⁻²⁶ This suggests the possibility that *hMLH1* mutation may play some role in these tumors.

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Human PMS1

Human *PMS1* was isolated using the procedures discussed with reference to Figure 1. Figure 10 shows the entire *hPMS1* cDNA nucleotide sequence. Figure 11 shows an alignment of the predicted human and yeast PMS1 protein sequences. We determined by FISH analysis that human *PMS1* is located on chromosome 7. Subsequent to our discovery of *hPMS1*, others have identified mutations in the gene which appear to confer HNPCC susceptibility.³⁹

Mouse MLHI

Using the procedure outlined above with reference to Figure 1, we have determined a partial nucleotide sequence of mouse *MLH1* cDNA, as shown in Figure 12 (SEQ ID NO: 135). Figure 13 shows the corresponding predicted amino acid sequence for mMLH1 protein (SEQ ID NO: 136) in comparison to

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the predicted hMLH1 protein sequence (SEQ ID NO: 5). Comparison of the mouse and human MLH1 proteins as well as the comparison of hMLH1 with yeast MLH1 proteins, as shown in Figure 9, indicate a high degree of conservation.

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Mouse PMS1

Using the procedures discussed above with reference to Figure 1, we isolated and sequenced the mouse *PMS1* gene, as shown in Figure 14 (SEQ ID NO: 137). This cDNA sequence encodes a predicted protein of 864 amino acids (SEQ ID NO: 138), as shown in Figure 15, where it is compared to the predicted amino acid sequence for hPMS1 (SEQ ID NO: 133). The degree of identity between the predicted mouse and human PMS1 proteins is high, as would be expected between two mammals. Similarly, as noted above, there is a strong similarity between the human PMS1 protein and the yeast DNA mismatch repair protein PMS1, as shown in Figure 11. The fact that yeast PMS1 and MLH1 function in yeast to repair DNA mismatches, strongly suggests that human and mice PMS1 and MLH1 are also mismatch repair proteins.

Uses for Mouse MLH1 and PMS1

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We believe our isolation and characterization of *mMLH1* and *mPMS1* genes will have many research applications. For example, as already discussed above, we have used our knowledge of the *mPMS1* gene to produce antibodies which react specifically with hPMS1. We have already explained that antibodies directed to the human proteins, MLH1 or PMS1 may be used for both research purposes as well as diagnostic purposes.

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We also believe that our knowledge of mPMS1 and mMLH1 will be useful for constructing mouse models in order to study the consequences of DNA mismatch repair defects. We expect that mPMS1 or mMLH1 defective mice will be highly prone to cancer because chromosome 2p and 3p-associated HNPCC are each due to a defect in a mismatch repair gene. As noted above, we have already produced chimeric mice which carry an mPMS1 defective gene. We are currently constructing mice heterozygous for mPMS1 or mMLH1 mutation. These

heterozygous mice should provide useful animal models for studying human cancer, in particular HNPCC. The mice will be useful for analysis of both intrinsic and extrinsic factors that determine cancer risk and progression. Also, cancers associated with mismatch repair deficiency may respond differently to conventional therapy in comparison to other cancers. Such animal models will be useful for determining if differences exist, and allow the development of regimes for the effective treatment of these types of tumors. Such animal models may also be used to study the relationship between hereditary versus dietary factors in carcinogenesis.

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Distinguishing Mutations From Polymorphisms

For studies of cancer susceptibility and for tumor identification and characterization, it is important to distinguish "mutations" from "polymorphisms". A "mutation" produces a "non-wild-type allele" of a gene. A non-wild-type allele of a gene produces a transcript and/or a protein product that does not function normally within a cell. "Mutations" can be any alteration in nucleotide sequence including insertions, deletions, substitutions, and rearrangements.

"Polymorphisms", on the other hand, are sequence differences that are found within the population of normally-functioning (i.e., "wild-type") genes. Some polymorphisms result from the degeneracy of the nucleic acid code. That is, given that most amino acids are encoded by more than one triplet codon, many different nucleotide sequences can encode the same polypeptide. Other polymorphisms are simply sequence differences that do not have a significant effect on the function of the gene or encoded polypeptide. For example, polypeptides can often tolerate small insertions or deletions, or "conservative" substitutions in their amino acid sequence without significantly altering function of the polypeptide.

"Conservative" substitutions are those in which a particular amino acid is substituted by another amino acid of similar chemical characteristics. For example, the amino acids are often characterized as "non-polar (hydrophobic)" including alanine, leucine, isoleucine, valine, proline, phenylaline, tryptophan, and methionine; "polar neutral", including glycine, serine, threonine, cysteine, tyrosine,

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asparagine, and glutamine; "positively charged (basic)", including arginine, lysine, and histidine; and "negatively charged (acidic)", including aspartic acid and glutamic acid. A substitution of one amino acid for another amino acid in the same group is generally considered to be "conservative", particularly if the side groups of the two relevant amino acids are of a similar size.

The first step in identifying a mutation or polymorphism in a mismatch repair gene sequence involves identification, using available techniques including those described herein, of a mismatch repair gene, (or gene fragment) sequence that differs from a known, normal (e.g. wild-type) sequence of the same mismatch repair gene (or gene fragment). For example, a hMLH1 gene (or gene fragment) sequence could be identified that differs in at least one nucleotide position from a known normal (e.g. wild-type) hMLH1 sequence such as any of SEO ID NOS: 6-24.

Mutations can be distinguished from polymorphisms using any of a variety of methods, perhaps the most direct of which is data collection and correlation with tumor development. That is, for example, a subject might be identified whose hMLH1 gene sequence differs from a sequence reported in SEQ ID. NOS: 6-24, but who does not have cancer and has no family history of cancer. Particularly if other, preferably senior, members of that subject's family have hMLH1 gene sequences that differ from SEQ ID NOS: 6-24 in the same way(s), it is likely that subject's hMLH1 gene sequence could be categorized as a "polymorphism". If other, unrelated individuals are identified with the same hMLH1 gene sequence and no family history of cancer, the categorization may be confirmed.

Mutations that are responsible for conferring genetic susceptibility to cancer can be identified because, among other things, such mutations are likely to be present in all tissues of an affected individual and in the germ line of at least one of that individual's parents, and are not likely to be found in unrelated families with no history of cancer.

When distinguishing mutations from polymorphisms, it can sometimes be valuable to evaluate a particular sequence difference in the presence of at least one known mismatch repair gene mutation. In some

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instances, a particular sequence change will not have a detectable effect (i.e., will appear to be a polymorphism) when assayed alone, but will, for example, increase the penetrance of a known mutation, such that individuals carrying both the apparent polymorphism difference and a known mutation have higher probability of developing cancer than do individuals carrying only the mutation. Sequence differences that have such an effect are properly considered to be mutations, albeit weak ones.

As discussed above and previously (U.S. Patent Application Nos. 08/168,877 and 08/209,521), mutations in mismatch repair genes or gene products produced non-wild-type versions of those genes or gene products. Some mutations can therefore be distinguished from polymorphisms by their functional characteristics in *in vivo* or *in vitro* mismatch repair assays. Any available mismatch repair assay can be used to analyze these characteristics. ⁴⁹⁻⁶³ It is generally desirable to utilize more than one mismatch repair assay before classifying a sequence change as a polymorphism, since some mutations will have effects that will not be observed in all assays.

For example, a mismatch repair gene containing a mutation would not be expected to be able to replace an endogenous copy of the same gene in a host cell without detectably affecting mismatch repair in that cell; whereas a mismatch repair gene containing a sequence polymorphism would be expected to be able to replace an endogenous copy of the same gene in a host cell without detectably affecting mismatch repair in that cell. We note that for such "replacement" studies, it is generally desirable to introduce the gene to be tested into a host cell of the same (or at least closely related) species as the cell from which the test gene was derived, to avoid complications due to, for example, the inability of a gene product from one species to interact with other mismatch repair gene products from another species. Similarly, a mutant mismatch repair protein would not be expected to function normally in an *in vitro* mismatch repair system (preferably from a related organism); whereas a polymorphic mismatch repair protein would be expected to function normally.

The methods described herein and previously allow identification of different kinds of mismatch repair gene mutations. The following examples

illustrate protocols for distinguishing mutations from polymorphisms in DNA mismatch repair genes.

EXAMPLE 3: We have developed a system for testing in yeast, S. cerevisiae the functional significance of mutations found in either the hMLH1 or hPMS1 genes. The system is described in this application using as an example, the serine (SER) to phenylalanine (PHE) causing mutation in hMLH1 that we found in a family with HNPCC, as described above. We have derived a yeast strain that it is essentially deleted for its MLH1 gene and hence is a strong mutator (i.e., 1000 fold above the normal rate in a simple genetic marker assay involving reversion from growth dependence on a given amino acid to independence (reversion of the hom3-10 allele, Prolla, Christie and Liskay, Mol Cell Biol, 14:407-415, 1994). When we placed the normal yeast MLH1 gene (complete with all known control regions) on a yeast plasma that is stably maintained as a single copy into the MLH1-deleted strain, the mutator phenotype is fully corrected using the reversion to amino acid independence assay. However, if we introduce a deleted copy of the yeast MLH1 there is no correction. We next tested the mutation that in the HNPCC family caused a SER to PHE alteration. We found that the resultant mutant yeast protein cannot correct the mutator phenotype, strongly suggesting that the alteration from the wild-type gene sequence probably confers cancer susceptibility, and is therefore classified as a mutation, not a polymorphism. We subsequently tested proteins engineered to contain other amino acids at the "serene" position and found that most changes result in a fully mutant, or at least partially mutant phenotype.

As other "point" mutations in *MLH1* and *PMS1* genes are found in cancer families, they can be engineered into the appropriate yeast homolog gene and their consequence on protein function studied. In addition, we have identified a number of highly conserved amino acids in both the *MLH1* and *PMS1* genes. We also have evidence that *hMLH1* interacts with yeast *PMS1*. This finding raises the possibility that mutations observed in the *hMLH1* gene can be more directly tested in the yeast system. We plan to systematically make mutations that will alter the amino acid at these conserved positions and determine what amino acid substitutions are tolerated and which are not. By

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collecting mutation information relating to *hMLH1* and *hPMS1*, both by determining and documenting actual found mutations in HNPCC families, and by artificially synthesizing mutants for testing in experimental systems, it may be eventually possible to practice a cancer susceptibility testing protocol which, once the individuals *hMLH1* or *hPMS1* structure is determined, only requires comparison of that structure to known mutation versus polymorphism data.

EXAMPLE 4: Another method which we have employed to study physical interactions between hMLH1 and hPMS1, can also be used to study whether a particular alteration in a gene product results in a change in the degree of protein-protein interaction. Information concerning changes in protein-protein interaction may demonstrate or confirm whether a particular genomic variation is a mutation or a polymorphism. Following our labs findings on the interaction between yeast MLH1 and PMS1 proteins in vitro and in vivo, (U.S. Patent Application Serial No. 08/168,877), the interaction between the human counterparts of these two DNA mismatch repair proteins was tested. The human MLH1 and human PMS1 proteins were tested for in vitro interaction using maltose binding protein (MBP) affinity chromatography. hMLH1 protein was prepared as an MBP fusion protein, immobilized on an amylose resin column via the MBP, and tested for binding to hPMS1, synthesized in vitro. The hPMS1 protein bound to the MBP-hMLH1 matrix, whereas control proteins showed no affinity for the matrix. When the hMLH1 protein, translated in vitro, was passed over an MBP-hPMS1 fusion protein matrix, the hMLH1 protein bound to the MBP-hPMS1 matrix, whereas control proteins did not.

Potential *in vivo* interactions between hMLH1 and hPMS1 were tested using the yeast "two hybrid" system.²⁸ Our initial results indicate that hMLH1 and hPMS1 interact *in vivo* in yeast. The same system can also be used to detect changes in protein-protein interaction which result from changes in gene or gene product structure and which have yet to be classified as either a polymorphism or a mutation which confers cancer susceptibility.

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Detection of HNPCC Families and Their Mutation(s)

It has been estimated that approximately 1,000,000 individuals in the United States carry (are heterozygous for) an HNPCC mutant gene.²⁹ Furthermore, estimates suggest that 50-60% of HNPCC families segregate mutations in the MSH2 gene that resides on chromosome 2p.^{1,2} Another significant fraction appear to be associated with the HNPCC gene that maps to chromosome 3p21-22, presumably due to hMLH1 mutations such as the C to T transition discussed above. Identification of families that segregate mutant alleles of either the hMSH2 or hMLH1 gene, and the determination of which individuals in these families actually have the mutation will be of great utility in the early intervention into the disease. Such early intervention will likely include early detection through screening and aggressive follow-up treatment of affected individuals. In addition, determination of the genetic basis for both familial and sporadic tumors could direct the method of therapy in the primary tumor, or in recurrences.

Initially, HNPCC candidate families will be diagnosed partly through the study of family histories, most likely at the local level, e.g., by hospital oncologists. One criterion for HNPCC is the observation of microsatellite instability in individual's tumo 3.36 The presenting patient would be tested for mutations in hMSH2, hM. II, hPMSI and other genes involved in DNA mismatch repair as they are identified. This is most easily done by sampling blood from the individual. Also highly useful would be freshly frozen tumor tissue. It is important to note for the screening procedure, that affected individuals are heterozygous for the offending mutation in their normal tissues.

The available tissues, e.g., blood and tumor, are worked up for PCR-based mutation analysis using one or both of the following procedures:

 Linkage analysis with a microsatellite marker tightly linked to the hMLH1 gene.

One approach to identify cancer prone families with a hMLH1 mutation is to perform linkage analysis with a highly polymorphic marker located within or tightly linked to hMLH1. Microsatellites are highly polymorphic and therefore are very useful as markers in linkage analysis. Because we possess the

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hMLH1 gene on a single large genomic fragment in a P1 phage clone (-100kbp), it is very likely that one or more microsatellites, e.g., tracts of dinucleotide repeats, exist within, or very close to, the hMLH1 gene. At least one such microsatellite has been reported.³⁸ Once such markers have been identified, PCR primers will be designed to amplify the stretches of DNA containing the microsatellites. DNA of affected and unaffected individuals from a family with a high frequency of cancer will be screened to determine the segregation of the MLH1 markers and the presence of cancer. The resulting data can be used to calculate a lod score and hence determine the likelihood of linkage between hMLH1 and the occurrence of cancer. Once linkage is established in a given family, the same polymorphic marker can be used to test other members of the kindred for the likelihood of their carrying the hMLH1 mutation.

2) Sequencing of reverse transcribed cDNA.

a) RNA from affected individuals, unaffected and unrelated individuals is reverse transcribed (RT'd), followed by PCR to amplify the cDNA in 4-5 overlapping portions. It should be noted that for the purposes of PCR, many different oligonucleotide primer pair sequences may potentially be used to amplify relevant portions of an individual's hMLH1 or hPMS1 gene for genetic screening purposes. With the knowledge of the cDNA structures for the genes, it is a straight-forward exercise to construct primer pairs which are likely to be effective for specifically amplifying selected portions of the gene. While primer sequences are typically between 20 to 30 bases long, it may be possible to use shorter primers, potentially as small as approximately 13 bases, to amplify specifically selected gene segments. The principal limitation on how small a primer sequence may be is that it must be long enough to hybridize specifically to the targeted gene segment. Specificity of PCR is generally improved by lengthening primers and/or employing nested pairs of primers.

The PCR products, in total representing the entire cDNA, are then sequenced and compared to known wild-type sequences. In most cases a mutation will be observed in the affected individual. Ideally, the nature of mutation will indicate that it is likely to inactivate the gene product. Otherwise,

the possibility that the alteration is not simply a polymorphism must be determined.

b) Certain mutations, e.g., those affecting splicing or resulting in translation stop codons, can destabilize the messenger RNA produced from the mutant gene and hence comprise the normal RT-based mutation detection method. One recently reported technique can circumvent this problem by testing whether the mutant cDNA can direct the synthesis of normal length protein in a coupled *in vitro* transcription/translation system.³²

3) Direct sequencing of genomic DNA.

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A second route to detect mutations relies on examining the exons and the intron/exon boundaries by PCR cycle sequencing directly off a DNA template. This method requires the use of oligonucleotide pairs, such as those described in Tables 2 and 3 above, that amplify individual exons for direct PCR cycle sequencing. The method depends upon genomic DNA sequence information at each intron/exon boundary (50bp, or greater, for each boundary). The advantage of the technique is two fold. First, because DNA is more stable than RNA, the condition of the material used for PCR is not as important as it is for RNA-based protocols. Second, most any mutation within the actual transcribed region of the gene, including those in an intron affecting splicing, will be detectable.

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For each candidate gene, mutation detection may require knowledge of both the entire cDNA structure, and all intron/exon boundaries of the genomic structure. With such information, the type of causal mutation in a particular family can be determined. In turn, a more specific and efficient mutation detection scheme can be adapted for the particular family. Screening for the disease (HNPCC) is complex because it has a genetically heterogeneous basis in the sense that more than one gene is involved, and for each gene, multiple types of mutations are involved.² Any given family is highly likely to segregate one particular mutation. However, as the nature of the mutation in multiple families is determined, the spectrum of the most prevalent mutations in the population will be determined. In general, determination of the most frequent mutations will direct and streamline mutation detection.

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Because HNPCC is so prevalent in the human population, carrier detection at birth could become part of standardized neonatal testing. Families at risk can be identified and all members not previously tested can be tested. Eventually, all affected kindreds could be determined.

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Mode of Mutation Screening and Testing

DNA-based Testing

Initial testing, including identifying likely HNPCC families by standard diagnosis and family history study, will likely be done in local and smaller DNA diagnosis laboratories. However, large scale testing of multiple family members, and certainly population wide testing, will ultimately require large efficient centralized commercial facilities.

Tests will be developed based on the determination of the most

common mutations for the major genes underlying HNPCC, including at least the hMSH2 gene on chromosome 2p and the MLH1 gene on chromosome 3p. A variety of tests are likely to be developed. For example, one possibility is a set of tests employing oligonucleotide hybridizations that distinguish the normal vs. mutant alleles.³³ As already noted, our knowledge of the nucleotide structures for hMLH1, hPMS1 and hMSH2 genes makes possible the design of numerous oligonucleotide primer pairs which may be used to amplify specific portions of an individual's mismatch repair gene for genetic screening and cancer risk analysis. Our knowledge of the genes' structures also makes possible the design of labeled probes which can be quickly used to determine the presence or absence of all or a portion of one of the DNA mismatch repair genes. For example, allele-specific oligomer probes (ASO) may be designed to distinguish between alleles. ASOs are short DNA segments that are identical in sequence except for a single base difference that reflects the difference between normal and mutant alleles. Under the appropriate DNA hybridization conditions, these probes can recognize a

single base difference between two otherwise identical DNA sequences. Probes

can be labeled radioactively or with a variety of non-radioactive reporter molecules, for example, fluorescent or chemiluminescent moieties. Labeled probes are then used to analyze the PCR sample for the presence of the disease-

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causing allele. The presence or absence of several different disease-causing genes can readily be determined in a single sample. The length of the probe must be long enough to avoid non-specific binding to nucleotide sequences other than the target. All tests will depend ultimately on accurate and complete structural information relating to hMLH1, hMSH2, hPMS1 and other DNA mismatch repair genes implicated in HNPCC.

Protein Detection-Based Screening

Tests based on the functionality of the protein product, *per se*, may also be used. The protein-examining tests will most likely utilize antibody reagents specific to either the hMLH1, hPMS1 and hMSH2 proteins or other related "cancer" gene products as they are identified.

For example, a frozen tumor specimen can be cross-sectioned and prepared for antibody staining using indirect fluorescence techniques. Certain gene mutations are expected to alter or destabilize the protein structure sufficiently such as to give an altered or reduced signal after antibody staining. It is likely that such tests will be performed in cases where gene involvement in a family's cancer has yet to be established. We are in the process of developing diagnostic monoclonal antibodies against the human MLH1 and PMS1 proteins. We are overexpressing MLH1 and PMS1 human proteins in bacteria. We will purify the proteins, inject them into mice and derive protein specific monoclonal antibodies which can be used for diagnostic and research purposes.

Identification and Characterization of DNA Mismatch Repair Tumors

In addition to their usefulness in diagnosing cancer susceptibility in a subject, nucleotide sequences that are homologous to a bacterial mismatch repair gene can be valuable for, among other things, use in the identification and characterization of mismatch-repair-defective tumors. Such identification and characterization is valuable because mismatch-repair-defective tumors may respond better to particular therapy regimens. For example, mismatch-repair-defective tumors might be sensitive to DNA damaging agents, especially when administered in combination with other therapeutic agents.

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Defects in mismatch repair genes need not be present throughout an individual's tissues to contribute to tumor formation in that individual. Spontaneous mutation of a mismatch repair gene in a particular cell or tissue can contribute to tumor formation in that tissue. In fact, at least in some cases, a single mutation in a mismatch repair gene is not sufficient for tumor development. In such instances, an individual with a single mutation in a mismatch repair gene is susceptible to cancer, but will not develop a tumor until a secondary mutation occurs. Additionally, in some instances, the same mismatch repair gene mutation that is strictly tumor-associated in an individual will be responsible for conferring cancer susceptibility in a family with a hereditary predisposition to cancer development.

In yet another aspect of the invention, the sequence information we have provided can be used with methods known in the art to analyze tumors (or tumor cell lines) and to identify tumor-associated mutations in mismatch repair genes. Preferably, it is possible to demonstrate that these tumor-associated mutations are not present in non-tumor tissues from the same individual. The information described in this application is particularly useful for the identification of mismatch repair gene mutations within tumors (or tumor cell lines) that display genomic instability of short repeated DNA elements.

The sequence information and testing protocols of the present invention can also be used to determine whether two tumors are related, i.e., whether a second tumor is the result of metastasis from an earlier found first tumor which exhibits a particular DNA mismatch repair gene mutation.

Isolating Additional Genes of Related Function

Proteins that interact physically with either hMLH1 and/or hPMS1, are likely to be involved in DNA mismatch repair. By analogy to hMLH1 and hMSH2, mutations in the genes which encode for such proteins would be strong candidates for potential cancer linkage. A powerful molecular genetic approach using yeast, referred to as a "two-hybrid system", allows the relatively rapid detection and isolation of genes encoding proteins that interact with a gene product of interest, e.g., hMLH1.²⁸

The two-hybrid system involves two plasmid vectors each intended to encode a fusion protein. Each of the two vectors contains a portion, or domain, of a transcription activator. The yeast cell used in the detection scheme contains a "reporter" gene. The activator alone cannot activate transcription. However, if the two domains are brought into close proximity then transcription may occur. The cDNA for the protein of interest, e.g., hMLH1 is inserted within a reading frame in one of the vectors. This is termed the "bait". A library of human cDNAs, inserted into a second plasmid vector so as to make fusions with the other domain of the transcriptional activator, is introduced into the yeast cells harboring the "bait" vector. If a particular yeast cell receives a library member that contains a human cDNA encoding a protein that interacts with hMLH1 protein, this interaction will bring the two domains of the transcriptional activator into close proximity, activate transcription of the reporter gene and the yeast cell will turn blue. Next, the insert is sequenced to determine whether it is related to any sequence in the data base. The same procedure can be used to identify yeast proteins in DNA mismatch repair or a related process. Performing the yeast and human "hunts" in parallel has certain advantages. The function of novel yeast homologs can be quickly determined in yeast by gene disruption and subsequent examination of the genetic consequences of being defective in the new found gene. These yeast studies will help guide the analysis of novel human "hMLH1-or hPMS1-interacting" proteins in much the same way that the yeast studies on PMS1 and MLH1 have influenced our studies of the human MLH1 and PMS1 genes.

Production of Antibodies

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By using our knowledge of the DNA sequences for hMLH1 and hPMS1, we can synthesize all or portions of the predicted protein structures for the purpose of producing antibodies. One important use for antibodies directed to hMLH1 and hPMS1 proteins will be for capturing other proteins which may be involved in DNA mismatch repair. For example, by employing coimmuno-precipitation techniques, antibodies directed to either hMLH1 or hPMS1 may be precipitated along with other associated proteins which are functionally and/or physically related. Another important use for antibodies will be for the purpose

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of isolating hMLH1 and hPMS1 proteins from tumor tissue. The hMLH1 and hPMS1 proteins from tumors can then be characterized for the purpose of determining appropriate treatment strategies.

We are in the process of developing monoclonal antibodies directed to the hMLH1 and hPMS1 proteins.

EXAMPLE 5: We have also used the following procedure to produce polyclonal antibodies directed to the human and mouse forms of PMS1 protein.

We inserted a 3' fragment of the mouse PMSI cDNA in the bacterial expression plasmid vector, pET (Novagen, Madison, WI). The expected expressed portion of the mouse PMS1 protein corresponds to a region of approximately 200 amino acids at the end of the PMS1 protein. This portion of the mPMS1 is conserved with yeast PMS1 but is not conserved with either the human or the mouse MLH1 proteins. One reason that we selected this portion of the PMS1 protein for producing antibodies is that we did not want the resulting antibodies to cross-react with MLH1. The mouse PMS1 protein fragment was highly expressed in E. coli., purified from a polyacrylamide gel and the eluted protein was then prepared for animal injections. Approximately 2 mg of the PMS1 protein fragment was sent to the Pocono Rabbit Farm (PA) for injections into rabbits. Sera from rabbits multiple times was tittered against the PMS1 antigen using standard ELISA techniques. Rabbit antibodies specific to mouse PMS1 protein were affinity-purified using columns containing immobilized mouse PMS1 protein. The affinity-purified polyclonal antibody preparation was tested further using Western blotting and dot blotting. We found that the polyclonal antibodies recognized, not only the mouse PMS1 protein, but also the human PMS1 protein which is very similar. Based upon the Western blots, there is no indication that other proteins were recognized strongly by our antibody, including either the human or mouse MLH1 proteins.

DNA Mismatch Repair Defective Mice

EXAMPLE 6: In order to create a experimental model system for studying DNA mismatch repair defects and resultant cancer in a whole animal

system we have derived DNA mismatch repair defective mice using embryonic stem (ES) cell technology. Using genomic DNA containing a portion of the *mPMS1* gene we constructed a vector that upon homologous recombination causes disruption of the chromosomal *mPMS1* gene. Mouse ES cells from the 129 mouse strain were confirmed to contain a disrupted *mPMS1* allele. The ES cells were injected into C57/BL6 host blastocysts to produce animals that were chimeric or a mixture of 129 and C57/BL6 cells. The incorporation of the ES cells was determined by the presence of patches of agouti coat coloring (indicative of ES cell contribution). All male chimeras were bred with C57/BL6 female mice.

Subsequently, twelve offspring (F_2) were born in which the agoutic coat color was detected indicating the germline transmission of genetic material from the ES cells. Analysis of DNA extracted from the tail tips of the twelve offspring indicated that six of the animals were heterozygous (contained one wild-type and one mutant allele) for the mPMSI mutation. Of the six heterozygous animals, three were female, (animals F_2 -8, F_2 -11 and F_2 -12) and three were males (F_2 , F_2 -10 and F_2 -13). Four breeding pens were set up to obtain mice that were homozygous for mPMSI mutation, and additional heterozygous mice. Breeding pen #1 which contained animals F_2 -11 and F_2 -10, yielded a total of thirteen mice in three litters, four of which have been genotyped. Breeding pen #2 (animals F_2 -8 and F_2 -13) gave twenty-two animals and three litters, three of which have been genotyped. Of the seven animals genotyped, three homozygous female animals have been identified. One animal died at six weeks of age from unknown causes. The remaining homozygous females are alive and healthy at twelve weeks of age. The results indicate that mPMSI homozygous defective mice are viable.

Breeding pens #3 and #4 were used to backcross the mPMSI mutation into the C57/BL6 background. Breeding pen #3 (animal F_2 -12 crossed to a C57/BL6 mouse) produced twenty-one animals in two litters, nine of which have been genotyped. Breeding pen #4 (animal F_2 -6 crossed with a C57/BL6 mouse) gave eight mice. In addition, the original male chimera (breeding pen #5) has produced thirty-one additional offspring.

To genotype the animals, a series of PCR primers have been developed that are used to identify mutant and wild-type mPMSI genes. They

are: (SEQ ID NOS: 143-148, respectively)

Primer 1: 5'TTCGGTGACAGATTTGTAAATG-3'

Primer 2: 5'TTTACGGAGCCCTGGC-3'

Primer 3: 5'TCACCATAAAAATAGTTTCCCG-3'

Primer 4: 5'TCCTGGATCATATTTTCTGAGC-3'

Primer 5: 5'TTTCAGGTATGTCCTGTTACCC-3'

Primer 6: 5'TGAGGCAGCTTTTAAGAAACTC-3'

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Primers 1+2 (5'targeted)

Primers 1+3 (5'untargeted)

Primers 4+5 (3'targeted)

Primers 4+6 (3'untargeted)

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The mice we have developed provide an animal model system for studying the consequences of defects in DNA mismatch repair and resultant HNPCC. The long term survival of mice homozygous and heterozygous for the mPMS1 mutation and the types and timing of tumors in these mice will be determined. The mice will be screened daily for any indication of cancer onset as indicated by a hunched appearance ir. combination with deterioration in coat condition. These mice carrying mPMS1 mutation will be used to test the effects of other factors, environmental and genetic, on tumor formation. For example, the effect of diet on colon and other type of tumors can be compared for normal mice versus those carrying mPMS1 mutation either in the heterozygous or homozygous genotype. In addition, the mPMS1 mutation can be put into different genetic backgrounds to learn about interactions between genes of the mismatch repair pathway and other genes involved in human cancer, for example, p53. Mice carrying mPMS1 mutations will also be useful for testing the efficacy of somatic gene therapy on the cancers that arise in mice, for example, the expected colon cancers. Further, isogenic fibroblast cell lines from the homozygous and heterozygous mPMS1 mice can be established for use in various cellular studies, including the determination of spontaneous mutation rates.

We are currently constructing a vector for disrupting the mouse *mMLH1* gene to derive mice carrying mutation in *mMLH1*. We will compare mice carrying defects in *mPMS1* to mice carrying defects in *mMLH1*. In addition, we will construct mice that carry mutations in both genes to see whether there is a synergistic effect of having mutations in two HNPCC genes. Other studies on the *mMLH1* mutant mice will be as described above for the *mPMS1* mutant mice.

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SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT: Liskay, Robert M.

Bronner, C. Eric

Baker, Sean M.

Bollag, Roni J.

Kolodner, Richard D.

- (ii) TITLE OF INVENTION: COMPOSITIONS AND METHODS RELATING TO DNA MISMATCH REPAIR GENES
 - (iii) NUMBER OF SEQUENCES: 148
 - (iv) CORRESPONDENCE ADDRESS:
- (A) ADDRESSEE: Kolisch, Hartwell, Dickinson, McCormack & Heuser
 - (B) STREET: 520 S.W. Yamhill Street, Suite 200
 - (C) CITY: Portland
 - (D) STATE: Oregon
 - (E) COUNTRY: U.S.A.
 - (F) ZIP: 97204
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - · (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:

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- (A) NAME: Van Rysselberghe, Pierre C.
- (B) REGISTRATION NUMBER: 33,557
- (C) REFERENCE/DOCKET NUMBER: OHSU 306B

(ix) TELECOMMUNICATION INFORMATION:

- (A) TELEPHONE: (503) 224-6655
- (B) TELEFAX: (503) 295-6679
- (C) TELEX: 360619

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 361 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
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- Asn Ser Leu Asp Ala Gly Ala Thr Arg Val Asp Ile Asp Ile Glu Arg 35 40 45
- Gly Gly Ala Lys Leu Ile Arg Ile Arg Asp Asn Gly Cys Gly Ile Lys 50 55 60
- Lys Glu Glu Leu Ala Leu Ala Leu Ala Arg His Ala Thr Ser Lys Ile
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- Ala Ser Leu Asp Asp Leu Glu Ala Ile Ile Ser Leu Gly Phe Arg Gly 85 90 95
- Glu Ala Leu Ala Ser Ile Ser Ser Val Ser Arg Leu Thr Leu Thr Ser 100 105 110
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(2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 538 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

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His	Ile 290		Pro	Tyr	Leu	Ala 295	Asp	Val	Asn	Val	His		Thr	Lys	Gln
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(T) la sea	71-	•		_	_		_	345					350		
		355		Leu			360					365			
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385					390					395					400
ser	TTE	GTĀ	Asn	Val 405	Asp	Gln	Ser	Gln	Gln 410	Gln	Leu	Leu	Val	Pro 415	Tyr
Ile	Phe	Glu	Phe 420	Pro	Ala	Asp	Asp	Ala 425	Leu	Arg	Leu	ГЛЗ	Glu 430	Arg	Met
Pro	Leu	Leu 435	Glu	Glu	Val	Gly	Val 440	Phe	Leu	Ala		Tyr 445	Gly	Glu	Asn
	Phe 450	Ile	Leu	Arg	Glu	His 455	Pro	Ile	Trp	Met			Glu	Glu	Ile
		Glv	Tle	Tyr	Glu		Cva	Acn	Wor	T 0		T	m in	T	01
465				-1-	470	1100	Cys	изр	Mec	475	Leu	rea	Thr	гÀг	
	Sar	Tla	Tara	Lys		7	212	C1	T					_	480
• • •	oer	116	ъys	485	ığı	ALG	NIG	GIU	490	ALA	116	Met	Met	Ser 495	Cys
Lys .	Arg		Ile 500	Lys	Ala	Asn	His	Arg 505	Ile	Asp	Asp	His		Ala	Arg
in '	ر 1.هـ، ۱			G) n	Lou	5a~	C1 -		N	N	D	m'	510	-	D
		515		Gln			520			asn		Tyr 525	Asn	CAa	Pro
lis (Gly	Arg	Pro	Val	Leu	Val	His	Phe	Thr						
!	530					535									

,	THEO	CULTAIN.	TOIL .	. OIC	20%											
	(i)	SEQ	UENC	E CH	ARAC'	reri:	STIC	S :								
		(A) LE	NGTH	: 60	7 am:	ino a	acid	8							
		(B) TY	PE: a	amino	o ac	id									
		(C) ST	RANDI	EDNE	ss: 1	sing:	le								
		(D) TO	POLO	3Y: 3	linea	ar									
	(ii)	MOL	ECULI	E TY	PE: I	ONA	(gene	omic)							
	(xi)	SEQ	UENC	E DES	SCRI	PTIO	v: 51	EQ II	ONO:	;3:	*					
	Met	Phe	His	His	Ile	Glu	Asn	Leu	Leu	Ile	Glu	Thr	Glu	Lys	Arg	Сув
	1				5					10					15	
	Lys	Gln	Lys	Glu	Gln	Arg	Tyr	Ile	Pro	Val	Lys	Tyr	Leu	Phe	Ser	Met
				20					25					30		
	Thr	Gln	Ile	His	Gln	Ile	Asn	Asp	Ile	Asp	Val	His	Arg	Ile	Thr	Ser
			35					40					45			
	Gly	Gln	Val	Ile	Thr	Asp	Leu	Thr	Thr	Ala	Val	Lys	Glu	Leu	Val	Asp
		50					55					60				
	Asn	Ser	Ile	Asp	Ala	Asn	Ala	Asn	Gln	Ile	Glu	Ile	Ile	Phe	Lys	Asp
	65					70					75					80
	Tyr	Gly	Leu	Glu	Ser	Ile	Glu	Cys	Ser	Asp	Asn	Gly	Asp	Gly	Ile	Asp
					85					90					95	
	Pro	Ser	Asn	Tyr	Glu	Phe	Leu	Ala	Leu	Lys	His	Tyr	Thr	Ser	Lys	Ile
				100					105					110		
	Ala	ГÄą	Phe	Gln	yab	Val	Ala	Lys	Val	Gln	Thr	Leu	Gly	Phe	Arg	Gly
			115					120					125	•		
	Glu		Leu	Ser	Ser	Leu	CAa	Gly	Ile	Ala	raa	Leu	Ser	Val	Ile	Thr
:		130					135					140				
		Thr	Ser	Pro.	Pro		Ala	Asp	Lys	Glu		Tyr	Asp	Met	Val	
	145			_	_	150			_		155					160
	His	Ile	Thr	Ser	Lys	Thr	Thr	Thr	Ser	-	Asn	Lys	Gly	Thr		Val
	•				165	9 1	•••			170					175	ъ.
	rea	vai	ser	180	Leu	Pne	HIS	ASN		Pro	vai	Arg	GIN		GIU	₽n€
	Co.	T	mb		Lys	N	Cln.	Dho	185	T	C	T 0	mh	190	T10	C1-
	Ser	rys	195	Pne	гÃя	ALG	GIII	200	THE	råa	Cys	reu	205	Val	116	GTI
	Gly	Tree-		Tle	Ile	Agn	Δla		T10	T.ve	Dhe	Ser		Trn) an	Tle
	GLY	210	n.a	116	1.40	ASII	215	11717	*16	Ly S	riie	220	AGT	115	Nam	7.1
	Thr		T.vs	Glv	Lys	Lvs		T.eu	Tle	T.e.11	Ser		Ma+	Ara	Aan	Ser
	225		_,_	4. ,	2,0	230					235		1166	9		240
		Met	Ara	Lvs	Asn		Ser	Ser	Val	Phe		Ala	Glv	Glv	Met	
			5	-1-	245					250	027	•••	,	0.27	255	
	Glv	Glu	Leu	Glu	Val	Asp	Leu	Va1	Lev		Len	Asn	Pro	Phe		Asr
	1			260					265	₽				270	-,-	
	Ara	Met	Leu		Lys	Tvr	Thr	Asp		Pro	Asp	Phe	Lev		Leu	Ast
	3		275	1	-1-	-4-		280					285			

M	Tura	Tle	Ara	Val	Lys	Gly	Tyr	Ile	Ser	Gln	Asn	Ser	Phe	Gly	Cys	3
						205					300					
Cly	Ard	Asn	ser	Lys	Asp	Arg	Gln	Phe	Ile	Tyr	Val	Asn	Lys	arg	PEC	5
					210					さてっ						
303	Glu	TVT	Ser	Thr	Leu	Leu	Lys	Сув	Сув	Asn	Glu	Val	Tyr	LAS	Tn:	r
				225					330							
n b	Nan	Aan	va 1	Gln	Phe	Pro	Ala	Val	Phe	Leu	Asn	Leu	Glu	Lev	Pr	0
			- 40					345					200	•		
		Tou	Tle	Ago	Val	Asn	Val	Thr	Pro	Asp	Lys	Arg	Va]	. Ile	e Le	·u
							360					202	,			
	•• !	333	G) u	Ara	Ala	Val	Ile	Asp	Ile	Phe	Lys	Thr	Th	Le	1 S€	er
						375	1				360	,				
	370		. 3.00	220	- Gln	Glu	Leu	Ala	Lev	Pro	Lys	Arg	y Me	t Cy	s Se	er
		TAT	Man	nry	390					395	5				40	00
385	i			_,	330		. 1	. Arc	. I.e.	ı Lev	ı Thi	Gl	ı Va	l Ph	e As	вþ
Glr	ser Ser	Glu	Glr			l GTI	, ny		410)				41	5	
				405	5				- T							
							•••	1 110	1 (1)	y Gl	n Phe	e As	n Le	u Gl	y P	he
Asj	Ası	p Phe	E Ly	s Lys	s Met	: GI	ı va.	LVa	_ GI.	y Gr			43	0		
			42	0				42	- *	a 66	~ Δα	n Le	u Ph	e Il	e V	al
Il	e Il	e Va	1 Th	r Ar	g Ly	g Va	I AS	p As	n ry	s Se		44	.5			
		43	5			_	44	0		- Dh	a (3)	-		eu G	ln A	la
As	p Gl	n Hi	s Al	a Se	r As	p Gl	u Ly	в ту	r As	n Ph	46	.n				
	45	0.				45	5						n P	ro V	al G	lu
۷a	1 Th	r Va	1 Ph	e Ly	s Se	r Gl	n Ly	s Le	u ii	e Il	e FL				4	180
46	5			*	47	0				47		n Te	911 P	ro V	al I	?he
Le	u Se	r Va	1 II	.e As	p Gl	u L∈	eu Va	il Va	il Le	eu As	sp As	, L		4	95	
				48	35.				4:	90) G	1., G			Gly
G)	Lu Ly	s As	n G	Ly Pi	e Ly	s Le	eu Ly	ys I.	le A:	sp G	Lu G.	Lu G.	<u>.</u> u 5	10		-
			50	00				5	05 		T	C	_		eu	Phe
S	er Ai	rg Va	al L	ys L	eu Le	eu S	er L	eu P	ro T	hr S	er r	ys G E	25	-		
							5	20					23			
A	sp L	eu G	ly A	sp P	he A	sn G	lu L	eu I	le H	is L	eu .	16 1	y S			_
						5	マニ				2	40				
G	ly L	eu A	rg A	rg A	sp A	sn I	le A	rg C	ys S	er L	Às T	Te P	irg i	Jer ,	100	560
					=	EO.				5	22					
A	la M	et A	rg A	la C	ys A	rg S	er S	er 1	le M	Met 1	le G	TA I	-ys	FIO	575	
					45					570					-	
Ť	vs I	ys T	hr M	let I	hr F	rg V	/al \	/al	lis l	Asn I	Leu S	ser (÷LU	Leu Loo	v∍ñ	-1-
				00					585					334		
τ	ero 1	rp I	Asn (Cys I	ero E	lis (Gly A	Arg	Pro '	Thr	Met :	Arg	His	Leu	WEC	
•			595	-				600					605			

- (2) INFORMATION FOR SEQ ID NO:4:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2484 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

CTTGGCTCTT CTGGCGCCAA AATGTCGTTC GTGGCAGGGG TTATTCGGCG GCTGGACGAG 60 ACAGTGGTGA ACCGCATCGC GGCGGGGGAA GTTATCCAGC GGCCAGCTAA TGCTATCAAA 120 GAGATGATTG AGAACTGTTT AGATGCAAAA TCCACAAGTA TTCAAGTGAT TGTTAAAGAG 180 GGAGGCCTGA AGTTGATTCA GATCCAAGAC AATGGCACCG GGATCAGGAA AGAAGATCTG 240 GATATTGTAT GTGAAAGGTT CACTACTAGT AAACTGCAGT CCTTTGAGGA TTTAGCCAGT 300 ATTTCTACCT ATGGCTTTCG AGGTGAGGCT TTGGCCAGCA TAAGCCATGT GGCTCATGTT 360 ACTATTACAA CGAAAACAGC TGATGGAAAG TGTGCATACA GAGCAAGTTA CTCAGATGGA 420 AAACTGAAAG CCCCTCCTAA ACCATGTGCT GGCAATCAAG GGACCCAGAT CACGGTGGAG 480 GACCTTTTT ACAACATAGC CACGAGGAGA AAAGCTTTAA AAAATCCAAG TGAAGAATAT 540 GGGAAAATTT TGGAAGTTGT TGGCAGGTAT TCAGTACACA ATGCAGGCAT TAGTTTCTCA GTTAAAAAAC AAGGAGAGAC AGTAGCTGAT GTTAGGACAC TACCCAATGC CTCAACCGTG 660 GACAATATTC GCTCCATCTT TGGAAATGCT GTTAGTCGAG AACTGATAGA AATTGGATGT 720 GAGGATAAAÄ CCCTAGECTT CAAAATGAAT GGTTACATAT CCAATGCAAA CTACTCAGTG 780 AAGAAGTGCA TCTTCTTACT CTTCATCAAC CATCGTCTGG TAGAATCAAC TTCCTTGAGA AAAGCCATAG AAACAGTGTA TGCAGCCTAT TTGCCCAAAA ACACACACCC ATTCCTGTAC CTGAGTTTAG AAATCAGTCC CCAGAATGTG GATGTTAATG TGCACCCCAC AAAGCATGAA GTTCACTTCC TGCACGAGGA GAGCATCCTG GAGCGGGTGC AGCAGCACAT CGAGAGCAAG 1020 CTCCTGGGCT CCAATTCCTC CAGGATGTAC TTCACCCAGA CTTTGCTACC AGGACTTGCT 1080 GGCCCCTCTG GGGAGATGGT TAAATCCACA ACAAGTCTGA CCTCGTCTTC TACTTCTGGA 1140 AGTAGTGATA AGGTCTATGC CCACCAGATG GTTCGTACAG ATTCCCGGGA ACAGAAGCTT 1200 GATGCATTTC TGCAGCCTCT GAGCAAACCC CTGTCCAGTC AGCCCCAGGC CATTGTCACA 1260 GAGGATAAGA CAGATATTTC TAGTGGCAGG GCTAGGCAGC AAGATGAGGA GATGCTTGAA 1320 CTCCCAGCCC CTGCTGAAGT GGCTGCCAAA AATCAGAGCT TGGAGGGGGA TACAACAAAG 1380 GGGACTTCAG AAATGTCAGA GAAGAGAGGA CCTACTTCCA GCAACCCCAG AAAGAGACAT 1440 CGGGAAGATT CTGATGTGGA AATGGTGGAA GATGATTCCC GAAAGGAAAT GACTGCAGCT 1500 TGTACCCCCC GGAGAAGGAT CATTAACCTC ACTAGTGTTT TGAGTCTCCA GGAAGAAATT 1560 AATGAGCAGG GACATGAGGT TCTCCGGGAG ATGTTGCATA ACCACTCCTT CGTGGGCTGT 1620 GTGAATCCTC AGTGGGCCTT GGCACAGCAT CAAACCAAGT TATACCTTCT CAACACCACC 1680 AAGCTTAGTG AAGAACTGTT CTACCAGATA CTCATTTATG ATTTTGCCAA TTTTGGTGTT 1740 CTCAGGTTAT CGGAGCCAGC ACCGCTCTTT GACCTTGCCA TGCTTGCCTT AGATAGTCCA 1800 GAGAGTGGCT GGACAGAGGA AGATGGTCCC AAAGAAGGAC TTGCTGAATA CATTGTTGAG 1860 TTTCTGAAGA AGAAGGCTGA GATGCTTGCA GACTATTTCT CTTTGGAAAT TGATGAGGAA 1920 GGGAACCTGA TTGGATTACC CCTTCTGATT GACAACTATG TGCCCCCTTT GGAGGGACTG 1980 CCTATCTTCA TTCTTCGACT AGCCACTGAG GTGAATTGGG ACGAAGAAAA GGAATGTTTT 2040 GAAAGCCTCA GTAAAGAATG CGCTATGTTC TATTCCATCC GGAAGCAGTA CATATCTGAG 2100 GAGTCGACCC TCTCAGGCCA GCAGAGTGAA GTGCCTGGCT CCATTCCAAA CTCCTGGAAG 2160 TGGACTGTGG AACACATTGT CTATAAAGCC TTGCGCTCAC ACATTCTGCC TCCTAAACAT 2220

240

TTCACAGAAG	ATGGAAATAT	CCTGCAGCTT	GCTAACCTGC	CTGATCTATA	CAAAGTCTTT	2280
GAGAGGTGTT	AAATATGGTT	ATTTATGCAC	TGTGGGATGT	GTTCTTCTTT	CTCTGTATTC	2340
CGATACAAAG	TGTTGTATCA	AAGTGTGATA	TACAAAGTGT	ACCAACATAA	GTGTTGGTAG	2400
CACTTAAGAC	TTATACTTGC	CTTCTGATAG	TATTCCTTTA	TACACAGTGG	ATTGATTATA	2460
aataaataga	TGTGTCTTAA	CATA				2484

(2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 756 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

(xi)	SEQ	JENCI	E DES	SCRI	PTIO	V: SI	EQ II	ON C	:5:						
Met	Ser	Phe	Val	Ala	Gly	Val	Ile	Arg	Arg	Leu	Asp	Glu	Thr	Val	Val
1				5			f .		10					15	
Asn	Arg	Ile	Ala	Ala	Gly	Glu	Val	Ile	Gln	Arg	Pro	Ala	Asn	Ala	Ile
			20					25					30		
Lys	Glu		Ile	Glu	Asn	Cys		Asp	Ala	rys	Ser		Ser	Ile	Gln
		35	_			_	40					45	_		
Val		Val	Lys	Glu	Gly	_	Leu	Lys	Leu	Ile		Ile	Gln	Asp	Asn
-1	50				_	55	_				60			_	
_	Thr	GTÅ	TTE	-	-	GIU	Asp	Leu	Asp		Val	сув	GIU	Arg	
65	Mh	0	*		70	a	Dh.a	.		75			-1 -		80
Inr	THE	Ser	rys	85	GIN	ser	Pne	GIU	Asp	red	Ala	ser	TIE	Ser 95	THE
T177	Gly	Dha	Arc		Glu	מות	Lau	212		Tle	502	ui e	t/a1	Ala	wia
TYL	· GIY	I II C	100	GLY	Gia	ura.	neu	105	Ser	116	Ser	UTB	110	VIG	****
Val	Thr	Ile		Thr	Lvs	Thr	Ala		Glv	Lvs	Cvs	Ala	-	Arg	Ala
	•	115			•		120	•	•	-	-	125	-	•	
Ser	Tyr	Ser	Asp	Gly	Lys	Leu	Lys	Ala	Pro	Pro	Lys	Pro	Сув	Ala	Gly
	130					135					140				
Asn	Gln	Gly	Thr	Gln	Ile	Thr	Val	Glu	Asp	Leu	Phe	Tyr	Asn	Ile	Ala
145					150					155					160
Thr	Arg	Arg	Lys	Ala	Leu	Lys	Asn	Pro	Ser	Glu	Glu	Tyr	Gly	Lys	Ile
				165				14.	170					175	
Leu	Glu	Val		Gly	Arg	Tyr	Ser		His	Asn	Ala	Gly		Ser	Phe
			180					185					190		
Ser	Val		Lys	Gln	Gly	Glu		Val	Ala	Asp	Val	_	Thr	Leu	Pro
_		195			_	_	200	_				205	_		
Asn		Ser	Thr	Val	Asp		Ile	Arg	Ser	Ile		Gly	Asn	Ala	vaı
0	210	01	T 01-	T1-	~1	215	C1	a	01	7	220	Mb s-	T -01-	7 J -	Dho
ser	wid	GIU	Leu	TTG	GIU	TTG	GTÀ	cys	GIU	Asp	TAR	rnr	rea	Ala	FILE

235

230

Lys	Met	Asn	Gly	Tyr 245	Ile	Ser	Asn	Ala	Asn 250	Tyr	Ser	Val	Lys	Lys 255	Cys
Ile	Phe	Leu	Leu 260	Phe	Ile	Asn	His	Arg 265	Leu	Val	Glu	Ser	Thr 270	Ser	Let
Arg	Lys	Ala 275	Ile	Glu	Thr	Val	Tyr 280	Ala	Ala	Tyr	Leu	Pro 285	Lys	Asn	Thr
His	Pro 290	Phe	Leu	Tyr	Leu	Ser 295	Leu	Glu	Ile	Ser	Pro 300	Gln	Asn	Val	Asp
Val 305	Asn	Val	His	Pro	Thr 310	Lys	His	Glu	Val	His 315	Phe	Leu	His	Glu	G10
Ser	Ile	Leu	Glu	Arg 325	Val	Gln	Gln	His	Ile 330	Glu	Ser	Lys	Leu	Leu 335	Gly
Ser	Asn	Ser	Ser 340	Arg	Met	Tyr	Phe	Thr 345	Gln	Thr	Leu	Leu	Pro 350	Gly	Let
Ala	Gly	Pro 355	Ser	Gly	Glu	Met	Val 360	Lys	Ser	Thr	Thr	Ser 365	Leu	Thr	Ser
Ser	Ser 370	Thr	Ser	Gly	Ser	Ser 375	Asp	Lys	Val	Tyr	Ala 380	His	Gln	Met	Va]
Arg 385	Thr	Asp	Ser	Arg	Glu 390	Gln	Lys	Leu	Asp	Ala 395	Phe	Leu	Gln	Pro	Leu 400
Ser	Lys	Pro	Leu	Ser	Ser	Gln	Pro	Gln	Ala 410	Ile	Val	Thr	Glu	Asp 415	Lys
Thr	Asp	Ile	Ser 420		Gly	Arg	Ala	Arg 425		Gln	Asp	Glu	Glu 430		Leu
Glu	Leu	Pro 435	,	Pro	Ala	Glu	Val		Ala	Lys	Asn	Gln 445		Leu	Glu
Gly	Asp 450		Thr	ГЛа	Gly	Thr		Glu	Met	Ser	Glu 460		Arg	Gly	Pro
Thr 465	Ser	Ser	Asn	Pro	Arg 470	Lys	Arg	His	Arg	Glu 475	Asp	Ser	Asp	Val	Glu 480
Met	Val	Glu	Asp	Asp 485	Ser	Arg	Lys	Glu	Met 490	Thr	Ala	Ala	Cys	Thr 495	Pro
Arg	Arg	Arg	Ile 500	Ile	Asn	Leu	Thr	Ser 505	Val	Leu	Ser	Leu	Gln 510	Glu	Glu
Ile	Asn	Glu 515	Gln	Gly	His	Glu	Val 520	Leu	Arg	Glu	Met	Leu 525	His	Asn	His
Ser	Phe 530	Val	Gly	Cys	Val	Asn 535	Pro	Gln	Trp	Ala	Leu 540	Ala	Gln	His	Glr
Thr 545	Lys	Leu	Tyr	Leu	Leu 550	Asn	Thr	Thr	Lys	Leu 555	Ser	Glu	Glu	Leu	Phe 560
Tyr	Gln	Ile	Leu	Ile 565	Tyr	Asp	Phe	Ala	Asn 570	Phe	Gly	Val	Leu	Arg 575	Leu
Ser	Glu	Pro	Ala	Pro	Leu	Phe	Asp	Leu 585	Ala	Met	Leu	Ala	Leu 590	Asp	Ser

Pro	Glu	Ser	Gly	Trp	Thr	Glu	Glu	Asp	Gly	Pro	Lys	Glu	Gly	Leu	Ala
		595					600					605			
Glu	Tyr	Ile	Val	Glu	Phe	Leu	Lys	Lys	Lys	Ala	Glu	Met	Leu	Ala	Asp
	610					615					620				
Tyr	Phe	Ser	Leu	Glu	Ile	Asp	Glu	Glu	Gly	Asn	Leu	Ile	Gly	Leu	Pro
625					630					635					640
Leu	Leu	Ile	Asp	Asn	Tyr	Val	Pro	Pro	Leu	Glu	Gly	Leu	Pro	Ile	Phe
				645					650					655	
Ile	Leu	Arg	Leu	Ala	Thr	Glu	Val	Asn	Trp	Asp	Glu	Glu	Lys	Glu	Cys
			660					665					670		
Phe	Glu	Ser	Leu	Ser	Lys	Glu	Cys	Ala	Met	Phe	Tyr	Ser	Ile	Arg	Lys
		675					680					685			
Gln	Tyr	Ile	Ser	Glu	Glu	Ser	Thr	Leu	Ser	Gly	Gln	Gln	Ser	Glu	Val
	690					695					700				
Pro	Gly	Ser	Ile	Pro	Asn	Ser	Trp	Lys	Trp	Thr	Val	Glu	His	Ile	Val
705	_				710		-	-	•	715					720
Tyr	Lys	Ala	Leu	Arq	Ser	His	Ile	Leu	Pro	Pro	Lvs	His	Phe	Thr	Glu
-	-			725					730		•			735	
Asp	Gly	Asn	Ile	Leu	Gln	Leu	Ala	Asn	Leu	Pro	Asp	Leu	Tyr	Lys	Val
			740					745			-		750	-	
Phe	Glu	Arg	Cys		•										
		755	=												

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 397 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

TGGCTGGATG	CTAAGCTACA	GCTGAAGGAA	GAACGTGAGC	ACGAGGCACT	GAGGTGATTG	60
GCTGAAGGCA	CTTCCGTTGA	GCATCTAGAC	GTTTCCTTGG	CTCTTCTGGC	GCCAAAATGT	120
CGTTCGTGGC	AGGGGTTATT	CGGCGGCTGG	ACGAGACAGT	GGTGAACCGC	ATCGCGGCGG	180
GGGAAGTTAT	CCAGCGGCCA	GCTAATGCTA	TCAAAGAGAT	GATTGAGAAC	TGGTACGGAG	240
GGAGTCGAGC	CGGGCTCACT	TAAGGGCTAC	GACTTAACGG	GCCGCGTCAC	TCAATGGCGC	300
GGACACGCCT	CTTTCCCCGG	GCAGAGGCAT	GTACAGCGCA	TGCCCACAAC	GGCGGAGGCC	360
GCCGGGTTCC	CTACGTGCCA	TAAGCCTTCT	CCTTTTC			397

WO 95/16793

PCT/US94/14746

70	
(2) INFORMATION FOR SEQ ID NO:7:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 393 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
AAACACGTTA ATGAGGCACT ATTGTTTGTA TTTGGAGTTT GTTATCATTG CTTGGCTCAT	6
ATTAAAATAT GTACATTAGA GTAGTTGCAG ACTGATAAAT TATTTTCTGT TTGATTTGCC	12
AGTTTAGATG CAAAATCCAC AAGTATTCAA GTGATTGTTA AAGAGGGAGG CCTGAAGTTG	18
ATTCAGATCC AAGACAATGG CACCGGGATC AGGGTAAGTA AAACCTCAAA GTAGCAGGAT	24
GTTTGTGCGC TTCATGGAAG AGTCAGGACC TTTCTCTGTT CTGGAAACTA GGCTTTTGCA	30
GATGGGATTT TTTCACTGAA AAATTCAACA CCAACAATAA ATATTTATTG AGTACCTATT	36
ATTTGCGGGG CACTGTTCAG GGGATGTGTC AGT	39
*	
(2) INFORMATION FOR SEQ ID NO:8:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 352 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
TTTCCTGGAT TAATCAAGAA ATGGAATTCA AAGAGATTTG GAAAATGAGT AACATGATTA	
TTTACTCATC TTTTTGGTAT CTAACAGAAA GAAGATCTGG ATATTGTATG TGAAAGGTTC	120
ACTACTAGTA AACTGCAGTC CTTTGAGGAT TTAGCCAGTA TTTCTACCTA TGGCTTTCGA	180
GGTG GGTAA GCTAAAGATT CAAGAAATGT GTAAAATATC CTCCTGTGAT GACATTGTCT	24
GTCATTTGTT AGTATGTATT TCTCAACATA GATAAATAAG GTTTGGTACC TTTTACTTGT	30
TAAATGTATG CAAATCTGAG CAAACTTAAT GAACTTTAAC TTTCAAAGAC TG	35
(2) INFORMATION FOR SEC ID NO.C.	
(2) INFORMATION FOR SEQ ID NO:9: (i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 287 base pairs	
(B) TYPE: nucleic acid	

- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

TGGAAGCAGC	AGCAGATAAC	CTTTCCCTTT	GGTGAGGTGA	CAGTGGGTGA	CCCAGCAGTG	60
AGTTTTTCTT	TCAGTCTATT	TTCTTTTCTT	CCTTAGGCTT	TGGCCAGCAT	AAGCCATGTG	120
GCTCATGTTA	CTATTACAAC	GAAAACAGCT	GATGGAAAGT	GTGCATACAG	GTATAGTGCT	180
GACTTCTTTT	ACTCATATAT	ATTCATTCTG	AAATGTATTT	TGGGCCTAGG	TCTCAGAGTA	240
ATCCTCTCTC	AACACCAGTG	TTATCTTTGG	CAGAGATCTT	GAGTACG		287

(2) INFORMATION FOR SEQ ID NO:10:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 336 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
TTGATATGAT TTTCTCTTTT CCCCTTGGGA TTAGTATCTA TCTCTCTACT GGATATTAAT	60
TTGTTATATT TTCTCATTAG AGCAAGTTAC TCAGATGGAA AACTGAAAGC CCCTCCTAAA	120
CCATGTGCTG GCAATCAAGG GACCCAGATC ACGGTAAGAA TGGTACATGG GAGAGTAAAT	180
TGTTGAAGCT TTGTTTGTAT AAATATTGGA ATAAAAAATA AAATTGCTTC TAAGTTTTCA	240
GGGTAATAAT AAAATGAATT TGCACTAGTT AATGGAGGTC CCAAGATATC CTCTAAGCAA	300
GATAAATGAC TATTGGCTTT TTGGCATGGC AGCCTG	336
(2) INFORMATION FOR SEQ ID NO:11:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 275 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
GCTTTTGCCA GGACCATCTT GGGTTTTATT TTCAAGTACT TCTATGAATT TACAAGAAAA	60
ATCAATCTTC TGTTCAGGTG GAGGACCTTT TTTACAACAT AGCCACGAGG AGAAAAGCTT	120
TAAAAAATCC AAGTGAAGAA TATGGGAAAA TTTTGGAAGT TGTTGGCAGG TACAGTCCAA	180
AATCTGGGAG TGGGTCTCTG AGATTTGTCA TCAAAGTAAT GTGTTCTAGT GCTCATACAT	240
TGAACAGTTG CTGAGCTAGA TGGTGAAAAG TAAAA	275
(2) INFORMATION FOR SEQ ID NO:12:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 389 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
CAGCAACCTA TAAAAGTAGA GAGGAGTCTG TGTTTTGACG CAGCACCTTT AGCATTTTTA	60
TTTGGATGAA GTTTCTGCTG GTTTATTTTT CTGTGGGTAA AATATTAATA GGCTGTATGG	120
AGATATTTT CTTTATATGT ACCTTTGTTT AGATTACTCA ACTCCACTAA TTTATTTAAC	180
TAAAAGGGGG CTCTGACATC TAGTGTGTGT TTTTGGCAAC TCTTTTCTTA CTCTTTTGTT	240
TTTCTTTTCC AGGTATTCAG TACACAATGC AGGCATTAGT TTCTCAGTTA AAAAAGTAAG	300
TTCTTGGTTT ATGGGGGATG GTTTTGTTTT ATGAAAAGAA AAAAGGGGAT TTTTAATAGT	360
TTGCTGGTGG AGATAAGGTT ATGATGTTT	389

60

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72	
(2) INFORMATION FOR SEQ ID NO:13:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 381 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:	
ATGTTTCAGT CTCAGCCATG AGACAATAAA TCCTTGTGTC TTCTGCTGTT TGTTTATCAG	60
CAAGGAGAGA CAGTAGCTGA TGTTAGGACA CTACCCAATG CCTCAACCGT GGACAATATT	120
CGCTCCATCT TTGGAAATGC TGTTAGTCGG TATGTCGATA ACCTATATAA AAAAATCTTT	180
TACATTTATT ATCTTGGTTT ATCATTCCAT CACATTATTT GGGAACCTTT CAAGATATTA	240
TGTGTGTTAA GAGTTTGCTT TAGTCAAATA CACAGGCTTG TTTTATGCTT CAGATTTGTT	300
AATGGAGTTC TTATTTCACG TAATCAACAC TTTCTAGGTG TATGTAATCT CCTAGATTCT	360
GTGGCGTGAA TCATGTGTTC T	381
(2) INFORMATION FOR SEQ ID NO:14:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 526 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	
ACTGAGTAGG GTAGGTGGGT GAGTGGGTGG GTGGGTGGG	300
GGATGGGTGG GTGAATGGGT GAACAGACAA ATGGATGGAT GAATGGACAG GCACAGGAGG	120
ACCTCAAATG GACCAAGTCT TCGGGGCCCT CATTTCACAA AGTTAGTTTA TGGGAAGGAA	180
CCTTGTGTTT TTAAATTCTG AT CITTTGT AATGTTTGAG TTTTGAGTAT TTTCAAAAGC TTCAGAATCT CTTTTCTAAT AGAGAACTGA TAGAAATTGG ATGTGAGGAT AAAACCCTAG	240
CCTTCAAAAT GAATGGTTAC ATATCCAATG CAAACTACTC AGTGAAGAAG TGCATCTTCT	300 360
TACTCTTCAT CAACCGTAAG TTAAAAAGAA CCACATGGGA AATCCACTCA CAGGAAACAC	420
CCACAGGGAA TTTTATGGGA CCATGGAAAA ATTTCTGAGT CCATAGGTTT GATTAAACAT	480
GGAGAAACCT CATGGCAAAG TTTGGTTTTA TTGGGAAGCA TGTATA	526
GANGAMACOL CALOCOMMO ILLOCITIM ILLOCAMOCA IGIALA	220
(2) INFORMATION FOR SEQ ID NO:15:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 434 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:	
5**	

GCATATCACT ACAGAAATGT CTTTCCTGAG GTGATGTCAT GACTTTGTGT GAATGTACAC 120 CTGTGACCTC ACCCCTCAGG ACAGTTTTGA ACTGGTTGCT TTCTTTTTAT TGTTTAGATC 180

618

73	
GTCTGGTAGA ATCAACTTCC TTGAGAAAAG CCATAGAAAC AGTGTATGCA GCCTATTTGC CCAAAAACAC ACACCCATTC CTGTACCTCA GGTAATGTAG CACCAAACTC CTCAACCAAG ACTCACAAGG AACAGATGTT CTATCAGGCT CTCCTCTTTG AAAGAGATGA GCATGCTAAT AGTACAATCA GAGTGAATCC CATACACCAC TGGCAAAAGG ATGTTCTGTC CCTTCTTACA GGTACAAGGC ACAG	300 360 420 434
(2) INFORMATION FOR SEQ ID NO:16: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 458 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ.ID NO:16:	
CTTACGCAAA GCTACACAGC TCTTAAGTAG CAGTGCCAAT ATTTGAACAC ACTCAGACTC	60
GAGCCTGAGG TTTTGACCAC TGTGTCATCT GGCCTCAAAT CTTCTGGCCA CCACATACAC	120
CATATGTGGG CTTTTCTCC CCCTCCCACT ATCTAGGTA ATTGTTCTCT CTTATTTTCC	180
TGACAGTTTA GAAATCAGTC CCCAGAATGT GGATGTTAAT GTGCACCCCA CAAAGCATGA	240
AGTTCACTTC CTGCACGAGG AGAGCATCCT GGAGCGGGTG CAGCAGCACA TCGAGAGCAA	300
GCTCCTGGGC TCCAATTCCT CCAGGATGTA CTTCACCCAG GTCAGGGCGC TTCTCATCCA	360
GCTACTTCTC TGGGGCCTTT GAAATGTGCC CGGCCAGACG TGAGAGCCCA GATTTTTGCT	420
GTTATTTAGG AACTTTTTT GAAGTATTAC CTGGATAG	458
(2) INFORMATION FOR SEQ ID NO:17: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 618 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
GATAATTATA CCTCATACTA GCTTCTTTCT TAGTACTGCT CCATTTGGGG ACCTGTATAT	60
CTATACTTCT TATTCTGAGT CTCTCCACTA TATATATATA TATATATATA TTTTTTTT	120
TTTTTTTTT TAATACAGAC TTTGCTACCA GGACTTGCTG GCCCCTCTGG GGAGATGGTT	180
AAATCCACAA CAAGTCTGAC CTCGTCTTCT ACTTCTGGAA GTAGTGATAA GGTCTATGCC	240
CACCAGATGG TTCGTACAGA TTCCCGGGAA CAGAAGCTTG ATGCATTTCT GCAGCCTCTG	300
AGCAAACCCC TGTCCAGTCA GCCCCAGGCC ATTGTCACAG AGGATAAGAC AGATATTTCT	360
AGTGGCAGGG CTAGGCAGCA AGATGAGGAG ATGCTTGAAC TCCCAGCCCC TGCTGAAGTG	420
GCTGCCAAAA ATCAGAGCTT GGAGGGGGAT ACAACAAAGG GGACTTCAGA AATGTCAGAG	480
ARRICAGIAC CTACTTCCAG CAACCCCAGG TATGGCCTTT TGGGAAAAGT ACAGCCTACC	540

TCCTTTATTC TGTAATAAAA CTGCCTTCTA ACTTTGGCTT TTCATGAATC ACTTGCATCT 600

TCTCTCTGCC GACTTCCC

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(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 478 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:	
CTGTGCTCCA GCACAGGTCA TCCAGCTCTG TAGACCAGCG CAGAGAAGTT GCTTGCTCCC	60
AAATGCAACC CACAAAATTT GGCTAAGTTT AAAAACAAGA ATAATAATGA TCTGCACTTC	120
CTTTTCTTCA TTGCAGAAAG AGACATCGGG AAGATTCTGA TGTGGAAATG GTGGAAGATG	180
ATTCCCGAAA GGAAATGACT GCAGCTTGTA CCCCCCGGAG AAGGATCATT AACCTCACTA	240
GTGTTTTGAG TCTCCAGGAA GAAATTAATG AGCAGGGACA TGAGGGTACG TAAACGCTGT	300
GGCCTGCCTG GGATGCATAG GGCCTCAACT GCCAAGGTTT TGGAAATGGA GAAAGCAGTC	360
ATGTTGTCAG AGTGGCACTA CAGTTTTGAT GGGCAAGCTC CTCTTCCTTT ACTAACCCAC	420
AATAGCATCA GCTTAAAGAC AATTTTTGAT TGGGAGAAAA GGGAGAAAAT AATCTCTG	478
(2) INFORMATION FOR SEQ ID NO:19:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 377 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	•
CAGTTTTCAC CAGGAGGCTC AAATCAGGCC TTTGCTTACT TGGTGTCTCT AGTTCTGGTG	60
CCTGGTGCTT TGGTCAATGA AGTGGGGTTG GTAGGATTCT ATTACTTACC TGTTTTTTGG	120
TTTTATTTT TGTTTTGCAG TTCTCCGGGA GATGTTGCAT AACACTCCT TCGTGGGCTG	180
TGTGAATCCT CAGTGGGCCT TGGCACAGCA TCAAACCAAG TTATACCTTC TCAACACCAC	240
CAAGCTTAGG TAAATCAGCT GAGTGTGTGA ACAAGCAGAG CTACTACAAC AATGGTCCAG	300
GGAGCACAGG CACAAAAGCT AAGGAGAGCA GCATGAAGGT AGTTGGGAAG GGCACAGGCT	360
TTGGAGTCAG CACATGT	377
(2) INFORMATION FOR SEQ ID NO:20:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 325 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
CCCCTGGTTG AAGCGTTGGA ATCCCACTCT TTGGAAGATT GTGTTAGACT GTTAACCAGA	60
TTECACAGCC AGGCAGAACT ATGTCTGTCT CATCCATGTG TCAGGGATTA CGTCTCCCAT	120
TTGTCCCAAC TGGTTGTATC TCAAGCATGA ATTCAGCTTT TCCTTAAAGT CACTTCATTT	180
TTATTTCAG TGAAGAACTG TTCTACCAGA TACTCATTTA TGATTTTGCC AATTTTGGTG	240

TTCTCAGGTT ATCGGTAAGT TTAGATCCTT TTCACTTCTG ACATTTCAAC TGACCGCCCC GCAAACAGTA GCTCTCCACT AAATA	300 325
(2) INFORMATION FOR SEQ ID NO:21:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 341 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	
CATTTATGGT TTCTCACCTG CCATTCTGAT AGTGGATTCT TGGGAATTCA GGCTTCATTT	60
GGATGCTCCG TTAAAGCTTG CTCCTTCATG TTCTTGCTTC TTCCTAGGAG CCAGCACCGC	120
TCTTTGACCT TGCCATGCTT GCCTTAGATA GTCCAGAGAG TGGCTGGACA GAGGAAGATG	180
GTCCCAAAGA AGGACTTGCT GAATACATTG TTGAGTTTCT GAAGAAGAAG GCTGAGATGC	240
TTGCAGACTA TTTCTCTTTG GAAATTGATG AGGTGTGACA GCCATTCTTA TACTTCTGTT	300
GTATTCTCCA AATAAAATTT CCAGCCGGGT GCATTGGCTC A	341
(2) INFORMATION FOR SEQ ID NO:22:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 260 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
CAGATAGGAG GCACAAGGCC TGGGAAAGGC ACTGGAGAAA TGGGATTTGT TTAAACTATG	60
ACAGCATTAT TTCTTGTTCC CTTGTCCTTT TTCCTGCAAG CAGGAAGGGA ACCTGATTGG	120
ATTACCCCTT CTGATTGACA ACTATGTGCC CCCTTTGGAG GGACTGCCTA TCTTCATTCT	180
TCGACTAGCC ACTGAGGTCA GTGATCAAGC AGATACTAAG CATTTCGGTA CATGCATGTG	240
TGCTGGAGGG AAAGGGCAAA	260
(2) INFORMATION FOR SEQ ID NO:23:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 340 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
CTATATCTTC CCAGCAATAT TCACAGTCCG TTTACAGTTT TAACGCCTAA AGTATCACAT	60
TTCGTTTTTT AGCTTTAAGT AGTCTGTGAT CTCCGTTTAG AATGAGAATG TTTAAATTCG	120
TACCTATTTT GAGGTATTGA ATTTCTTTGG ACCAGGTGAA TTGGGACGAA GAAAAGGAAT	180
CTTTTCARAC CCTCAGTARA GRATCCCCTR TCTTCTATTC CATCCGCRAG CAGTACATAT	240

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CTGAGGAGTC GACCCTCTCA GGCCAGCAGG TACAGTGGTG ATGCACACTG GCACCCCAGG	300
ACTAGGACAG GACCTCATAC ATCTTAGGAG ATGAAACTTG	340
(2) INFORMATION FOR SEQ ID NO:24:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 563 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:	
AATCCTCTTG TGTTCAGGCC TGTGGATCCC TGAGAGGCTA GCCCACAAGA TCCACTTCAA	60
AAGCCCTAGA TAACACCAAG TCTTTCCAGA CCCAGTGCAC ATCCCATCAG CCAGGACACC	120
AGTGTATGTT GGGATGCAAA CAGGGAGGCT TATGACATCT AATGTGTTTT CCAGAGTGAA	180
GTGCCTGGCT CCATTCCAAA CTCCTGGAAG TGGACTGTGG AACACATTGT CTATAAAGCC	240
TTGCGCTCAC ACATTCTGCC TCCTAAACAT TTCACAGAAG ATGGAAATAT CCTGCAGCTT	300
GCTAACCTGC CTGATCTATA CAAAGTCTTT GAGAGGTGTT AAATATGGTT ATTTATGCAC	360
TGTGGGATGT GTTCTTCTTT CTCTGTATTC CGATACAAAG TGTTGTATCA AAGTGTGATA	420
TACAAAGTGT ACCAACATAA GTGTTGGTAG CACTTAAGAC TTATACTTGC CTTCTGATAG	480
TATTCCTTTA TACACAGTGG ATTGATTATA AATAAATAGA TGTGTCTTAA CATAATTTCT	540
TATTTAATTT TATTATGTAT ATA	563
•	
(2) INFORMATION FOR SEQ ID NO:25:	
(i) SEQUENCE CHARACTERISTICS:	•
(A) LENGTH: 137 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:	
CTTGGCTCTT CTGGCGCCAA AATGTCGTTC GTGGCAGGGG TTATTCGGCG GCTGGACGAG	60
ACAGTGGTGA ACCGCATCGC GGCGGGGGAA GTTATCCAGC GGCCAGCTAA TGCTATCAAA	120
GAGATGATTG AGAACTG	137
(2) INFORMATION FOR SEQ ID NO:26:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 91 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:	
TTTAGATGCA AAATCCACAA GTATTCAAGT GATTGTTAAA GAGGGAGGCC TGAAGTTGAT	60
TCAGATCCAA GACAATGCCA CCGGGATCAG G	91
TOTALL GUOUNTOGON OCCUPANTOUS A	2 1

(2) INFORMATION FOR SEQ ID NO:27:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 99 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:	
AAAGAAGATC TGGATATTGT ATGTGAAAGG TTCACTACTA GTAAACTGCA GTCCTTTGAG	60
GATTTAGCCA GTATTTCTAC CTATGGCTTT CGAGGTGAG	99
(2) INFORMATION FOR SEQ ID NO:28:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 74 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:	
GCTTTGGCCA GCATAAGCCA TGTGGCTCAT GTTACTATTA CAACGAAAAC AGCTGATGGA	60
AAGTGTGCAT ACAG	74
(2) INFORMATION FOR SEQ ID NO:29:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 73 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:	
AGCAAGTTAC TCAGATGGAA AACTGAAAGC CCCTCCTAAA CCATGTGCTG GCAATCAAGG	60
GACCCAGATC ACG	73
(2) INFORMATION FOR SEQ ID NO:30:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 92 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:	~ ^
GTGGAGGACC TTTTTTACAA CATAGCCACG AGGAGAAAAG CTTTAAAAAA TCCAAGTGAA	60
GAATATGGGA AAATTTTGGA AGTTGTTGGC AG	92

(2) INFORMATION FOR SEQ ID NO:31:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 43 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:	
GTATTCAGTA CACAATGCAG GCATTAGTTT CTCAGTTAAA AAA	43
(2) INFORMATION FOR SEQ ID NO: 32:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 89 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:	
CAAGGAGAGA CAGTAGCTGA TGTTAGGACA CTACCCAATG CCTCAACCGT GGACAATATT	60
CGCTCCATCT TTGGAAATGC TGTTAGTCG	89
•	
(2) INFORMATION FOR SEQ ID NO:33:	•
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 113 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:	
AGAACTGATA GAAATTGGAT GTGAGGATAA AACCCTAGCC TTCAAAATGA ATGGTTACAT	60
ATCCAATGCA AACTACTCAG TGAAGAAGTG CATCTTCTTA CTCTTCATCA ACC	113
(2) INFORMATION FOR SEQ ID NO:34:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 94 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:	
ATCGTCTGGT AGAATCAACT TCCTTGAGAA AAGCCATAGA AACAGTGTAT GCAGCCTATT	60
TGCCCAAAAA CACACACCCA TTCCTGTACC TCAG	94

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(2) INFORMATION FOR SEQ ID NO:35:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 154 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:	
TTTAGAAATC AGTCCCCAGA ATGTGGATGT TAATGTGCAC CCCACAAAGC ATGAAGTTCA	
CTTCCTGCAC GAGGAGAGCA TCCTGGAGCG GGTGCAGCAG CACATCGAGA GCAAGCTCCT	
GGGCTCCAAT TCCTCCAGGA TGTACTTCAC CCAG	154
(2) INFORMATION FOR SEQ ID NO:36:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 371 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:	
ACTITGCTAC CAGGACTIGC TGGCCCCTCT GGGGAGATGG TTAAATCCAC AACAAGTCTG	60
ACCTCGTCTT CTACTTCTGG AAGTAGTGAT AAGGTCTATG CCCACCAGAT GGTTCGTACA	120
GATTCCCGGG AACAGAAGCT TGATGCATTT CTGCAGCCTC TGAGCAAACC CCTGTCCAGT	180
CAGCCCCAGG CCATTGTCAC AGAGGATAAG ACAGATATTT CTAGTGGCAG GGCTAGGCAG	240
CAAGATGAGG AGATGCTTGA ACTCCCAGCC CCTGCTGAAG TGGCTGCCAA AAATCAGAGC	300
TTGGAGGGGG ATACAACAAA GGGGACTTCA GAAATGTCAG AGAAGAGAGG ACCTACTTCC	360
AGCAACCCCA G	371
.0	
(2) INFORMATION FOR SEQ ID NO:37:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 149 base pairs (B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:	
AAAGAGACAT CGGGAAGATT CTGATGTGGA AATGGTGGAA GATGATTCCC GAAAGGAAAT	60
GACTGCAGCT TGTACCCCC GGAGAAGGAT CATTAACCTC ACTAGTGTTT TGAGTCTCCA	
GGAAGAAATT AATGAGCAGG GACATGAGG	149
CONTRACT INTERCENTAL CONTRACT .	

(2) INFORMATION FOR SEQ ID NO:38:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 109 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:	
TTCTCCGGGA GATGTTGCAT AACCACTCCT TCGTGGGCTG TGTGAATCCT CAGTGGGCCT	60
TGGCACAGCA TCAAACCAAG TTATACCTTC TCAACACCAC CAAGCTTAG	109
(2) INFORMATION FOR SEQ ID NO:39:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 64 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:	
TGAAGAACTG TTCTACCAGA TACTCATTTA TGATTTTGCC AATTTTGGTG TTCTCAGGTT	60
ATCG	64
(2) INFORMATION FOR SEQ ID NO:40:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 165 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:	
GAGCCAGCAC CGCTCTTTGA CCTTGCCATG CTTGCCTTAG ATAGTCCAGA GAGTGGCTGG	60
ACAGAGGAAG ATGGTCCCAA AGAAGGACTT GCTGAATACA TTGTTGAGTT TCTGAAGAAG	
AAGGCTGAGA TGCTTGCAGA CTATTTCTCT TTGGAAATTG ATGAG	165
(2) INFORMATION FOR SEQ ID NO:41:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 93 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: CDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:	
GAAGGGAACC TGATTGGATT ACCCCTTCTG ATTGACAACT ATGTGCCCCC TTTGGAGGGA	60
CTGCCTATCT TCATTCTTCG ACTAGCCACT GAG	93
	-

AGGCACTGAG GTGATTGGC

(2) INFORMATION FOR SEQ ID NO:42:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 114 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 42:	
GTGAATTGGG ACGAAGAAAA GGAATGTTTT GAAAGCCTCA GTAAAGAATG CGCTATGTTC	60
TATTCCATCC GGAAGCAGTA CATATCTGAG GAGTCGACCC TCTCAGGCCA GCAG	114
(2) INFORMATION FOR SEQ ID NO:43:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 360 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:	
AGTGAAGTGC CTGGCTCCAT TCCAAACTCC TGGAAGTGGA CTGTGGAACA CATTGTCTAT	60
AAAGCCTTGC GCTCACACAT TCTGCCTCCT AAACATTTCA CAGAAGATGG AAATATCCTG	120
CAGCTTGCTA ACCTGCCTGA TCTATACAAA GTCTTTGAGA GGTGTTAAAT ATGGTTATTT	180
ATGCACTGTG GGATGTGTTC TTCTTTCTCT GTATTCCGAT ACAAAGTGTT GTATCAAAGT	240
GTGATATACA AAGTGTACCA ACATAAGTGT TGGTAGCACT TAAGACTTAT ACTTGCCTTC	300
TGATAGTATT CCTTTATACA CAGTGGATTG ATTATAAATA AATAGATGTG TCTTAACATA	360
(2) INFORMATION FOR SEQ ID NO:44:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 19 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc feature	
(B) LOCATION: 1	
(B) LOCATION: 1 (D) OTHER INFORMATION: /note= "primers directed to geno	mic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:44:	
(VT) DESCRIBE DEPONITITION. DES ID NO. 11.	

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	02	
(2) INF	FORMATION FOR SEQ ID NO:45:	
(i	.) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 19 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
(ix) FEATURE:	
	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
	(D) OTHER INFORMATION: /note= "primers directed to genom	nic
	intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:45:	
TCGTAGC	CCT TAAGTGAGC	19
	ORMATION FOR SEQ ID NO:46:	
(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 22 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
(ix) FEATURE:	
	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
	(D) OTHER INFORMATION: /note= "primers directed to genom	ic
	intron DNA"	
) SEQUENCE DESCRIPTION: SEQ ID NO:46:	
AATATGT	ACA TTAGAGTAGT TG	22
/2\ TNEC	ORMATION FOR SEQ ID NO:47:	
) SEQUENCE CHARACTERISTICS:	
(±)	(A) LENGTH: 19 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
(iv)	FEATURE:	
(12)	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
	(D) OTHER INFORMATION: /note= "primers directed to genom	٠.
	intron DNA"	TG
(ri)	SEQUENCE DESCRIPTION: SEQ ID NO:47:	
(**)	opsorion publication: Spa in Main's	

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CAGAGAAAGG TCCTGACTC

AACCTTTCCC TTTGGTGAGG

			••		
(2)	INFO	RMATIC	ON FOR SEQ ID NO:48:		
(i) SEQUENCE CHARACTERISTICS:					
		(A)	LENGTH: 22 base pairs		
		(B)	TYPE: nucleic acid		
		(C)	STRANDEDNESS: single		
		(D)	TOPOLOGY: linear		
	(ix)	FEATU	TRE:		
		(A)	NAME/KEY: misc_feature		
		(B)	LOCATION: 1		
		(D)	OTHER INFORMATION: /note= "primers direct	ed to	genomic
			intron DNA"	*	
	(xi)	SEQUE	NCE DESCRIPTION: SEQ ID NO:48:		
AGAG	ATTT	GG AAA	ATGAGTA AC		22
(2)	INFO	RMATIC	ON FOR SEQ ID NO:49:		
	(i)	SEQUE	NCE CHARACTERISTICS:		
		(A)	LENGTH: 19 base pairs		
		(B)	TYPE: nucleic acid		
		(C)	STRANDEDNESS: single		
		(D)	TOPOLOGY: linear		
	(ix)	FEATU	JRE:		
		(A)	NAME/KEY: misc_feature		
		(3)	LOCATION: 1		
		(D)	OTHER INFORMATION: /note= "primers direct	ed to	genomic
			intron DNA"		
	(xi)	SEQUE	NCE DESCRIPTION: SEQ ID NO:49:		-
ACAF	ATGTC	AT CAC	AGGAGG		19
(2)	INFO	RMATIC	on FOR SEQ ID NO:50:		
	(i)	SEQUE	INCE CHARACTERISTICS:		
		(A)	LENGTH: 20 base pairs		
		• ,	TYPE: nucleic acid		
		• •	STRANDEDNESS: single		
		• •	TOPOLOGY: linear		
	(ix)	FEATU			
			NAME/KEY: misc_feature		
		• ,	LOCATION: 1		
		(D)	OTHER INFORMATION: /note= "primers direct	ed to	genomic
			intron DNA"		
	(xi)	SEQUE	ENCE DESCRIPTION: SEQ ID NO:50:		

(2) INFORMATION FOR SEQ ID NO:51: (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:51:
GATTACTCTG AGACCTAGGC 20
(2) INFORMATION FOR SEQ ID NO:52:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 22 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:52:
GATTITCTCT TTTCCCCTTG GG 22
• *
(2) INFORMATION FOR SEQ ID NO:53:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:53:
CAAACAAAGC TTCAACAATT TAC 23

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	(2) INFORMATION FOR SEQ ID NO:54:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 26 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(ix) FEATURE:	
,	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
	(D) OTHER INFORMATION: /note= "primers directed to genom	nic
	intron DNA"	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:54:	
	GGGTTTTATT TTCAAGTACT TCTATG	26
	(2) INFORMATION FOR SEQ ID NO:55:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 26 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(ix) FEATURE:	
	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
	. (D) OTHER INFORMATION: /note= "primers directed to genor	nic
	intron DNA"	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:55:	
	GCTCAGCAAC TGTTCAATGT ATGAGC	26
	(2) INFORMATION FOR SEQ ID NO:56:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 18 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
	(ix) FEATURE:	

- (A) NAME/KEY: misc_feature
- (B) LOCATION: 1
- (D) OTHER INFORMATION: /note= "primers directed to genomic intron DNA"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:56:
 CTAGTGTGTG TTTTTGGC 18

(D) OTHER INFORMATION: /note= "primers directed to genomic

DNA"

(B) LOCATION: 1

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:59: GGTTCCCAAA TAATGTGATG G

24

(2) INFORMATION FOR SEQ ID NO:60:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 18 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	•
(D) OTHER INFORMATION: /note= "primers directed	to genomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:60:	
CAAAAGCTTC AGAATCTC	10
(2) INFORMATION FOR SEQ ID NO:61:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 23 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	-
(D) OTHER INFORMATION: /note= "primers directed	to genomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:61:	
CTGTGGGTGT TTCCTGTGAG TGG	2:
(2) INFORMATION FOR SEQ ID NO:62:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 24 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed	to genomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:62:	

CATGACTTTG TGTGAATGTA CACC

AAAATCTGGG CTCTCACG

(2)	INFO	RMATI	ON FOR SEQ ID NO:63:	
	(i)	SEQU	ENCE CHARACTERISTICS:	
		(A)	LENGTH: 24 base pairs	
		(B)	TYPE: nucleic acid	
		(C)	STRANDEDNESS: single	
		(D)	TOPOLOGY: linear	
	(ix)	FEAT	URE:	
		(A)	NAME/KEY: misc_feature	
		(B)	LOCATION: 1	
		(D)	OTHER INFORMATION: /note= "primers directed to genom	nic
			intron DNA"	
	(xi)	SEQU	ENCE DESCRIPTION: SEQ ID NO:63:	
GAG	AGAG	CC TG	ATAGAACA TCTG	24
(2)			ON FOR SEQ ID NO:64:	
	(i)	-	ENCE CHARACTERISTICS:	
		(A)	LENGTH: 20 base pairs	
		(B)	TYPE: nucleic acid	
			STRANDEDNESS: single	
	,	(D)	TOPOLOGY: linear	
	(ix)	FEAT	URE:	
			NAME/KEY: misc_feature	
			LOCATION: 1	
		(D)	OTHER INFORMATION: /note= "primers directed to genome	aic
			intron DNA"	
			ENCE DESCRIPTION: SEQ ID NO:64:	
GGGC	TTTT!	rc TC	CCCCTCCC	20
	T1170		ON TOP STO ID NO. SE.	
(2)		-	ON FOR SEQ ID NO:65:	
	(1)	_	ENCE CHARACTERISTICS:	
			LENGTH: 18 base pairs TYPE: nucleic acid	
			STRANDEDNESS: single	
			TOPOLOGY: linear	
	/ i == \	FEAT		
	(11)			
			NAME/KEY: misc_feature LOCATION: 1	
				ni c
		(0)	OTHER INFORMATION: /note= "primers directed to genor intron DNA"	u I C
	/vi)	SECT	ENCE DESCRIPTION: SEO ID NO:65:	

89
(2) INFORMATION FOR SEQ ID NO:66:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:66:
AATTATACCT CATACTAGC 19
(2) INFORMATION FOR SEQ ID NO:67:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:67:
GTTTTATTAC AGAATAAAGG AGG 23
(2) INFORMATION FOR SEQ ID NO:68:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:68: 19 AAGCCAAAGT TAGAAGGCA

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(2) INFORMATION FOR SEQ ID NO:69:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 20 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to genomi intron DNA"	.C
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:69:	
TGCAACCCAC AAAATTTGGC	20
(2) INFORMATION FOR SEQ ID NO:70:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 20 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to genomi intron DNA"	C
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:70:	
CTTTCTCCAT TTCCAAAACC	20
(2) INFORMATION FOR SEQ ID NO:71:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 18 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	

(A) NAME/KEY: misc_feature

(B) LOCATION: 1

(D) OTHER INFORMATION: /note= "primers directed to genomic intron DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:71:

91
(2) INFORMATION FOR SEQ ID NO:72:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:72:
CATTGTTGTA GTAGCTCTGC 20
(2) INFORMATION FOR SEQ ID NO:73:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:73:
CCCATTTGTC CCAACTGG 18
(2) INFORMATION FOR SEQ ID NO:74:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 19 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:74:
CGGTCAGTTG AAATGTCAG
19

intron DNA"

(2) INFORMATION FOR SEQ ID NO:75:		
(i) SEQUENCE CHARACTERISTICS:		
(A) LENGTH: 22 base pairs		
(B) TYPE: nucleic acid		
(C) STRANDEDNESS: single		
(D) TOPOLOGY: linear		
(ix) FEATURE:		
(A) NAME/KEY: misc_feature		
(B) LOCATION: 1		
(D) OTHER INFORMATION: /note= "primers directed	to	genomic
intron DNA"		
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:75:		
CATTIGGATG CTCCGTTAAA GC		22
(2) INFORMATION FOR SEQ ID NO:76:		
(i) SEQUENCE CHARACTERISTICS:		
(A) LENGTH: 23 base pairs		
(B) TYPE: nucleic acid		
(C) STRANDEDNESS: single		
(D) TOPOLOGY: linear		
(ix) FEATURE:		
(A) NAME/KEY: misc_feature		
(B) LOCATION: 1		
(D) OTHER INFORMATION: /note= "primers directed	tó	genomic.
intron DNA"		
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:76:		
CACCCGGCIG GAAATTTTAT TTG	•	23
•		
(2) INFORMATION FOR SEQ ID NO:77:		
(i) SEQUENCE CHARACTERISTICS:		
(A) LENGTH: 22 base pairs		
(B) TYPE: nucleic acid		
(C) STRANDEDNESS: single		
(D) TOPOLOGY: linear		
(ix) FEATURE:		
(A) NAME/KEY: misc_feature		
(B) LOCATION: 1		
(D) OTHER INFORMATION: /note= "primers directed	to	genomic
intron DNA"		
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:77:		

GGAAAGGCAC TGGAGAAATG GG

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(2) INFORMATION FOR SEQ ID NO:78:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 25 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to gene	omic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:78:	
CCCTCCAGCA CACATGCATG TACCG	25
(2) INFORMATION FOR SEQ ID NO:79:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 20 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to gene	mic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:79:	
TAAGTAGTCT GTGATCTCCG	20
(2) INFORMATION FOR SEQ ID NO:80:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 18 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to generate to gen	omic
intron DNA"	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:80:

ATGTATGAGG TCCTGTCC

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(2) INFORMATION FOR SEQ ID NO:81:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 18 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers direc	ted to genomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:81:	
GACACCAGTG TATGTTGG	18
(2) INFORMATION FOR SEQ ID NO:82:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 20 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers direc	ted to genomic
intron DNA"	
(x;) SEQUENCE DESCRIPTION: SEQ ID NO:82:	
GAGAP AAG AACACATCCC	20
(2) INFORMATION FOR SEQ ID NO:83:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 38 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers direc	ted to denowic

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:83:
TGTAAAACGA CGGCCAGTCA CTGAGGTGAT TGGCTGAA

intron DNA"

	95
(2)	INFORMATION FOR SEQ ID NO:84:
` '	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 19 base pairs
	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: single
	(D) TOPOLOGY: linear
	(ix) FEATURE:
	(A) NAME/KEY: misc_feature
	(B) LOCATION: 1
	(D) OTHER INFORMATION: /note= "primers directed to genomic
	intron DNA"
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:84:
TAGC	CCTTAA GTGAGCCCG 1
(2)	INFORMATION FOR SEQ ID NO:85:
	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 38 base pairs
	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: single
	(D) TOPOLOGY: linear
	(ix) FEATURE:
	(A) NAME/KEY: misc_feature
	(B) LOCATION: 1
	(D) OTHER INFORMATION: /note= "primers directed to genomic
	intron DNA"
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:85:
TGTA	AAACGA CGGCCAGTTA CATTAGAGTA GTTGCAGA
(2)	INFORMATION FOR SEQ ID NO:86:
	(i) SEQUENCE CHARACTERISTICS:
	(A) LENGTH: 19 base pairs
	(B) TYPE: nucleic acid
	(C) STRANDEDNESS: single
	(D) TOPOLOGY: linear
	(ix) FEATURE:
	(A) NAME/KEY: misc_feature

- (B) LOCATION: 1
- (D) OTHER INFORMATION: /note= "primers directed to genomic intron DNA"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:86: AGGTCCTGAC TCTTCCATG

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(2) INFORMATION FOR SEQ ID NO:87:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 40 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to ge	nomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:87:	
TGTAAAACGA CGGCCAGTTT GGAAAATGAG TAACATGATT	40
(2) INFORMATION FOR SEQ ID NO:88:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 19 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to ge	nomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:88:	
TGTCATCACA GGAGGATAT	19
(2) INFORMATION FOR SEC ID NO. 90.	
(2) INFORMATION FOR SEQ ID NO:89:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 38 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(C) STRANDEDNESS: SINGLE (D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	

- (D) OTHER INFORMATION: /note= "primers directed to genomic intron DNA"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:89: TGTAAAACGA CGGCCAGTCT TTCCCTTTGG TGAGGTGA

			97	
(2)	INFO	RMATI	ON FOR SEQ ID NO:90:	
	(i)	SEQU	ENCE CHARACTERISTICS:	
		(A)	LENGTH: 20 base pairs	
		(B)	TYPE: nucleic acid	
		(C)	STRANDEDNESS: single	
		(D)	TOPOLOGY: linear	
	(ix)	FEAT	URE:	
		(A)	NAME/KEY: misc_feature	
		(B)	LOCATION: 1	
		(D)	OTHER INFORMATION: /note= "primers directed to g	enomic
			intron DNA"	
	(xi)	SEQU	ENCE DESCRIPTION: SEQ ID NO:90:	
TACT	CTGA	GA CC	TAGGCCCA	20
(2)	INFO	RMATI	ON FOR SEQ ID NO:91:	
	(i)	SEQU	ENCE CHARACTERISTICS:	
		(A)	LENGTH: 40 base pairs	
		(B)	TYPE: nucleic acid	
		(C)	STRANDEDNESS: single	
		(D)	TOPOLOGY: linear	
	(ix)	FEAT	URE:	
		(A)	NAME/KEY: misc_feature	
		(B)	LOCATION: 1	
•	,	(D)	OTHER INFORMATION: /note= "primers directed to g	genomic
			intron DNA"	
	(xi)	SEQU	ENCE DESCRIPTION: SEQ ID NO:91:	
TGTA	AAAC	GA CG	GCCAGTTC TCTTTTCCCC TTGGGATTAG	40
(2)	INFO	RMATI	ON FOR SEQ ID NO:92:	
	(i)	SEQU	ENCE CHARACTERISTICS:	
		, ,	LENGTH: 23 base pairs	
			TYPE: nucleic acid	
		(C)	STRANDEDNESS: single	

- (D) TOPOLOGY: linear
- (ix) FEATURE:
 - (A) NAME/KEY: misc_feature
 - (B) LOCATION: 1
 - (D) OTHER INFORMATION: /note= "primers directed to genomic intron DNA"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:92:

ACAAAGCTTC AACAATTTAC TCT

98
(2) INFORMATION FOR SEQ ID NO:93:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 46 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:93:
TGTAAAACGA CGGCCAGTGT TTTATTTCA AGTACTTCTA TGAATT 46
(2) INFORMATION FOR SEQ ID NO:94:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:94:
CAGCAACTGT TCAATGTATG AGCACT 26
·
(2) INFORMATION FOR SEQ ID NO:95:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 36 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

- (ix) FEATURE:
 - (A) NAME/KEY: misc_feature
 - (B) LOCATION: 1
 - (D) OTHER INFORMATION: /note= "primers directed to genomic intron DNA"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:95: TGTAAAACGA CGGCCAGTGT GTGTGTTTTT GGCAAC

99	
(2) INFORMATION FOR SEQ ID NO:96:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 18 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to	genomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:96:	
AACCTTATCT CCACCAGC	1
(2) INFORMATION FOR SEQ ID NO:97:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 41 base pairs	
(B) TYPE: nucleic acid	÷
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to	genomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:97:	
TGTAAAACGA CGGCCAGTAG CCATGAGACA ATAAATCCTT G	4
(2) INFORMATION FOR SEQ ID NO:98:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 22 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to	genomic
intron DNA"	

TCCCAAATAA TGTGATGGAA TG 22

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:98:

	100	
(2) INFORMATIO	N FOR SEQ ID NO:99:	
• •	NCE CHARACTERISTICS:	
(A)	LENGTH: 37 base pairs	
(B) '	TYPE: nucleic acid	
(C)	STRANDEDNESS: single	
(D) ⁴	TOPOLOGY: linear	
(ix) FEATU	RE:	
(A) 1	NAME/KEY: misc_feature	
(B) 1	LOCATION: 1	
(D) (OTHER INFORMATION: /note= "primers directed to	genomic
	intron DNA"	
(xi) SEQUE	NCE DESCRIPTION: SEQ ID NO:99:	
TGTAAAACGA CGG	CCAGTAA GCTTCAGAAT CTCTTTT	3.
	N FOR SEQ ID NO:100:	
, ,	NCE CHARACTERISTICS:	
1 1	LENGTH: 23 base pairs	
	TYPE: nucleic acid	
`\\ `	STRANDEDNESS: single	
	TOPOLOGY: linear	
(ix) FEATUR		
	NAME/KEY: misc_feature	
, ,	LOCATION: 1	
, (D) (OTHER INFORMATION: /note= "primers directed to	genomic
	intron DNA"	
	NCE DESCRIPTION: SEQ ID NO:100:	2.
TGGGTGTTTC CTG	IGAGIGG AIT	23
(3) INFORMATION	N FOR SEQ ID NO:101:	
, ,	N FOR SEQ ID NOTION: NCE CHARACTERISTICS:	
	LENGTH: 42 base pairs	
i i	TYPE: nucleic acid	
` '	STRANDEDNESS: single	
• • • • • • • • • • • • • • • • • • • •	TOPOLOGY: linear	
(ix) FEATUR		
• •	NAME/KEY: misc feature	
	LOCATION: 1	
, i	THER INFORMATION: (note: "primers directed to	zenomi c

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:101:
TGTAAAACGA CGGCCAGTAC TTTGTGTGAA TGTACACCTG TG 42

intron DNA"

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1	U	1

101	
(2) INFORMATION FOR SEQ ID NO:102:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 24 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to genomi	¢
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:102:	
GAGAGCCTGA TAGAACATCT GTTG	2
(2) INFORMATION FOR SEQ ID NO:103:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 39 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
' (D) OTHER INFORMATION: /note= "primers directed to genomic	c
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:103:	
TGTAAAACGA CGGCCAGTCT TTTTCTCCCC CTCCCACTA	3
(2) INFORMATION FOR SEQ ID NO:104:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 17 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to genomic	C
intron DNA"	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:104:

17

TCTGGGCTCT CACGTCT

102

102	
(2) INFORMATION FOR SEQ ID NO:105:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 18 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note≈ "primers directed to	genomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:105:	
CTTATTCTGA GTCTCTCC	18
(2) INFORMATION FOR SEQ ID NO:106:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 35 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to	genomic.
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 106:	
TGTAAAACGA CGGCCAGTGT TTGCTCAGAG GCTGC	35
•	
(2) INFORMATION FOR SEQ ID NO:107:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 21 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(ix) FEATURE:	
(A) NAME/KEY: misc_feature	
(B) LOCATION: 1	
(D) OTHER INFORMATION: /note= "primers directed to	genomic
intron DNA"	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:107:	

GATGGTTCGT ACAGATTCCC G

103
(2) INFORMATION FOR SEQ ID NO:108:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 41 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:108:
TGTAAAACGA CGGCCAGTTT ATTACAGAAT AAAGGAGGTA G
(2) INFORMATION FOR SEQ ID NO:109:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 39 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:109:
TGTAAAACGA CGGCCAGTAA CCCACAAAAT TTGGCTAAG
(2) INFORMATION FOR SEQ ID NO:110:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1

(D) OTHER INFORMATION: /note= "primers directed to genomic

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:110:

intron DNA"

TCTCCATTTC CAAAACCTTG

	104	
(2) INFORM	MATION FOR SEQ ID NO:111:	
(i) S	SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 18 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
(ix) F	FEATURE:	
	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
	(D) OTHER INFORMATION: /note= "primers directed to go	enomic
	intron DNA"	
(xi) S	SEQUENCE DESCRIPTION: SEQ ID NO:111:	
TGTCTCTAGT	T TCTGGTGC	1
(2) INFORM	MATION FOR SEQ ID NO:112:	
(i) S	SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 38 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
(ix) F	FEATURE:	
	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
,	(D) OTHER INFORMATION: /note= "primers directed to go	enomic
	intron DNA"	
(xi) S	SEQUENCE DESCRIPTION: SEQ ID NO:112:	•
TGTAAAACGA	A CGGCCAGTTG TTGTAGTAGC TCTGCTTG	3
, ,	MATION FOR SEQ ID NO:113:	
(1) S	SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 20 base pairs	
	(B) TYPE: nucleic acid	
•	(C) STRANDEDNESS: single	
, , , , , , , , , , , , , , , , , , , ,	(D) TOPOLOGY: linear	
` '	FEATURE: (A) NAME/KEY: misc feature	
	(A) NAME/ALI: MISC TEATURE	

(B) LOCATION: 1

(D) OTHER INFORMATION: /note= "primers directed to genomic intron DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO NETT ATTTGTCCCA ACTGGTTGTA

105
(2) INFORMATION FOR SEQ ID NO:114:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 39 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:114:
TGTAAAACGA CGGCCAGTTC AGTTGAAATG TCAGAAGTG 39
(2) INFORMATION FOR SEQ ID NO:115:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:115:
TGTAAAACGA CGGCCAGT 18
(2) INFORMATION FOR SEQ ID NO:116:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:116: CCGGCTGGAA ATTTTATTTG GAG

(2) INFORMATION FOR SEQ ID NO:117:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 41 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:117:
TGTAAAACGA CGGCCAGTAG GCACTGGAGA AATGGGATTT G
(2) INFORMATION FOR SEQ ID NO:118:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primers directed to genomic
intron DNA"
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:118: TCCAGC CAC ATGCATGTAC CGAAAT
TCCAGC. JAC ATGCATGTAC CGAAAT
(2) INFORMATION FOR SEQ ID NO:119:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ix) FEATURE:
(A) NAME/KEY: misc feature
(B) LOCATION: 1
(D) OTHER INFORMATION: /note= "primer directed to genomic
intron DNA"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:119:

GTAGTCTGTG ATCTCCGTTT

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39

107

(2) INF	ORMATION FOR SEQ ID NO:120:	,
(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 36 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
(ix) FEATURE:	
	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
	(D) OTHER INFORMATION: /note= "primers directed to genomic	3
	intron DNA"	
(xi)) SEQUENCE DESCRIPTION: SEQ ID NO:120:	
TGTAAAA	CGA CGGCCAGTTA TGAGGTCCTG TCCTAG	36
(2) INFO	DRMATION FOR SEQ ID NO:121:	
(i)	SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 19 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
(ix)	FEATURE:	
	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
,	(D) OTHER INFORMATION: /note= "primers directed to genomic	;
	intron DNA"	
(xi)	SEQUENCE DESCRIPTION: SEQ ID NO:121:	
ACCAGTGT	TAT GTTGGGATG	19
	? **	
(2) INFO	DRMATION FOR SEQ ID NO:122:	
(i)	SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 39 base pairs	
	(B) TYPE: nucleic acid	
	(C) STRANDEDNESS: single	
	(D) TOPOLOGY: linear	
(ix)	FEATURE:	
	(A) NAME/KEY: misc_feature	
	(B) LOCATION: 1	
	(D) OTHER INFORMATION: /note= "primers directed to genomic	2
	intron DNA"	
(xi)	SEQUENCE DESCRIPTION: SEO ID NO:122:	

TGTAAAACGA CGGCCAGTGA AAGAAGAACA CATCCCACA

(2)	INFO	RMAT	ION	FOR	SEQ	ID N	0:12	3:								
	(i)	SEQ	UENC	E CH	ARAC	TERI	STIC	s:								
		(A) LE	ngth	: 77	0 am	ino	acid	8							
		(B) TY	PE:	amin	o ac	iđ									
		(C) ST	RAND	EDNE	SS:	sing	le								
		(D) TO:	POLO	GY:	line	ar									
	(ii)	MOL	ECUL	E TY	PE: 1	prote	ein									
	(xi)	SEQ	UENC	E DE	SCRI	PTIO	N: S	EQ I	ои о	: 123	:					
	Met	Ser	Leu	Arg	Ile	Lys	Ala	Leu	Asp	Ala	Ser	Val	Val	Asn	Lys	11
	1				5					10					15	
	Ala	Ala	Gly	Glu	Ile	Ile	Ile	Ser	Pro	Val	Asn	Ala	Leu	Lys	Glu	Me
				20					25					30		
	Met	Glu	Asn	Ser	Ile	Asp	Ala	Asn	Ala	Thr	Met	Ile	Asp	Ile	Leu	۷a
			35					40		+			45			
	Lys	Glu	Gly	Gly	Ile	Lys	Val	Leu	Gln	Ile	Thr	Asp	Asn	Gly	Ser	G1
		50					55					60				
	Ile	Asn	Lys	Ala	Asp	Leu	Pro	Ile	Leu	Cys	Glu	Arg	Phe	Thr	Thr	Se
	65					70					75					80
	Lys	Leu	Gln	Lys		Glu	Asp	Leu	Ser		Ile	Gln	Thr	Tyr	_	Ph
					85					90					95	
	Arg	Gly	Glu		Leu	Ala	Ser	lle		His	Val	Ala	i.rg			۷a
				100					105					110		
	Thr	Thr		Val	Lys	Glu	Asp	Arg	Cys	Ala	Trp	Arg		Ser	Tyr	Al
	,	a 1	115	.	T	a 3	0	120		_	,		125		_	
	GIU		гÀг	Met	Leu	GIU		Pro	Lys	Pro	Val		Gly	Lys	Asp	GT.
	mh	130	T 10	T 011	1701		135	Leu	7h a	D b =	*	140	D	0	>	• -
	145	TILL	116	Deu.	Val	150	nsp	Leu	Pile	Pne	155	TIG	Pro	ser	Arg	16
		Δla	T.en	Ara	Ser		Aan	Asp	Glu	Tree.		Tuc	Tlo	Tou	hen	
	••••	*	200	9	165			ap	GZU	170	261	Lys	116	neu.	175	٧.
	Val	Glv	Ara	Tvr		Ile	Hig	Ser	Lvs		Tle	Glv	Phe	Ser		T.v
		0-1	9	180					185			O_y		190	٠,٢٥	_,
	Lvs	Phe	Glv		Ser	Asn	Tvr	Ser		Ser	Val	T.vg	Pro		Tvr	Th
			195	•			2	200				-1-	205		-1-	
	Val	Gln	Asp	Arg	Ile	Ārg	Thr	Val	Phe	Asn	Lvs	ser	Val	Ala	Ser	As
		210	_	_		_	215				•	220				
	Leu	Ile	Thr	Phe	His	Ile	Ser	Lys	Val	Glu	Asp		Asn	Leu	Glu	Se
	225					230		-			235					24
	Val	Asp	Gly	Lys	Val	Cys	Asn	Leu	Asn	Phe	Ile	Ser	Lys	Lys	Ser	Il
		-	-		245					250			-	_	255	
	Ser	Leu	Ile	Phe	Phe	Ile	Asn	Asn	Arg	Leu	Val	Thr	Cys	Asp	Leu	Le
				260					265				-	270		
	Arg	Arg	Ala	Leu	Asn	Ser	Val	Tyr	Ser	Asn	Tyr	Leu	Pro	Lys	Gly	Pho
			275					280					285			

Arg	Pro 290	Phe	Ile	Tyr	Leu	Gly 295	Ile	Val	Ile	Asp	Pro	Ala	Ala	Val	Ası
W-1		1721	u i o	Bro	mb~		2	61 11	Va 1	Ara		Lou	Ser	Gl n	N er
305	nsn	Val	urs	PIO	310	nys	arg	GIU	Vai	315	rne	Leu	Ser	GIII	320
Glu	Ile	Ile	Glu	Lys	Ile	Ala	Asn	Gln	Leu	His	Ala	Glu	Leu	Ser	Ala
				325					330					335	
Ile	Asp	Thr	Ser	Arg	Thr	Phe	Lys	Ala	Ser	Ser	Ile	Ser	Thr	Asn	Lys
			340					345					350		
Pro	Glu	Ser	Leu	Ile	Pro	Phe	Asn	Asp	Thr	Ile	Glu	Ser	Asp	Arg	Asr
		355					360					365			
Arg	Lys	Ser	Leu	Arq	Gln	Ala	Gln	Val	Val	Glu	Asn	Ser	Tyr	Thr	Thr
_	370			_		375					380		•		
Δla		Ser	Gln	T.e.11	Ara		λla	T.vc	Ara	Gln		Aen	Lys	T.011	Val
385	*****	201	02	200	390	my s		y 5	**** 9	395	314		₽¥ ª	Dea	400
		_	_ •	_			_					_	_		
Arg	Ile	Asp	Ala		Gln	Ala	Lys	Ile		Ser	Phe	Leu	Ser		Ser
				405					410					415	
Gln	Gln	Phe	Asn	Phe	Glu	Gly	Ser	Ser	Thr	Lys	Arg	Gln	Leu	ser	Glu
			420					425					430		
Pro	Lys	Val	Thr	Asn	Val	Ser	His	Ser	Gln	Glu	Ala	Glu	Lys	Leu	Thr
		435					440					445			
Leu	Asn	Glu	Ser	Glu	Gln	Pro	Arg	Asp	Ala	Asn	Thr	Ile	Asn	Asp	Asr
	450					455					460				
Asp	Lėu	Lys	Asp	Gln	Pro	Lys	Lys	Lys	Gln	Lys	Gln	Leu	Gly	Asp	Tyr
465					470					475					480
Lys	Val	Pro	Ser	Ile	Ala	Asp	Asp	Glu	Lys	Asn	Ala	Leu	Pro	Ile	Ser
				485					490					495	
Lys	Asp	Gly	Tyr	Ile	Arq	Val	Pro	Lys	Glu	Arg	Val	Asn	Val	Asn	Leu
-	-		500		_			505		-			510		
Thr	Ser	Ile	Lvs	Lvs	Leu	Ara	Glu	Lvs	Val	αzA	asa	Ser	Ile	His	Arc
		515	-1-			5	520					525			3
Glu	Leu		Asp	Ile	Phe	Ala		Leu	Asn	Tvr	Val		Val	Val	Agr
	530					535				-1-	540	- 1		•	
Gl.		Ara	A ===	T Ou	71 -		T10	C1 n	u:c	A a m		T vo	Leu	Dho	T or
545	G_14	nrg	nr 9	Deu	550	AT.	116	Gin	nra	555	Leu	пув	Deu	rne	
	N		01	0		0	m	a1	*			01 -	71 -	91	560
TTE	Asp	TYE	GTĀ		vaı	Cys	Tyr	GIU		Pne	TYT	GIN	Ile		Let
			_	565				_	570					575	_
Thr	Asp	Phe		Asn	Phe	Gly	Lys		Asn	Leu	Gln	Ser	Thr	Asn	Val
			580					585					590		
Ser	Asp	Asp	Ile	Val	Leu	Tyr	Asn	Leu	Leu	Ser	Glu	Phe	Asp	Glu	Lev
		595					600					605			
Asn	Asp	Asp	Ala	Ser	Lys	Glu	Lys	Ile	Ile	Ser	Lys	Ile	Trp	Asp	Met
	610					615					620				
Ser	Ser	Met	Leu	Asn	Glu	Tyr	Tyr	Ser	Ile	Glu	Leu	Val	Asn	qeA	Gly
625					630					635					640

110

Leu Asp Asn Asp Leu Lys Ser Val Lys Leu Lys Ser Leu Pro Leu Leu 645 650 Leu Lys Gly Tyr Ile Pro Ser Leu Val Lys Leu Pro Phe Phe Ile Tyr 665 Arg Leu Gly Lys Glu Val Asp Trp Glu Asp Glu Gln Glu Cys Leu Asp 680 Gly Ile Leu Arg Glu Ile Ala Leu Leu Tyr Ile Pro Asp Met Val Pro 695 700 Lys Val Asp Thr Leu Asp Ala Ser Leu Ser Glu Asp Glu Lys Ala Gln 705 710 715 Phe Ile Asn Arg Lys Glu His Ile Ser Ser Leu Leu Glu His Val Leu 725 730 Phe Pro Cys Ile Lys Arg Arg Phe Leu Ala Pro Arg His Ile Leu Lys 745 Asp Val Val Glu Ile Ala Asn Leu Pro Asp Leu Tyr Lys Val Phe Glu 760 765 Arg Cys 770

- (2) INFORMATION FOR SEQ ID NO:124:
 - (i) SEQUENCE CHARACTERISTICS:
 - ' (A) LENGTH: 64 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein
 - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:124:
 - Val Asn Arg Ile Ala Ala Gly Glu Val Ile eln Arg Pro Ala Asn Ala
 - Ile Lys Glu Met Ile Glu Asn Cys Leu Asp Ala Lys Phe Thr Ser Ile
 20 25 30
 - Gln Val Ile Val Lys Glu Gly Gly Leu Lys Leu Ile Gln Ile Gln Asp 35 40 45
 - Asn Gly Thr Gly Ile Arg Lys Glu Asp Leu Asp Ile Val Cys Glu Arg
 50 55 60
- (2) INFORMATION FOR SEQ ID NO:125:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 64 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: protein

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:125: Val Asn Arg Ile Ala Ala Gly Glu Val Ile Gln Arg Pro Ala Asn Ala 10 Ile Lys Glu Met Ile Glu Asn Cys Leu Asp Ala Lys Ser Thr Ser Ile 25 Gln Val Ile Val Lys Glu Gly Gly Leu Lys Leu Ile Gln Ile Gln Asp 40 Asn Gly Thr Gly Ile Arg Lys Glu Asp Leu Asp Ile Val Cys Glu Arg 50 (2) INFORMATION FOR SEQ ID NO:126: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 52 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:126: Pro Ala Asn Ala Ile Lys Glu Met Ile Glu Asn Cys Leu Asp Ala Lys 10 Ser Thr Asn Ile Gln Val Val Val Lys Glu Gly Gly Leu Lys Leu Ile 25 30 Gin Ile Gin Asp Asn Gly Thr Gly Ile Arg Lys Glu Asp Leu Asp Ile 40 Val Cys Glu Arg 50 (2) INFORMATION FOR SEQ ID NO:127: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 64 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:127: Val Asn Lys Ile Ala Ala Gly Glu Ile Ile Ser Pro Val Asn Ala 10 Leu Lys Glu Met Met Glu Asn Ser Ile Asp Ala Asn Ala Thr Met Ile 25 Asp Ile Leu Val Lys Glu Gly Gly Ile Lys Val Leu Gln Ile Thr Asp 40 Asn Gly Ser Gly Ile Asn Lys Ala Asp Leu Pro Ile Leu Cys Glu Arg 50 55

								112								
(2)	INFO	RMAT:	ION I	FOR :	SEQ	ID N	0:12	8:								
	(i)	SEQ	UENC	E CH	ARAC	TERI	STIC	s:								
		(A) LE	NGTH	: 64	ami	no a	cids								
		(B)) TY	PE: a	amin	o ac	id									
		(C)) STI	RANDI	EDNE	ss: s	sing:	le								
		(D)) TO	POLO	GY:	linea	ar									
	(ii)	MOLI	ECULI	TYI	?E:]	prote	ein									
	(xi)	SEQ	JENCI	E DES	SCRI	PTIO	N: S1	EQ II	ONO:	128	:					
	Val	His	Arg	Ile	Thr	Ser	Gly	Gln	Val	Ile	Thr	Asp	Leu	Thr	Thr	Ala
	1				5					10					15	
	Val	Lys	Glu	Leu	Val	Asp	Asn	Ser	Ile	Asp	Ala	Asn	Ala	Asn	Gln	Ile
				20					25					30		
	Glu	Ile	Ile	Phe	Lys	Asp	Tyr	Gly	Leu	Glu	Ser	Ile	Glu	Cys	Ser	Asp
			35					40					45			
	Asn	Gly	Asp	Gly	Ile	Asp	Pro	Ser	Asn	Tyr	Glu	Phe	Leu	Ala	Leu	Lys
		50					55					60				
(2)	INFO	RMATI	ON I	FOR S	SEQ :	ID NO	0:129	€:								
	(i)	SEQU	JENCE	E CH	ARAC:	reri:	STIC	5:	•							
		(A)	LE	IGTH:	64	amir	no a	cids								
		(B)	TYI	?E: a	amino	o aci	id									
		(C)	STE	RANDI		SS: 6		le		•					•	
		(D)	TO	POLO	EDNE:	SS: s linea	sing: ar	le		٠						
•	(ii)	(D)	TOI CUL!	POLOC E TYI	EDNE: GY: :	SS: 6 linea prote	sing: ar ein									
•	(xi)	(D) MOLE SEQU	TOI ECULI JENCI	POLOC E TYPE E DES	EDNE: GY: : PE: : BCRII	SS: s lines prote	sing: ar ein N: SI	EQ II								
•	(xi) Ala	(D) MOLE SEQU	TOI ECULI JENCI	POLOC E TYPE E DES	EDNE: GY: : PE: : CRII Ala	SS: s lines prote	sing: ar ein N: SI	EQ II		Val		Arg	Pro	Ala		val
•	(xi) Ala 1	(D) MOLE SEQU Asn	TOI ECULE JENCI Gln	POLOC E TYPE E DES	EDNE: GY: : PE: : CRII Ala 5	SS: s lines prote PTION Ala	sing: ar ein N: SI Gly	EQ II Glu	Val	Val 10	Glu				15	
•	(xi) Ala 1	(D) MOLE SEQU Asn	TOI ECULE JENCI Gln	POLOC E TYPE E DES Ile Leu	EDNE: GY: : PE: : CRII Ala 5	SS: s lines prote PTION Ala	sing: ar ein N: SI Gly	EQ II Glu	Val Leu	Val 10	Glu		Pro Ala	Thr	15	
•	(xi) Ala 1 Val	(D) MOLE SEQU Asn	TOI ECULE JENCE Gln Glu	POLOCE TYPE DES	EDNE: GY: GY: GY: GY: GY: GY: GY: GY: GY: GY	SS: solines prote PTION Ala Glu	sing: ar ein N: SI Gly Asn	EQ II Glu Ser	Val Leu 25	Val 10 Asp	Glu Ala	Gly	Ala	Thr 30	15 Arg	Ile
•	(xi) Ala 1 Val	(D) MOLE SEQU Asn	TOI ECULE JENCE Gln Glu Asp	POLOCE TYPE DES	EDNE: GY: GY: GY: GY: GY: GY: GY: GY: GY: GY	SS: solines prote PTION Ala Glu	sing: ar ein N: SI Gly Asn	GQ II Glu Ser Gly	Val Leu 25	Val 10 Asp	Glu Ala	Gly	Ala Arg	Thr 30	15 Arg	Ile
	(xi) Ala 1 Val Asp	(D) MOLE SEQUAS Asn Lys	TOI ECULE JENCE Gln Glu Asp 35	POLOC E TYPE E DES Ile Leu 20 Ile	EDNE: FE: p SCRII Ala 5 Val	SS: a linea prote PTION Ala Glu Arg	sing: ar ein N: SI Gly Asn	Glu Ser Gly 40	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp
	(xi) Ala 1 Val Asp	(D) MOLE SEQUASE Lys Lys Gly	TOI ECULE JENCE Gln Glu Asp 35	POLOC E TYPE E DES Ile Leu 20 Ile	EDNE: FE: p SCRII Ala 5 Val	SS: a linea prote PTION Ala Glu Arg	sing: ar ein N: SI Gly Asn Gly	Glu Ser Gly 40	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly Ile Leu	Ala Arg	Thr 30 Ile	15 Arg Arg	Ile Asp
•	(xi) Ala 1 Val Asp	(D) MOLE SEQUAS Asn Lys	TOI ECULE JENCE Gln Glu Asp 35	POLOC E TYPE E DES Ile Leu 20 Ile	EDNE: FE: p SCRII Ala 5 Val	SS: a linea prote PTION Ala Glu Arg	sing: ar ein N: SI Gly Asn	Glu Ser Gly 40	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp
. (2)	(xi) Ala 1 Val Asp	(D) MOLE SEQUE Asn Lys Ile Gly 50	Glu Asp Cys	POLOG E TYPE E DES Ile Leu 20 Ile Gly	EDNE: SY: : SCRII Ala 5 Val Glu	SS: solines prote PTION Ala Glu Arg Lys	sing: ar ein N: SI Gly Asn Gly Lys 55	Glu Ser Gly 40 Asp	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly Ile Leu	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp
(2)	(xi) Ala 1 Val Asp Asn	(D) MOLE SEQUE Asn Lys Ile Gly 50	TOPECULE GIN Glu Asp 35 Cys	POLOC E TYPE E DES Ile Leu 20 Ile Gly	EDNES GY: : SCRII Ala 5 Val Glu Ile	SS: solines prote PTION Ala Glu Arg Lys	sing: ar ein N: SI Gly Asn Gly Lys 55	EQ III Glu Ser Gly 40 Asp	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly Ile Leu	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp
(2)	(xi) Ala 1 Val Asp Asn	(D) MOLE SEQUE Asn Lys Ile Gly 50 RMATI	Glu Asp Cys	POLOC E TYPE E DES Ile Leu 20 Ile Gly	EDNES GY: : SCRII Ala 5 Val Glu Ile	SS: solines prote PTION Ala Glu Arg Lys	sing: ar ein N: SI Gly Asn Gly Lys 55 0:130	Ser Gly 40 Asp	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly Ile Leu	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp
(2)	(xi) Ala 1 Val Asp Asn	(D) MOLE SEQU Asn Lys Ile Gly 50 RMATI SEQU (A)	Glu Asp 35 Cys	POLOCE TYPE DESTRUCTION OF THE CHARACTER CHARA	EDNES SY: : SCRII Ala 5 Val Glu Ile	SS: Elinea prote PTION Ala Glu Arg Lys	sing: ar ein N: SI Gly Asn Gly Lys 55 0:130 STICS	Ser Gly 40 Asp	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly Ile Leu	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp
(2)	(xi) Ala 1 Val Asp Asn	(D) MOLE SEQUE AST Lys Ile Gly 50 RMATI SEQUE (A) (B)	Glu Asp Cys Con i	POLOCE TYPE DESTRUCTION OF CHARACTERS OF CHA	EDNES SY: : SCRII Ala 5 Val Glu Ile SEQ : ARACS ARACS	SS: slines prote PTION Ala Glu Arg Lys ID NO TERIS amir	sing: ar ein N: SI Gly Asn Gly Lys 55 0:130 STICS no actid	Ser Gly 40 Asp	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly Ile Leu	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp
(2)	(xi) Ala 1 Val Asp Asn	(D) MOLE SEQUE Asn Lys Ile Gly 50 RMATI SEQUE (A) (B) (C)	Glu Asp Cys Con i	POLOCE TYPE E TYPE Leu 20 Ile Gly FOR SE CHA	EDNES GY: CRI SCRII Ala 5 Val Glu Ile ARAC: 64 Amino	SS: Sines prote prior Ala Glu Arg Lys ID NO TERIS amir c aci	sing: ar ein N: SI Gly Asn Gly Lys 55 C:130 STICS no ac id sing:	Ser Gly 40 Asp	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly Ile Leu	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp
(2)	(xi) Ala 1 Val Asp Asn	(D) MOLE SEQUE Asn Lys Ile Gly 50 RMATI SEQUE (A) (B) (C) (D)	Glu Asp STI	POLOCE TYPE E TYPE E DES ILE 20 ILE Gly FOR SE E CHA IGTH: PE: A RANDE	EDNES SY: : SCRII Ala 5 Val Glu Ile SEQ : ARACT Amino	SS: slines prote PTION Ala Glu Arg Lys ID NO TERIS amir caci	sing: ar ein N: SI Gly Asn Gly Lys 55 0:130 STICS no ac id sing:	Ser Gly 40 Asp	Val Leu 25 Ala	Val 10 Asp Lys	Glu Ala Leu	Gly Ile Leu	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp
(2)	(xi) Ala 1 Val Asp Asn INFOR	(D) MOLE SEQUE Asn Lys Ile Gly 50 RMATI SEQUE (A) (B) (C) (D) MOLE	Glu Asp 35 Cys CON II IENCE ION II TYI STE TOE	POLOGE TYPE Leu 20 Ile Gly FOR SECHE	EDNES SY: : SCRII Ala 5 Val Glu Ile SEQ : ARACS 64 amino	SS: slines prote PTION Ala Glu Arg Lys ID NO TERIS amir caci	sing: ar ein N: SI Gly Asn Gly Lys 55 0:130 STICS no ac id sing: ein	Glu Ser Gly 40 Asp	Val Leu 25 Ala Glu	Val 10 Asp Lys Leu	Glu Ala Leu Ala	Gly Ile Leu	Ala Arg 45	Thr 30 Ile	15 Arg Arg	Ile Asp

PCT/US94/14746 WO 95/16793

							113								
Val	Lys	Glu	Leu 20	Val	Glu	Asn	Ser	Leu 25	Asp	Ala	Gly	Ala	Thr 30	Arg	Val
Asp	Ile	Авр 35	Ile	Glu	Arg	Gly	Gly 40	Ala	Lys	Leu	Ile	Arg 45	Ile	Arg	Asp
Asn	Gly 50	Сув	Gly	Ile	Lys	Lys 55	Glu	Glu	Leu	Ala	Leu 60	Ala	Leu	Ala	Arg
(2) INFO	TAMS:	ION I	FOR S	SEQ :	ID NO	0:13	1:								
(i)	SEQ	JENC	E CH	ARAC:	reri:	STIC	5:								
	(A)) LEI	NGTH:	64	amin	no a	cids								
	(B)	TYI	PE: a	amino	aci	id									
	(C)	STI	RANDE	DNE	SS: 8	sing:	le								
	(D	TO	POLO	Y:	linea	ar									
(ii)	MOL	CULI	TYI	E: 1	prote	ein									
(xi)	SEQ	JENCI	E DES	CRI	PTIO	N: SI	II QE	NO:	:131	3					
Ala	Asn	Gln	Ile	Ala	Ala	Gly	Glu	Val	Ile	Glu	Arg	Pro	Ala	Ser	Val
1				5					10					15	
Cys	Lys	Glu	Leu	Val	Glu	Asn	Ala	Ile	Asp	Ala	Gly	Ser	Ser	Gln	Ile
			20					25					30		
Ile	Ile	Glu	Ile	Glu	Glu	Ala	Gly	Leu	Lys	Lys	Val	Gln	Ile	Thr	Asp
		35					40					45			
Asn	Gly	His	Gly	Ile	Ala	His	Asp	Glu	Val	Glu	Leu	Ala	Leu	Arg	Arg
	50			•		55					60				
•															
(2) INFOR	TAM	ON 1	FOR S	EQ :	ED NO	0:13	2:	٠							
(i)	SEQ	JĖNCI	E CHA	RAC:	reris	STICS	S:								
	(A)) LEI	NGTH:	268	37 ba	ase p	pairs	3							
•	(B)	TY	PE: r	nucle	eic a	acid									
	(C)	ST	RANDE	EDNE	5S: :	sing:	le.								
	(D)	TO	POLO	Y: :	linea	ar									
(ii)						(gend	omic))							
(viii)															
	• •		P POS			-									
(xi)															
CCATGGAGC															60
GGAAGTCAG															120
AGGAGTTAG															180
ACTATGGAG															240
TCGAAGGCT															300
AGGTTGAAA															360
TCACCATTI															420
ATGGGAAAA															480 540

AGTATGCCAA AATGGTCCAG GTCTTACATG CATACTGTAT CATTTCAGCA GGCATCCGTG TAAGTTGCAC CAATCAGCTT GGACAAGGAA AACGACAGCC TGTGGTATGC ACAGGTGGAA

600

	GCCCCAGCAT	AAAGGAAAAT	ATCGGCTCTG	TGTTTGGGCA	GAAGCAGTTG	CAAAGCCTCA	720
	TTCCTTTTGT	TCAGCTGCCC	CCTAGTGACT	CCGTGTGTGA	AGAGTACGGT	TTGAGCTGTT	780
	CGGATGCTCT	GCATAATCTT	TTTTACATCT	CAGGTTTCAT	TTCACAATGC	ACGCATGGAG	840
	TTGGAAGGAG	TTCAACAGAC	AGACAGTTTT	TCTTTATCAA	CCGGCGGCCT	TGTGACCCAG	900
	CAAAGGTCTG	CAGACTCGTG	AATGAGGTCT	ACCACATGTA	TAATCGACAC	CAGTATCCAT	960
	TTGTTGTTCT	TAACATTTCT	GTTGATTCAG	AATGCGTTGA	TATCAATGTT	ACTCCAGATA	1020
	AAAGGCAAAT	TTTGCTACAA	GAGGAAAAGC	TTTTGTTGGC	agttttaaag	ACCTCTTTGA	1080
	TAGGAATGTT	TGATAGTGAT	GTCAACAAGC	TAAATGTCAG	TCAGCAGCCA	CTGCTGGATG	1140
	TTGAAGGTAA	CTTAATAAAA	ATGCATGCAG	CGGATTTGGA	AAAGCCCATG	GTAGAAAAGC	1200
	AGGATCAATC	CCCTTCATTA	AGGACTGGAG	AAGAAAAAA	AGACGTGTCC	ATTTCCAGAC	1260
	TGCGAGAGGC	CTTTTCTCTT	CGTCACACAA	CAGAGAACAA	GCCTCACAGC	CCAAAGACTC	1320
	CAGAACCAAG	AAGGAGCCCT	CTAGGACAGA	AAAGGGGTAT	GCTGTCTTCT	AGCACTTCAG	1380
	GTGCCATCTC	TGACAAAGGC	GTCCTGAGAT	CTCAGAAAGA	GGCAGTGAGT	TCCAGTCACG	1440
	GACCCAGTGA	CCCTACGGAC	AGAGCGGAGG	TGGAGAAGGA	CTCGGGGCAC	GGCAGCACTT	1500
	CCGTGGATTC	TGAGGGGTTC	AGCATCCCAG	ACACGGGCAG	TCACTGCAGC	AGCGAGTATG	1560
	CGGCCAGCTC	CCCAGGGGAC	AGGGGCTCGC	AGGAACATGT	GGACTCTCAG	GAGAAAGCGC	1620
	CTGAAACTGA	CGACTCTTTT	TCAGATGTGG	ACTGCCATTC	AAACCAGGAA	GATACCGGAT	1680
•	GTAAATTTCG	AGTTTTGCCT	CAGCCAACTA	ATCTCGCAAC	CCCAAACACA	AAGCGTTTTA	1740
	AAAAAGAAGA	AATTCTTTCC	AGTTCTGACA	TTTGTCAAAA	GTTAGTAAAT	ACTCAGGACA	1800
•	TGTCAGCCTC	TCAGGTTGAT	TGAGCTGTGA	AATTAATAA	GAAAGTTGTG	CCCCTGGACT	1860
•	TTTCTATGAG	TTCTTTAGCT	AAACGAATAA	AGCAGTTACA	TCATGAAGCA	CAGCAAAGTG	1920
i	AAGGGGAACA	GAATTACAGG	AAGTTTAGGG	CAAAGATTTG	TCCTGGAGAA	AATCAAGCAG	1980
					TGCAGAAATG		2040
						ATAGTGGACC.	2100
					GCAGCACACC		2160
	•				TGTTAATGAA	•	2220
٠	TAGAAAATCT	GGAAATATTT	AGAAAGAATG	GCTTTGATTT	TGTTATCGAT	GAAAATGCTC	2280
					TAAAAACTGG		2340
					CCCTGGGGTC		2400
(CTTCCCGAGT	CAAGCAGATG	TTTGCCTCCA	GAGCCTGCCG	GAAGTCGGTG	ATGATTGGGA	2460
(CTGCTCTCAA	CACAAGCGAA	TGAAGAAACT	GATCACCCAC	ATGGGGGAGA	TGGGCCACCC	2520
					CCAACCTGGG		2580
	,				TCGCAGATTT	TTATGTTTTG	2640
2	AAAGACAGAG	TCTTCACTAA	CCTTTTTTGT	TTTAAAATGA	AACCTGC		2687

(2) INFORMATION FOR SEQ ID NO:133:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 862 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:133:

Met Glu Arg Ala Glu Ser Ser Ser Thr Glu Pro Ala Lys Ala Ile Lys

1 5 10 15

Pro	Ile	Asp	Arg 20	Lys	Ser	Val	His	Gln 25	Ile	Сув	Ser	Gly	Gln 30	Val	Va]
Leu	Ser	Leu 35	Ser	Thr	Ala	Val	Lys 40	Glu	Leu	Val	Glu	Asn 45	Ser	Leu	Ası
Ala	Gly 50	Ala	Thr	Asn	Ile	Asp 55	Leu	Lys	Leu	Lys	Asp 60	Tyr	Gly	Val	Ası
65	Ile				70					75					80
Glu	Gly	Leu	Thr	Leu 85	Lys	His	His	Thr	Ser 90	Lys	Ile	Gln	Glu	Phe 95	Ala
qaA	Leu	Thr	Gln 100	Val	Glu	Thr	Phe	Gly 105	Phe	Arg	Gly	Glu	Ala 110	Leu	Ser
Ser	Leu	Cys 115	Ala	Leu	Ser	Asp	Val 120	Thr	Ile	Ser	Thr	Cys 125	His	Ala	Ser
Ala	Lys 130	Val	Gly	Thr	Arg	Leu 135	Met	Phe	Asp	His	Asn 140	Gly	Lys	Ile	Ile
Gln 145	Lys	Thr	Pro	Tyr	Pro 150	Arg	Pro	Arg	Gly	Thr 155	Thr	Val	Ser	Val	Glr 160
Gln	Leu	Phe	ser	Thr 165	Leu	Pro	Val	Arg	His 170	Lys	Glu	Phe	Gln	Arg 175	Asr
Ile [.]	Lys	Lys	Ģlu 180	Tyr	Ala	Lys	Met	Val 185	Gln	Val	Leu	His	Ala 190	Tyr	Cys
Ile	Ile	Ser 195	Ala	Gly	Ile	Arg	Val 200	Ser	Cys	Thr	Asn	Gln 205	Leu-	Gly	Glr
Gly	Lys 210	Arg	Gln	Pro	Val	Val 215	Cys	Ile	Gly	Gly	Ser 220	Pro	Ser	Ile	Lys
Glu 225	Asn	Ile	Gly	Ser	Val 230	Phe	Gly	Gln	Lys	Gln 235	Leu	Gln	Ser	Leu	11e
Pro	Phe	Val	Gln	Leu 245	Pro	Pro	Ser	Asp	Ser 250	Val	Cys	Glu	Glu	Tyr 255	Gly
Leu	Ser	Cys	Ser 260	Asp	Ala	Leu	His	Asn 265	Leu	Phe	Tyr	Ile	Ser 270	Gly	Phe
	Ser	275	_				280					285			
Phe	Phe 290	Phe	Ile	Asn	Arg	Arg 295	Pro	Cys	Asp	Pro	Ala 300	Lys	Val	Cys	Arq
Leu 305	Val	Asn	Glu	Val	Tyr 310	His	Met	Tyr	Asn	Arg 315	His	Gln	Tyr	Pro	Phe 320
Val	Val	Leu	Asn	11e 325	Ser	Val	Asp	Ser	Glu 330	Cys	Val	Asp	Ile	Asn 335	Va:
Thr	Pro	Asp	Lys 340	Arg	Gln	Ile	Leu	Leu 345	Gln	Glu	Glu	Lys	Leu 350	Leu	Le
Ala	Val	Leu 355	Lys	Thr	Ser	Leu	Ile	_	Met	Phe	Asp	Ser 365	Asp	Val	Ası

	_	-			a - ·	~ -	A 1					**- *	m 1	71	B	T -
	Lys	Leu 370	Asn	val	ser	Gln	Gln 375	Pro	Leu	Leu	Asp	Val 380	GIU	Gly	Asn	re
	Ile	Lys	Met	His	Ala	Ala	Asp	Leu	Glu	Lys	Pro	Met	Val	Glu	His	Gl
	385					390					395					400
	Asp	Gln	Ser	Pro	Ser	Leu	Arg	Ile	Gly	Glu	Glu	Lys	ГÄв	Asp	Val	Sex
					405					410					415	
	Ile	Ser	Arg	Leu	Arg	Glu	Ala	Phe	Ser	Leu	Arg	His	Thr	Thr	Glu	Ası
				420					425					430		
	Lys	Pro	His	Ser	Pro	Lys	Thr	Pro	Glu	Pro	Arg	Arg	Ser	Pro	Leu	Gly
			435					440					445			
	Gln	Lys	Arg	Gly	Met	Leu	Ser	Ser	Ser	Thr	Ser	Gly	Ala	Ile	Ser	Ası
		450					455					460				
	Lys	Gly	Val	Leu	Arg	Ser	Gln	Lys	Glu	Ala	Val	Ser	Ser	Ser	His	Gly
	465	-			•	470		-			475					480
	Pro	Ser	Asp	Pro	Thr	Asp	Ara	Ala	Glu	Val	Glu	Lvs	Asp	Ser	Gly	His
					485		3			490			•		495	
	Glv	Ser	Thr	Ser		Asp	Ser	Glu	Glv		Ser	Ile	Pro	Asp		Gly
	1			500					505					510		
	Sor	Hic	Cve		Sar	Glu	Tur	A 1 =		Sor	Sar	Pro	Glw	Asp	Ara	Gli
	Jer	HIL	515	Ser	Ser	GIU	TYL	520	MIG	Ser	Ser	FLO	525	nsp	nry	GL
	50×	Cln		ui.	17 - 1	N C TO	50=		C111	Two	7.1 -	D×o		Th-	A cm	A G =
	ser		Glu	ure	AGI	wab		GIII	GIU	гàа	WIG		GIU	Thr	Asp	veř
		530	C		*** 1	3	535	***		3	~1	540	x	m1	01	O
		Pne	ser	Авр	vai		Cys	HIS	ser	Asn		GIU	Asp	Thr	GTA	
	545	Dh a	3	*** 1	T	550	01 -		*1.	.	555		ml	5	3	560
	гÀа	Pne	Arg	Val		Pro	Gin	Pro	TIE		rėn	Ala	Thr	Pro		THE
			-1.		565	-1		-1-	_	570		_	_		575	~ 7
	Lys	Arg	Pne		Lys	GLu	GIu	IIe		Ser	Ser	Ser	Asp	Ile	Cys	Gir
	_			580			_		585	_				590		
	Lys	Leu		Asn	Thr	Gln	Asp		Ser	Ala	Ser	Gln		Asp	Val	Ala
			595					600					605			
	Val	_	Ile	Asn	Lys	Lys		Val	Pro	Leu	Asp		Ser	Met	Ser	Ser
		610					615					620				
	Leu	Ala	Lys	Arg	Ile	_	Gln	Leu	His	His	Glu	Ala	Gln	Gln	Ser	
	625					630					635					640
,	Gly	Glu	Gln	Asn	Tyr	Arg	Lys	Phe	Arg	Ala	Lys	Ile	Cys	Pro	Gly	Glu
					645					650					655	
	Asn	Gln	Ala	Ala	Glu	Asp	Glu	Leu	Arg	Lys	Glu	Ile	Ser	Lys	Thr	Met
				660					665					670		
	Phe	Ala	Glu	Met	Glu	Ile	Ile	Gly	Gln	Phe	Asn	Leu	Gly	Phe	Ile	Ile
			675					680					685			
	Thr	Lys	Leu	Asn	Glu	Asp	Ile	Phe	Ile	Val	Asp	Gln	His	Ala	Thr	Asp
		690				_	695				-	700				~
	Glu	Lys	Tyr	Asn	Phe	Glu	Met	Leu	Gln	Gln	His	Thr	Val	Leu	Gln	Gly
	705	-	-			710										720

Gln	Arg	Leu	Ile	Ala	Pro	Gln	Thr	Leu	Asn	Leu	Thr	Ala	Val	Asn	Glu
				725					730					735	
Ala	Val	Leu	Ile	Glu	Asn	Leu	Glu	Ile	Phe	Arg	Lys	Asn	Gly	Phe	Asp
			740					745					750		
Phe	Val	Ile	Asp	Glu	Asn	Ala	Pro	Val	Thr	Glu	Arg	Ala	Lys	Leu	Ile
		755					760					765			
Ser	Leu	Pro	Thr	Ser	Lys	Asn	Trp	Thr	Phe	Gly	Pro	Gln	Asp	Val	Asp
	770					775					780				
Glu	Leu	Ile	Phe	Met	Leu	Ser	Asp	Ser	Pro	Gly	Val	Met	Сув	Arg	Pro
785					790					795					800
Ser	Arg	Val	Lys	Gln	Met	Phe	Ala	Ser	Arg	Ala	Сув	Arg	Lys	Ser	Val
				805					810					815	
Met	Ile	Gly	Thr	Ala	Leu	Asn	Thr	Ser	Glu	Met	Lys	Lys	Leu	Ile	Thr
			820					825					830		
His	Met	Gly	Glu	Met	Gly	His	Pro	Trp	Asn	Cys	Pro	His	Gly	Arg	Pro
		835					840					845			
Thr	Met	Arg	His	Ile	Ala	Asn	Leu	Gly	Val	Ile	Ser	Gln	Asn		
	850					855					860				
NFOF	ITAM	ON F	OR S	EQ 1	סא סז):134	:								
(i)	SEQU	ENCE	CHA	RACI	TERIS	STICS	3 :								

(2) II

- (A) LENGTH: 903 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:134:

Met Phe His His Ile Glu Asn Leu Leu Ile Glu Thr Glu Lys Arg Cys

Lys Gln Lys Glu Gln Arg Tyr Ile Pro Val Lys Tyr Leu Phe Ser Met

20 25

Thr Gln Ile His Gln Ile Asn Asp Ile Asp Val His Arg Ile Thr Ser 40

Gly Gln Val Ile Thr Asp Leu Thr Thr Ala Val Lys Glu Leu Val Asp 55

Asn Ser Ile Asp Ala Asn Ala Asn Gln Ile Glu Ile Ile Phe Lys Asp

Tyr Gly Leu Glu Ser Ile Glu Cys Ser Asp Asn Gly Asp Gly Ile Asp

Pro Ser Asn Tyr Glu Phe Leu Ala Leu Lys His Tyr Thr Ser Lys Ile 105

Ala Lys Phe Gln Asp Val Ala Lys Val Gln Thr Leu Gly Phe Arg Gly 120

Glu Ala Leu Ser Ser Leu Cys Gly Ile Ala Lys Leu Ser Val Ile Thr 130 135 140

mb	mb	C	D	D	T	A 7 -	7000	T	T 6	G1 ··	m	Acr	Me+	17 = 1	G1
145	Tnr	ser	Pro	PTO	150	WIS	Asp	тÄв	ren	155	TAL	Asp	uec	AGT	160
	T1.	mh	602	Y		Th-	802	A	Agn		Gly	Thr	Thr	Val:	
UTR	TIE	THE	ser	165	7117	1111	Ser	nrg	170	nys	GLY	1111		175	Dea
17-1	602	G1n	Ten		uia	Aan	T.Au	Pro		Ara	Gln	Lys	Glu		Ser
Val	Ser	GIII	180	FIIG	UTO	ASII	neu	185	AUT	ary	GIII	БÃВ	190	r ne	Del
Tara	Thr	Dha		Ara	Gln	Phe	Thr		Cvs	Len	Thr	Val		Gln	Glv
ם עַת	1111	195	Dy D	my	G 111		200	27.0	0,0	204	****	205			1
Tvr	Ala		Ile	Agn	Ala	Ala		Lvs	Phe	Ser	Val	Trp	Asn	Ile	Thr
-,-	210					215		-1-			220				
Pro		Glv	Lvs	Lvs	Asn		Ile	Leu	Ser	Thr		Arg	Asn	Ser	Ser
225		•	•	•	230					235		-			240
	Ara	Lys	Asn	Ile	Ser	Ser	Val	Phe	Gly	Ala	Gly	Gly	Met	Phe	Gly
	_	•		245					250		•	-		255	_
Leu	Glu	Glu	Val	Asp	Leu	Val	Leu	Asp	Leu	Asn	Pro	Phe	Lys	Asn	Arg
			260	•				265					270		_
Met	Leu	Gly	Lys	Tyr	Thr	Asp	Asp	Pro	Asp	Phe	Leu	Asp	Leu	Asp	Tyr
		275	_	-		_	280		-			285			
Lys	Ile	Arg	Val	Lys	Gly	Tyr	Ile	Ser	Gln	Asn	Ser	Phe	Gly	Cys	Gly
	290					295					300				
Arg	'Asn	Ser	Lys	Asp	Arg	Gln	Phe	Ile	Tyr	Val	Asn	Lys	Arg	Pro	Val
305					310					315					320
Glu	Tyr	Ser	Thr	Leu	Leu	Lys	Cys	Cys	Asn	Glu	Val	Tyr	Lys	Thr	Phe
· .				325					330	•				335	
Asn	Asn	Val	Gln	Phe	Pro	Ala	Val	Phe	Leu	Asn	Leu	Glu	Leu	Pro	Met
			340					345					350		
Ser	Leu	Ile	Asp	Val	Asn	Val	Thr	Pro	Asp	Lys	Arg	Val	Ile	Leu	Leu
		355					360					365			
His	Asn	Glu	Arg	Ala	Val	Ile	Asp	Ile	Phe	Lys	Thr	Thr	Leu	Ser	Asp
	370					375					380				
_	Tyr	Asn	Arg	Gln		Leu	Ala	Leu	Pro	-	Arg	Met	Суз	Ser	
385					390					395					400
Ser	Glu	Gln	Gln		Gln	Lys	Arg	Leu		Thr	Glu	Val	Phe		Asp
_	_			405		_	_		410	_	_	•		415	_
Arg	ser	Thr		His	Glu	Ser	Asp		Glu	Asn	Tyr	His		Ala	Arg
_			420			_		425				_	430		
Ser	Glu		Asn	Gln	Ser	Asn		Ala	His	Phe	Asn	Ser	Thr	Thr	Gly
		435		_	_		440				_	445			
vaı		Asp	rys	ser	Asn	_	Thr	Glu	Leu	Thr		Val	Met	Asp	GTA
.	450	m1- · ·	3	**. *	æ.	455			~ 3	0 -	460	-	۵.	*** 1	G
	тĂг	Inr	ASN	vaı		Asp	vaı	TTE	GTĀ		GIU	Cys	GLU	val	
465	7	6	C	77 - 7	470	. .	_		~ 1-	475	6 ~	C	mı- ··	D	480
vall	Азр	ser	ser	Va1			_	Glu			ser	Ser	rnr	Pro 495	
				400					441)					470	

Lys	Lys	Leu	Pro 500	Ser	Ile	Lys	Thr	Asp 505	ser	Gln	Asn	Leu	Ser 510	Asp	Leu
Agn	T.eu	Agn	-	Dhe	Ser	Aan	Pro		Phe	Gln	Agn	Tle		Ser	Pro
		515					520					525			
Yab	Lys 530	Ala	Arg	Ser	Leu	Glu 535	Lys	Val	Val	Glu	Glu 540	Pro	Val	Tyr	Phe
Asp	Ile	Asp	Gly	Glu	Lys	Phe	Gln	Glu	Lys	Ala	Val	Leu	Ser	Gln	Ala
545		-			550					555					560
Asp	Gly	Leu	Val	Phe	Val	Asp	Asn	Glu	Сув	His	Glu	His	Thr	Asn	Asp
				565					570					575	
Cys	Cys	His	Gln	Glu	Arg	Arg	Gly	Ser	Thr	Asp	Ile	Glu	Gln	Asp	Asp
			580					585					590		
Glu	Ala	Asp	Ser	Ile	Tyr	Ala	Glu	Ile	Glu	Pro	Val	Glu	Ile	Asn	Val
		595					600					605			
Arg	Thr	Pro	Leu	Lys	Asn	Ser	Arg	Lys	Ser	Ile	Ser	Lys	Asp	Asn	Tyr
	610					615					620				
Arg	Ser	Leu	Ser	Asp	Gly	Leu	Thr	His	Arg	Lys	Phe	Glu	Asp	Glu	Ile
625					630					635					640
Leu	Glu	Tyr	Asn	Leu	Ser	Thr	Lys	Aśn	Phe	Lys	Glu	Ile	Ser	Làs	Asn
				645					650					655	
Gly	Lys	Gln	Met	Ser	Ser	Ile	Ile	Ser	ГÀа	Arg	Lys	Ser		Ala	Gln
			660					665	•				670		
Glu	Asn		Ile	Lys	Asn	Lys		Glu	Leu	Glu	Asp		Glu	Gln	Gly
1	_	675	•		_		680		_			685		_	
Glu	-	Tyr	Leu	Thr	Leu		Val	Ser	Lys	Asn	-	Phe	Lys	rys	Met
~ 3	690	••- •	~1	61 -	nh -	695	•		71. -	71.	700	**- *	mb	*	T
	var	vai	GIĀ	GIN	Phe 710	Asn	Leu	GIY	Pne	715	TIE	var	THE	Arg	720
705 Val	Acn	Acn	T.3765	Sor	Lýs	T.e.u	Pho	Tle	Val		Gln	Hie	Δla	Ser	
Val	rob	non	Dys	725	пуз	Dea	rne	116	730	пор	G111		Ata	735	F
Glu	Lvs	Tvr	Asn		Glu	Thr	Leu	Gln		Val	Thr	Val	Phe		Ser
	•	-	740					745					750	_	
Gln	Lys	Leu	Ile	Ile	Pro	Gln	Pro	Val	Glu	Leu	Ser	Val	Ile	Asp	Glu
		755					760					765			
Leu	Val	Val	Leu	Asp	Asn	Leu	Pro	Val	Phe	Glu	Lys	Asn	Gly	Phe	Lys
	770					775					780				
Leu	Lys	Ile	Asp	Glu	Glu	Glu	Glu	Phe	Gly	Ser	Arg	Val	Lys	Leu	Leu
785					790					795					800
Ser	Leu	Pro	Thr	Ser	Lys	Gln	Thr	Leu	Phe	Asp	Leu	Gly	Asp	Phe	Asn
				805					810					815	
Glu	Leu	Ile		Leu	Ile	Lys	Glu	Asp	Gly	Gly	Leu	Arg		Asp	Asn
			820					825					830		
Ile				Lys	Ile	_			Phe	Ala				Cys	Arg
		835					840					845			

120

(2) INFORMATION FOR SEQ ID NO:135:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2577 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:135:

TTCCGGCCAA TGCTATCAAA GAGATGATAG AAAACTGTTT AGATGCAAAA TCTACAAATA TTCAAGTGGT TGTTAAGGAA GGTGGCCTGA AGCTAATTCA GATCCAAGAC AATGGCACTG 120 GAATCAGGAA GGAAGATCTG GATATTGTGT GTGAGAGGTT CACTACGAGT AAACTGCAGA 180 CTTTTGAGGA TTTAGCCAGT ATTCTACCT ATGGCTTTCG TGGTGAGCAT TTGGCAAGCA 240 TAAGTCATGT GGCCCATGTC ACTATTACAA CCAAAACAGC TGATGGGAAA TGTGCGTACA 300 GAGCAAGTTA CTCAGATGGA AAGCTGCAAG CCCCTCTAA ACCCTGTGCA GGCAACCAGG 360 GCACCCTGAT CACGGTGGAA GACCTTTTTT ACAACATAAT CACAAGGAGG AAAGCTTTAA 420 AAAATCCAAG TGAAGAGTAC GGAAAAATTT TGGAAGTTGT TGGCAGGTAT TCAATACACA 480 ATTCAGGCAT TAGTATCTCA GTTAAAAAAC AAGGTGAGAC AGTATCTGAT GTCAGAACAC 540 TGCCCAATGC CACAACCGTG GACAACCTTC GCTCCATCTT TGGAAATGCG GTTAGTCGAG 600 AACTGATAGA AGTTGGGTGT GAG' AAAA CCCTAGCTTT CAAAATGAAT GGCTATATAT 660 720 CGAATGCAAA GTATTCAGTG AAGTGCA TTTTCCTACT CTTCATCAAC CACCGTCTGG 780 TAGAATCAGC TGCCTTGAGA AAAGCCATTG AAACTGTATA TGCAGCATAC TTGCCAAAAA CACACACCCA TTCCTGTACC TCAGTTTGAA ATCAGCCCTC AGAACGTGAC GTCAATGTAC 840 ACCCCACCAA GACAGAAGTT CATTTTCTGC ACGAGGAGAG CATTCTGCAG CGTGTGCAGC 900 AGCACATTGA GAGCAAGCTG CTGGGCTCCA ATTCCTCCAG GATGTATTTC ACCCAGACCT 960 TGCTTCCAGG ACTTGCTGGG CCTCTGGGGA GGCAGCTAGA CCCACGACAG GGGTGGCTTC 1020 CTCATCCACT AGTGGAAGTG GCGACAAGGT CTACGCTTAC CAGATGTCGC GTACGGACTC 1080 CCGGGATCAG AAGCTTGACG CCTTTCTGCA GCCTGTAACC AGCCTTGTGC CCAGCCAGCC 1140 CCAGGACCCT CGCCCTGTCC GAGGGGCCAG GACAGAGGGC TCTCCTGAAA GGGCCACGCG 1200 GGAGGATGAG GAGATGCTTG CTCTCCCAGC CCCCGCTGAA GCAGCTGCTG AGAGTGAGAA 1260 CTTGGAGAGG GAATCACTAA TGGAGACTTC AGACGCAGCC CAGAAAGCGG CACCCACTTC 1320 CAGTCCAGGA AGCTCCAGAA AGAGTCATCG GGAGGACTCT GATGTGGAAA TGGTGGAAAA 1380 TGCTTCCGGG AAGGAAATGA CAGCTGCTTG CTACCCCAGG AGGAGGATCA TTAACCTCAC 1440 CAGCGTCTTG AGTCTCCAGG AAGAGATTAG TGAGCGGTGC CATGAGACTC TCCGGGAGAT 1500 ACTCCGTAAC CATTCCTTTG TGGGCTGTGT GAATCCTCAG TGGGCCTTGG CACAGCACCA 1560 GACCAAGCTA TACCTCCTCA ACACTACCAA GCTCAGTGAA GAGCTGTTCT ACCAGATACT 1620 CATTTATGAT TTTGCCAACT TTGGTGTTCT GAGGTTATCG GAACCAGCGC CACTCTTCGA 1680 CCTGGCCATG CTGGCTTAGA CAGTCCTGAA AGTGGCTGGA CAGAGGACGA CGGCCCGAAG 1740

	AAGGGCTTGC	AGAGTACATT	GTCGAGTTTC	TGAAGAGAAG	CGAGATGCTT	GCAGACTATT	1800
,	CTCTGTGAGA	TCGATGAGAA	GGGAACCTGA	TTGATTACTC	TTCTGATGAC	AGCTATGTGC	1860
	CACCTTTGGA	GGGACTGCCT	ATCTTCATTC	TTCGACTGGC	CACTGAGGTG	AATTGGGTGA	1920
	AGAAAAGGAG	TGTTTTGAAA	GTCTCAGTAA	AGAATGTGCT	ATGTTTTACT	CCATTCGGAA	1980
(GCAGTATATA	CTGGAGGAGT	CGACCCTCTC	AGGCCAGCAG	AGTGACATGC	CTGGCTCCAC	2040
•	GTCAAAGCCC	TGGAAGTGGA	CTGTGGAGCA	CATTATCTAT	AAAGCCTTCC	GCTCACACCT	2100
(CCTACCTCCG	AAGCATTTCA	CAGAAGATGG	CAATGTCCTG	CAGCTTGCCA	ACCTGCCAGA	2160
	TCTATACAAA	GTCTTTGAGC	GGTGTTAAAT	ACAATCATAG	CCACCGTAGA	GACTGCATGA	2220
4	CCATCCAAGG	CGAAGTGTAT	GGTACTAATC	TGGAAGCCAC	AGAATAGGAC	ACTTGGTTTC	2280
	AGCTCCAGGG	TTTTCAGTGC	TCACTATTCT	TGTTCTGTAT	CCCAGTATTG	GTGCTGCAAC	2340
•	TTAATGTACT	TCACCTGTGG	ATTGGCTGCA	AATAAACTCA	CGTGTATTGG	AAAAAAGGAA	2400
	TTCCTGCAGC	CCGGGGGATC	CACTAGTTCT	AGAGCGGCCG	CCACCGGTGG	AGCTCCAGCT	2460
•	TTTGTTCCCT	TTAGTGAGGG	TTAATTTCGA	GCTTGGCGTA	ATCATGGTCA	TAGCTGTTTC	2520
(CTGTGTGAAA	TTGTTATCCG	CTCACAATTC	CACACAACAT	ACGAGCCGGA	AGCATAA	2577

(2) INFORMATION FOR SEQ ID NO:136:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 728 amino acids
 - (B) TYPE: amino acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:136:

Pro Ala Asn Ala Ile Lys Glu Met Ile Glu Asn Cys Leu Asp Ala Lys

Ser Thr Asn Ile Gln Val Val Val Lys Glu Gly Gly Leu Lys Leu Ile 20 25 30

Gln Ile Gln Asp Asn Gly Thr Gly Ile Arg Lys Glu Asp Leu Asp Ile 35 40 45

Val Cys Glu Arg Phe Thr Thr Ser Lys Leu Gln Thr Phe Glu Asp Leu 50 55 60

Ala Ser Ile Ser Thr Tyr Gly Phe Arg Gly Glu His Leu Ala Ser Ile
65 70 75 80

Ser His Val Ala His Val Thr Ile Thr Thr Lys Thr Ala Asp Gly Lys

Cys Ala Tyr Arg Ala Ser Tyr Ser Asp Gly Lys Leu Gln Ala Pro Pro 100 105 110

Lys Pro Cys Ala Gly Asn Gln Gly Thr Leu Ile Thr Val Glu Asp Leu
115 120 125

Phe Tyr Asn Ile Ile Thr Arg Arg Lys Ala Leu Lys Asn Pro Ser Glu

130 135 140

Glu Tyr Gly Lys Ile Leu Glu Val Val Gly Arg Tyr Ser Ile His Asn

Ser Gly Ile Ser Ile Ser Val Lys Lys Gln Gly Glu Thr Val Ser Asp 165 170 175

•	Val	Arg	Thr	Leu 180	Pro	Asn	Ala	Thr	Thr 185	Val	Asp	Asn	Ile	Arg 190	Ser	Ile
1	Phe	Gly	Asn		Val	Ser	Arg			Ile	Glu	Val			Glu	Asp
			195					200					205			
1	Lys	Thr 210	Leu	Ala	Phe	Lys	Met 215	Asn	Gly	Tyr	Ile	Ser 220	Asn	Ala	Lys	Tyr
	202		Tue	Twa	Cura	Ile		Tan	Leu	Dhe	Tla		wie	Ara	Len	Va l
		Val	my a	mys	Cys		rne	neu	neu	riie		non	1110	nrg	Deu	
	225	_			_	230	_		~		235		_			240
(ilu	Ser	Ala	ATA	245	Arg	răs	Ala	11e	250	Thr	vaı	Tyr	Ala	255	туг
1	Leu	Pro	Lys	Thr	His	Thr	His	Ser	Cys	Thr	Ser	Val	Glx	Asn	Gln	Pro
				260					265					270		
5	Ser	Glu	Ara	Asp	Val	Asn	Val	His	Pro	Thr	Lvs	Thr	Glu	Val	His	Phe
			275			•		280			-, -		285			
•		uio		~1	C	T1_	Tour		X ~~~	₹7 . 1	C1-	61 =		T10	C1	60.
1	Jeu		GIU	GIU	ser	Ile		GIII	Arg	vai	GIII		ura	TIE	GIU	Ser
		290					295					300				
	_	Leu	Leu	Gly	Ser	Asn	Ser	Ser	Arg	Met		Phe	His	Pro	Asp	Leu
3	305					310					315					320
Z	lla	Ser	Arg	Thr	Cys	Trp	Ala	Ser	Gly	Glu	Ala	Ala	Arg	Pro	Thr	Thr
					325					330					335	
C	ly	Val	Ala	Ser	Ser	Ser	Thr	Ser	Gly	ser	Gly	Asp	Lys	Val	Tyr	Ala
				340				,	345					350	,	
7	'yr	Gln	Met	Ser	Arg	Thr	Asp	Ser	Arg	Asp	Gln	Lys	Leu	Asp	Ala	Phe
			355					360					365			
I	éu	Gln	Pro	Val	Ser	Ser	Leu	Val	Pro	Ser	Gln	Pro	Gln	qsA	Pro	Arg
		370					375					380		_		_
F	ro	Val	Arq	Gly	Ala	Arg	Thr	Glu	Gly	Ser	P.	Glu	Ara	Ala	Thr	Arc
	885		_	•		390			•		395					400
		Asp	Glu	Glu	Met	Leu	Ala	T.en	Pro	Ala		Ala	Glu	Δla	Δla	
•			014	0.14	405			200		410			014	AIG	415	21.10
_		805	C3	7 ~ ~		~1.v	N == ==	C1	50×		Woh	C1	Mh	C		* 7 ~
G	, Lu	Ser	GIU		rea	Glu	Arg	GIU		Leu	Met	GIU	THE		Asp	Ala
_	_		_	420		_			425					430		
A	la	Gln		Ala	Ala	Pro	Thr		Ser	Pro	Gly	Ser		Arg	Lys	Ser
			435					440					445			
H	lis	Arg	Glu	Asp	Ser	Asp	Val	Glu	Met	Val	Glu	Asn	Ala	Ser	Gly	Lys
		450					455					460				
G	lu	Met	Thr	Ala	Ala	Суз	Tyr	Pro	Arg	Arg	Arg	Ile	Ile	Asn	Leu	Thr
4	65					470					475					480
S	er	Val	Leu	Ser	Leu	Gln	Glu	Glu	Ile	Ser	Glu	Arg	Cys	His	Glu	Thr
					485					490		-			495	
I	,eu	Ara	Glu	Ile		Arg	Asn	His	Ser		Val	G) v	Cvs	Val		Pro
_		7		500		79			505			1	-10	510		
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G	-11	TLD	515		vra	Gln		GIN		тÄд	Leu	rAL	Leu		ASN	ınr
			2012					5 7 [1					476			

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Thr	Lys	Leu	Ser	Glu	Glu	Leu	Phe	Tyr	Gln	Ile	Leu	Ile	Tyr	Asp	Phe
	530					535					540				
Ala	Asn	Phe	Gly	Val	Leu	Arg	Leu	Ser	Glu	Pro	Ala	Pro	Leu	Phe	Asp
545					550					555					560
Leu	Ala	Met	Leu	Ala	Glx	Thr	Val	Leu	Lys	Val	Ala	Gly	Gln	Arg	Thr
				565					570					575	
Thr	Ala	Arq	Arg	Ara	Ala	Cvs	Ara	Val	His	Cvs	Ara	Val	Ser	Glu	Glu
		_	580			•	•	585		•	-		590		
Lvs	Ara	Asp	-	Cvs	Ara	Leu	Phe		Val	Ara	Ser	Met		Ara	Glu
-1-		595		-1-			600			******		605	9	9	
Dro	Aco		Tou	7 011	Dho	C1 w		C1 n	Leu	C	71.	_	Dho	~1	~1
PLO	_	GIX	Leu	Leu	rne		GIX	GIII	Leu	Сув		Int	Pne	GTA	GLY
	610					615					620				
Thr	Ala	Tyr	Leu	His	Ser	Ser	Thr	Gly	His	Glx	Gly	Glu	Leu	Gly	Glu
625					630					635					640
Glu	Lys	Glu	Cys	Phe	Glu	Ser	Leu	Ser	Lys	Glu	Cys	Ala	Met	Phe	Tyr
	•			645					650					655	
Ser	Ile	Arq	Lys	Gln	Tyr	Ile	Leu	Glu	Glu	Ser	Thr	Leu	Ser	Glv	Gln
		_	660		•			665					670	-	
Gln	Ser	Asp	Met	Pro	Glv	Ser	Thr	Ser	Lys	Pro	Trn	T.vs		Thr	٧al
		675			1		680		-,-			685			
G1:1	Wie		T16	Tree	Tue	Als		A ====	Ser	ui a	T 011		D=0	Dro.	T
GIU	690	***	11,6	TAT	ъys	695	FILE	ary	Ser	nrs	700	Leu	PIO	FIO	The
			-•	_				_		_				_	
	Pne	Thr	Glu	Asp		Asn	Val	Leu	Gln		Ala	Asn	Leu	Pro	
705					710					715					720
Leu	Tyr	Lys	Val	Phe	Glu	Arg	Cys								
				725						-					

(2) INFORMATION FOR SEQ ID NO:137:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3065 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:137:

CGGTGAAGGT CCTGAAGAAT TTCCAGATTC CTGAGTATCA TTGGAGGAGA CAGATAACCT GTCGTCAGGT AACGATGGTG TATATGCAAC AGAAATGGGT GTTCCTGGAG ACGCGTCTTT 120 TCCCGAGAGC GGCACCGCAA CTCTCCCGCG GTGACTGTGA CTGGAGGAGT CCTGCATCCA 180 TGGAGCAAAC CGAAGGCGTG AGTACAGAAT GTGCTAAGGC CATCAAGCCT ATTGATGGGA 240 AGTCAGTCCA TCAAATTTGT TCTGGGCAGG TGATACTCAG TTTAAGCACC GCTGTGAAGG 300 AGTTGATAGA AAATAGTGTA GATGCTGGTG CTACTACTAT TGATCTAAGG CTTAAAGACT 360 ATGGGGTGGA CCTCATTGAA GTTTCAGACA ATGGATGTGG GGTAGAAGAA GAAAACTTTG 420 AAGGTCTAGC TCTGAAACAT CACACATCTA AGATTCAAGA GTTTGCCGAC CTCACGCAGG 480 TTGAAACTTT CGGCTTTCGG GGGGAAGCTC TGAGCTCTCT GTGTGCACTA AGTGATGTCA 540 CTATATCTAC CTGCCACGGG TCTGCAAGCG TTGGGACTCG ACTGGTGTTT GACCATAATG 600 GGAAAATCAC CCAGAAAACT CCCTACCCC GACCTAAAGG AACCACAGTC AGTGTGCAGC 660

ACTTATTTTA	TACACTACCC	GTGCGTTACA	AAGAGTTTCA	GAGGAACATT	AAAAAGGAGT	720
ATTCCAAAAT	GGTGCAGGTC	TTACAGGCGT	ACTGTATCAT	CTCAGCAGGC	GTCCGTGTAA	780
GCTGCACTAA	TCAGCTCGGA	CAGGGGAAGC	GGCACGCTGT	GGTGTGCACA	AGCGGCACGT	840
CTGGCATGAA	GGAAAATATC	GGGTCTGTGT	TTGGCCAGAA	GCAGTTGCAA	AGCCTCATTC	900
CTTTTGTTCA	GCTGCCCCCT	AGTGACGCTG	TGTGTGAAGA	GTACGGCCTG	AGCACTTCAG	960
GACGCCACAA	AACCTTTTCT	ACGTTTTCGG	GCTTCATTTC	ACAGTGCACG	CACGGCGCCG	1020
ggaggagtgc	AACAGACAGG	CAGTTTTTCT	TCATCAATCA	GAGGCCCTGT	GACCCAGCAA	1080
aggtctctaa	GCTTGTCAAT	GAGGTTTATC	ACATGTATAA	CCGGCATCAG	TACCCATTTG	1140
TCGTCCTTAA	CGTTTCCGTT	GACTCAGAAT	GTGTGGATAT	TAATGTAACT	CCAGATAAAA	1200
GGCAAATTCT	ACTACAAGAA	GAGAAGCTAT	TGCTGGCCGT	TTTAAAGACC	TCCTTGATAG	1260
GAATGTTTGA	CAGTGATGCA	AACAAGCTTA	ATGTCAACCA	GCAGCCACTG	CTAGATGTTG	1320
AAGGTAACTT	AGTAAAGTCG	CATACTGCAG	AACTAGAAAA	GCCTGTGCCA	GGAAAGCAAG	1380
ATAACTCTCC	TTCACTGAAG	AGCACAGCAG	ACGAGAAAAG	GGTAGCATCC	ATCTCCAGGC	1440
TGAGAGAGGC	CTTTTCTCTT	CATCCTACTA	AAGAGATCAA	GTCTAGGGGT	CCAGAGACTG	1500
CTGAACTGAC	ACGGAGTTTT	CCAAGTGAGA	AAAGGGGCGT	GTTATCCTCT	TATCCTTCAG	1560
ACGTCATCTC	TTACAGAGGC	CTCCGTGGCT	CGCAGGACAA	ATTGGTGAGT	CCCACGGACA	1620
GCCCTGGTGA	CTGTATGGAC	AGAGAGAAAA	TAGAAAAAGA	CTCAGGGCTC	AGCAGCACCT	1680
Cagctggctc	TGAGGAAGAG	TTCAGCACCC	CAGAAGTGGC	CAGTAGCTTT	AGCAGTGACT	1740
ATAACGTGAG	CTCCCTAGAA	GACAGACCTT	CTCAGGAAAC	CATAAACTGT	GGTGACCTGC	1800
IGCCGTCCTC	CAGGTACAGG	ACAGTCCTTG	AAGCCAGAAG	ACCATGGATA	TCAATGCAAA	1860
GCTCTACCTC	TAGCTCGTCT	GTCACCCACA	AATGCCAAGC	GCTTCAAGAC	AGAGGAAGAC	1920
CCTCAAATGT	CAACATATCT	CAAAGATTGC	CTGGTCCTCA	GAGCACCTCA	GCAGCTGAGG	1980
ICGATGTAGC	CATAAAAATG	AATAAGAGAT	CGTGCTCCTC	GAGTTCTCTA	GCTAAGCGAA	2040
IGAAGCAGTT	ACAGCACCTA	AAGGCGCAGA	ACAAACATGA	ACTGAGTTAC	AGAAATTTA	2100
GGGCCAAGAT	TTGCCCTGGA	GAAAACCAAG	CAGCAGAAGA	TGAACTCAGA	Aaagagatta	2160
	GTTTGCAGAG					2220
CCAAACTGAA	AGAGGACCTC	TTCCTGGTGG	ACCAGCATGC	TGCGGATGAG	AAGTACAACT	30
TTGAGATGCT	GCAGCAGCAC	ACGGTGCTCC	AGGCGCAGAG	GCTCATCACG	TGGGTGCACF	340
	AGTTCCCAGA					2400
IGATAGAAAA	TCTGGAAATA	TTCAGAAAGA	ATGGCTTTGA	CTTTGTCATT	GATGAGGATG	2460
CTCCAGTCAC	TGAAAGGGCT	AAATTGATTT	CCTTACCAAC	TAGTAAAAAC	TGGACCTTTG	2520
FACCCCAAGA	TATAGATGAA	CTGATCTTTA	TGTTAAGTGA	CAGCCCTGGG	GTCATGTGCC	2580
GCCCTCACG	AGTCAGACAG	ATGTTTGCTT	CCAGAGCCTG	TCGGAAGTCA	GTGATGATTG	2640
SAACGGCGCT	CAATGCGAGC	GAGATGAAGA	AGCTCATCAC	CCACATGGGT	GAGATGGACC	2700
ACCCCTGGAA	CTGCCCCCAC	GGCAGGCCAA	CCATGAGGCA	CGTTGCCAAT	CTGGATGTCA	2760
CTCTCAGAA	CTGACACACC	CCTTGTAGCA	TAGAGTTTAT	TACAGATTGT	TCGGTTCGCA	2820
AGAGAAGGT	TTTAAGTAAT	CTGATTATCG	TTGTACAAAA	ATTAGCATGC	TGCTTTAATG	2880
	ATTTAAAAGC					2940
	GATCCGGTGG					3000
CATTCATGA	GACTCAATTC	AAGGACAAAA	AAAAAAAGAT	ATTTTTGAAG	CCTTTTAAAA	3060
AAAA						2065

(2)	INFO	RMAT	ION :	FOR	SEQ	ID N	0:13	8:								
	(i)	SEQ	UENC	E CH	ARAC'	TERI	STIC	S:								
		(A) LE	ngth	: 86	4 am:	ino a	acid	8							
		(B) TY	PE:	amin	o ac	id									
		•	•			5S: 1		le								
		•	•			line										
	(ii)					•										
	(xi)											-1-	-		- 1 -	•
	net 1	GIU	GIN	THE	5	GIÀ	var	ser	THE	10	Сув	Ala	Lys	Ala	11e	гÃ
		Tle	Agn	Glw		Ser	17 a 1	uia	Gla		Cua	Sar	Gly	Gln		T 3 4
				20	27.0	001	V 44.	*****	25		Cyb	Der	GLY	30	*41	**
	Leu	Ser	Leu		Thr	Ala	Val	Lvs		Leu	Ile	Glu	Asn		Val	Ası
			35					40					45			
	Ala	Gly	Ala	Thr	Thr	Ile	Asp	Leu	Arg	Leu	Lys	Asp	Tyr	Gly	Val	Ası
		50					55		_		_	60	_	_		-
	Leu	Ile	Glu	Val	Ser	Asp	Asn	Gly	Суз	Gly	Val	Glu	Glu	Glu	Asn	Phe
	65					70					75					80
	Glu	Gly	Leu	Ala	Leu	Lys	His	His	Thir	Ser	Lys	Ile	Gln	Glu	Phe	Ala
					85					90					95	
	Asp	Leu	Thr	•	Val	Glu	Thr	Phe	_	Phe	Arg	Gly	Glu			Sea
		*	a	100					105		_		_	110	•	_
	ser	Leu	115	Ala	ren	Ser	Asp	120	Thr	lle	Ser	Thr	Сув	His	Gly	Sei
	Ala	Ser		Glv	Thr	Ara	T.ou		Pho	Acn	Wie	A a n	125 Gly	T ***	Tle	Thi
		130	***	01,	****	mry,	135	V 0.1	F 1.1C	veb	1115	140	GIY	гур	116	1111
	Gln	Lys	Thr	Pro	Tyr	Pro	Arg	Pro	Lys	Gly	Thr		Val	Ser	Val	Glr
	145	_			_	150	-		-	•	155					160
	His	Leu	Phe	Tyr	Thr	Leu	Pro	Val	Arg	Tyr	Lys	Glu	Phe	Gln	Arg	Ası
					165					170					175	
	Ile	Lys	Lys		Tyr	Ser	Lys	Met	Val	Gln	Val	Leu	Gln	Ala	Tyr	Cyt
			_	180			_		185					190		
	IIe	IIe	Ser 195	Ala	GIA	Val	Arg		Ser	Cys	Thr	Asn	Gln	Leu	Gly	Glı
	Glv	Luc		uia	λla	Wa l	Wa 1	200	Th.∞	50×	C1	mh sa	205 Ser	G1	Wot	T
	O.J	210	my	******	nia	Val	215	Cys	1111	Ser	GIY	220	ser	GIY	met	гЪ
	Glu		Ile	Glv	Ser	Val		Glv	Gln	Lvs	Gln		Gln	Ser	T.eu	Tle
	225			-		230		•			235					240
	Pro	Phe	Val	Gln	Leu	Pro	Pro	Ser	Asp	Ala		Cys	Glu	Glu	Tyr	Gly
					245					250		_			255	
	Leu	Ser	Thr	Ser	Gly	Arg	His	ГЛЗ	Thr	Phe	Ser	Thr	Phe	Ser	Gly	Phe
				260					265					270		
	Ile	Ser	Gln	Cys	Thr	His	Gly	Ala	Gly	Arg	Ser	Ala	Thr	Asp	Arg	Gl
			275					280					285			

Phe		Phe	Ile	Asn	Gln	_	Pro	Сув	Asp	Pro		Lys	Val	Ser	Lys
	290					295					300				
Leu 305	Val	Asn	Glu	Val	Tyr 310	His	Met	Tyr	Asn	Arg 315	His	Gln	Tyr	Pro	320
٧al	Val	Leu	Asn	Val	Ser	Val	Asp	Ser	Glu	Сув	Val	Asp	Ile	Asn	Val
				325					330			_		335	
Thr	Pro	Asp	Lys	Arg	Gln	Ile	Leu	Leu	Gln	Glu	Glu	Fåa	Leu	Leu	Lev
			340					345					350		
Ala	Val	Leu	Lys	Thr	Ser	Leu	Ile	Gly	Met	Phe	Asp	Ser	Asp	Ala	Asn
		355					360					365			
Lys	Leu	Asn	Val	Asn	Gln		Pro	Leu	Leu	Asp	Val	Glu	Gly	Asn	Leu
	370					375					380				
Val	Lys	Ser	His	Thr	Ala	Glu	Leu	Glu	Lys	Pro	Val	Pro	Gly	Lys	Gln
385					390					395					400
Asp	Asn	Ser	Pro	Ser	Leu	Lys	Ser	Thr	Ala	Asp	Glu	Lys	Arg	Val	Ala
				405					410					415	
Ser	Ile	Ser	Arg	Leu	Arg	Glu	Ala		Ser	Leu	His	Pro	Thr	Lys	Glu
			420					425					430		
Ile	Lys		Arg	Gly	Pro	Glu		Ala	Glu	Leu	Thr		Ser	Phe	Pro
		435					440					445			
Ser		Lys	Arg	Gly	Val	_	Ser	Ser	Tyr	Pro		Asp	Val	Ile	Ser
_	450					455					460				
Tyr 465	Arg	Gly	Leu	Arg	Gly 470	Ser	Gln	Asp	Lys	Leu 475	Val	Ser	Pro	Thr	Asp 480
,	Pro	Glv	Agn	Cva	Met	Agn	Ara	Glu	T.va		G1.,	Larg	Aen	Sar	
		1		485			9		490		Jau	Ly 5	nop	495	U±y
Leu	Ser	Ser	Thr	Ser	Ala	Gly	Ser	Glu	Glu	Glu	Phe	Ser	Thr	Pro	Glu
			500					505					510		
Val	Ala	Ser	Ser	Phe	Ser	Ser	Asp	Tyr	Asn	Val	Ser	Ser	Leu	Glu	Asp
	-	515					520					525			
Arg	Pro	Ser	Gln	Glu	Thr	Ile	Asn	Cys	Gly	Asp	Leu	Leu	Pro	Ser	Ser
	530					535					540				
Arg	Tyr	Arg	Thr	Val	Leu	Glu	Ala	Arg	Arg	Pro	Trp	Ile	Ser	Met	Gln
545					550					555					560
Ser	ser	Thr	Ser	Ser	Ser	Ser	Val	Thr	His	Lys	Cys	Gln	Ala	Leu	Gln
				565					570					575	
Asp	Arg	Gly	Arg	Pro	Ser	Asn	Val	Asn	Ile	Ser	Gln	Arg	Leu	Pro	Gly
			580					585					590		
Pro	Gln	Ser	Thr	Ser	Ala	Ala	Glu	Val	Asp	Val	Ala	Ile	Lys	Met	Asn
		595					600					605			
Lys	Arg	Ser	Cys	Ser	Ser		Ser	Leu	Ala	Lys	Arg	Met	Lys	Gln	Leu
	610					615					620				
	His	Leu	Lys	Ala	Gln	Asn	Lys	His			Ser	Tyr	Arg	Lys	Phe
625					630					635					640

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Arg	Ala	Lys	Ile	Cys	Pro	Gly	Glu	Asn	Gln	Ala	Ala	Glu	Asp	Glu	Leu
				645					650					655	
Arg	Lys	Glu	Ile	ser	Lys	Ser	Met	Phe	Ala	Glu	Met	Glu	Ile	Leu	Gly
			660					665					670		
Gln	Phe	Asn	Leu	Gly	Phe	Ile	Val	Thr	Lys	Leu	Lys	Glu	Asp	Leu	Phe
		675					680					685			
Leu		Asp	Gln	His	Ala		Asp	Glu	Lys	Tyr		Phe	Glu	Met	Leu
	690					695					700				
Gln	Gln	His	Thr	Val	Leu	Gln	Ala	Gln	Arg	Leu	Ile	Thr	Trp	Val	His
705					710					715					720
Thr	Gly	Phe	Arg	Val	Pro	Arg	Pro	Gln	Thr	Leu	Asn	Leu	Thr	Ala	Val
				725					730		-			735	
Asn	Glu	Ala	Val	Leu	Ile	Glu	Asn	Leu	Glu	Ile	Phe	Arg	Lys	Asn	Gly
			740					745					750		
Phe	Asp	Phe	Val	Ile	Asp	Glu	Asp	Ala	Pro	Val	Thr	Glu	Arg	Ala	Lys
		755					760					765			
Leu	Ile	Ser	Leu	Pro	Thr	Ser	Lys	Asn	Trp	Thr	Phe	Gly	Pro	Gln	Asp
	770					775					780				
Ile	Asp	Glu	Leu	Ile	Phe	Met	Leu	Ser	Asp	Ser	Pro	Gly	Val	Met	Сув
785					790					795					800
Arg	Pro	Ser	Arg	Val	Arg	Gln	Met	Phe	Ala	Ser	Arg	Ala	Сув	Arg	Lys
				805	÷				810					815	
Ser	Val	Met	Ile	Gly	Thr	Ala	Leu	Asn	Ala	Ser	Glu	Met	Lys	Lys	Leu
			820					825					830		
Ile	Thr	His	Met	Gly	Glu	Met	qaA	His	Pro	Trp	Asn	Cys	Pro	His	Gly
		835					840					845			
Arg	Pro	Thr	Met	Arg	His	Val	Ala	Asn	Leu	Asp	Val	Ile	Ser	Gln	Asn
	850					855					860				

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- (2) INFORMATION FOR SEQ ID NO:139:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:139: CTTGATTCTA GAGCYTCNCC NCKRAANCC

(2) INFORMATION FOR SEQ ID NO:140:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:140: AGGTCGGAGC TCAARGARYT NGTNGANAA	29
MOGICOOMSC TOTALINITE MOTHERINE.	-
(2) INFORMATION FOR SEQ ID NO:141:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 15 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:141:	
ACTTGTGGAT TTTGC	15
(2) INFORMATION FOR SEQ ID NO:142:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 15 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:142:	
ACTTGTGAAT TTTGC	15
•	
(2) INFORMATION FOR SEQ ID NO:143:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 22 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:143:	
TTCGGTGACA GATTTGTAAA TG	22
(2) INFORMATION FOR SEQ ID NO:144:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 16 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:144: TTTACGGAGC CCTGGC	16
TITACGGAGC CCIGGC	70

(2) INFORMATION FOR SEQ ID NO:145:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 22 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:145:	
TCACCATAAA AATAGTTTCC CG	22
(2) INFORMATION FOR SEQ ID NO:146:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 22 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:146:	
TCCTGGATCA TATTTCTGA GC	22
(2) INFORMATION FOR SEQ ID NO:147:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 22 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
· (D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:147:	
TTTCAGGTAT GTCCTGTTAC CC	22
(2) INFORMATION FOR SEQ ID NO:148:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 22 base pairs	
(B) TYPE: nucleic acid	
(C) STRANDEDNESS: single	
(D) TOPOLOGY: linear	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:148:	
TGAGGCAGCT TTTAAGAAAC TC	22

WE CLAIM:

- 1. A method of diagnosing cancer susceptibility in a subject comprising detecting a mutation in a *mutL* homolog gene or gene product in a tissue of the subject, the mutation being indicative of the subject's susceptibility to cancer.
- 2. A method of identifying and classifying a DNA mismatch-repair-defective tumor comprising detecting in a tumor a mutation in a *mutL* homolog gene or gene product, the mutation being indicative of a defect in a mismatch repair system of the tumor.
- 3. The method of claim 1 or claim 2 wherein the step of detecting comprises detecting a mutation in hMLH1 or hPMS1.
- 4. The method of claim 1 or claim 2 wherein the step of detecting mprises isolating nucleic acid from the subject;

amplifying a segment of the mismatch repair gene or gene product from the isolated nucleic acid;

comparing the amplified segment with an analogous segment of a wild-type allele of the mismatch repair gene or gene product; and

detecting a difference between the amplified segment and the analogous segment, the difference being indicative of a mutation in the mismatch repair gene or gene product.

5. The method of claim 4 wherein the step of detecting comprises determining whether the difference between the amplified segment and the analogous segment causes an affected phenotype.

- 6. The method of claim 4 wherein the difference in nucleotide sequence is selected from the group consisting of deletions of at least one nucleotide, insertions of at least one nucleotide, substitutions of at least one nucleotide and nucleotide rearrangements.
- 7. The method of claim 4 wherein the step of amplifying comprises:

reverse transcribing all or a portion of an RNA mismatch repair gene product to DNA; and

amplifying a segment of the DNA produced by reverse transcription.

8. The method of claim 4 wherein the step of amplifying comprises:

selecting a pair of oligonucleotide primers capable of hybridizing to opposite strands of the mismatch repair gene, and in opposite orientation;

performing a polymerase chain reaction utilizing the oligonucleotide primers such that nucleic acid of the mismatch repair chain intervening between the primers is amplified to become the amplified segment.

- 9. The method of claim 8 wherein the intervening nucleic acid comprises at least a fragment of at least one exon of the mismatch repair gene.
- 10. The method of claim 9 wherein the at least one exon has a nucleotide sequence selected from the group consisting of SEQ ID NOS: 25-43.

- 11. The method of claim 1 or claim 2 wherein the step of detecting comprises detecting a mutation in a *mutL* homolog mismatch repair protein.
- 12. The method of claim 4 wherein the analogous segment of a wild-type allele of the mismatch repair gene or gene product comprises a wild-type hMLH1 gene fragment having a unique portion of nucleotide sequence selected from the group consisting of: SEQ ID NOS: 6-24.
- 13. The method of claim 8 wherein the step of selecting comprises selecting a pair of oligonucleotide primers, each primer of the pair comprising a nucleotide sequence selected from the group consisting of: SEQ ID NOS: 44-82.
- 14. The method of claim 8 wherein the intervening nucleotide sequence that is amplified comprises a unique portion of at least one nucleotide sequence selected from the group consisting of: SEQ ID NOS: 6-24.
- 15. The method of claim 4 wherein the step of detecting a difference comprises detecting an hMLH1 mutation characterized by a C to T transition mutation which produces a non-conservative amino acid substitution at position 44 of the hMLH1 protein.

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16. The method of claim 5 wherein the step of determining comprises:

deriving a yeast strain that is deleted for its hMLH1 gene; constructing a yeast homolog of the amplified segment including the

difference;

introducing the yeast homolog of the amplified segment into the yeast strain; and

assaying the yeast strains ability to correct DNA mispairs.

- 17. The method of claim 5 wherein the step of determining comprises producing an hMLH1 protein including amino acids corresponding to the difference; and determining the extent of interaction between the hMLH1 protein and an hPMS1 protein compared to the degree of protein-protein interaction observed with wild-type hMLH1 and hPMS1 proteins.
- 18. An isolated oligonucleotide primer capable of hybridizing specifically to all or a fragment of an hMLH1 genomic sequence with a T_m of greater than about 55-degrees° C_o .
- 19. The isolated oligonucleotide primer of claim 18, the oligonucleotide primer being extendable by a DNA polymerase.
- 20. The isolated oligonucleotide primer of claim 19, the oligonucleotide primer being capable of amplifying at least a portion of an *hMLH1* gene when used in a polymerase chain reaction including another primer.

- 21. The isolated oligonucleotide primer of claim 20, the oligonucleotide primer being at least 13 nucleotides in length.
- 22. The isolated oligonucleotide primer of claim 21 comprising a nucleotide sequence selected from the group consisting of SEQ ID NOS: 44-82.
- 23. An isolated nucleic acid including a segment having a nucleotide sequence substantially identical to a nucleotide sequence selected from the group consisting of SEQ ID NOS: 6-24.
- 24. An isolated nucleic acid including a segment having a nucleotide sequence substantially identical to a nucleotide sequence selected from the group consisting of SEQ ID NOS: 25-43.
- 25. A unique fragment of the nucleic acid of claim 23 or claim 24.
- 26. A method of detecting a mutation in a eukaryotic *mutL* homolog gene or fragment thereof comprising the steps of:

isolating a eukaryotic *mutL* homolog gene or fragment thereof; and detecting a difference in activity between the isolated gene or fragment thereof and a wild-type allele of the gene or fragment thereof; the difference in activity being indicative of a mutation in the eukaryotic *mutL* homolog gene or fragment thereof.

- 27. A method of detecting a mutation in a eukaryotic *mutL* homolog gene or gene product comprising detecting a difference in activity between the gene or gene product and a wild-type version of the gene or gene product, the difference in activity being indicative of a mutation in the *mutL* homolog gene or gene product.
- 28. The method of claim 26 wherein the eukaryotic *mutL* homolog gene or fragment thereof comprises a human gene or fragment thereof.
- 29. The method of claim 27 wherein the *mutL* homolog gene or gene product comprises a human gene or gene product.
- · 30. The method of claim 28 or claim 29 wherein the gene comprises an *hMLH1* and the wild-type version of the gene comprises a wild-type allele of the *hMLH1* gene.
- 31. The method of claim 28 or claim 29 wherein the gene comprises a *hPMS1* and the wild-type version of the gene comprises a wild-type allele of the *hPMS1* gene.
- 32. The method of claim 30 wherein the wild-type version of the hMLH1 gene comprises a nucleotide sequence substantially identical to a nucleotide sequence selected from the group consisting of SEQ ID NOS: 6-24, and unique fragments thereof.

- 33. The method of claim 30 wherein the wild-type version of the *hMLH1* gene encodes a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO: 5 and unique fragments thereof.
- 34. The method of claim 28 or claim 29 wherein the human mismatch repair gene product comprises a hMLH1 protein or unique fragment thereof.
- 35. The method of claim 34 wherein the hMLH1 protein comprises an amino acid sequence selected from the group consisting of SEQ ID NO: 5 and unique fragments thereof.
- 36. An isolated nucleotide or protein structure including a segment sequentially corresponding to a unique portion of a human *mutL* homolog gene or gene product.
- 37. The nucleotide of claim 36 wherein the *mutL* homolog gene is *hMLH1* or *hPMS1*.
- 38. A pair of oligonucleotide primers capable of being used together in a polymerase chain reaction to amplify specifically a unique segment of a human *mutL* homolog gene.
- 39. The pair of oligonucleotide primers of claim 38 wherein the *mutL* homolog gene is *hMLH1* or *hPMS1*.

- 40. A probe comprising
- a nucleotide sequence capable of binding specifically by Watson/Crick pairing to complementary bases in a portion of a human *mutL* homolog gene; and
- a label-moiety attached to the sequence, wherein the label-moiety has a property selected from the group consisting of fluorescent, radioactive and chemiluminescent.
- 41. The probe of claim 40 wherein the human *mutL* homolog gene is *hMLH1* or *hPMS1*.
- 42. An amplified quantity of a nucleotide including a segment corresponding to a unique portion of a human *mutL* homolog gene.
- 43. The nucleotide of claim 42 wherein the human mutL he molog gene is hMLH1 or hPMS1.
- 44. A pair of oligonucleotide primers capable of being employed in a polymerase chain reaction to amplify specifically a single exon from a human *mutL* homolog gene along with selected portions of flanking upstream and downstream introns.
- 45. The primers of claim 44 wherein the human *mutL* homolog gene is *hMLH1* or *hPMS1*.

- 46. The method of claim 1 wherein the detecting step comprises detecting a mutation in a portion of the individual's *hMLH1* gene, the portion being homologous to the DNA sequence including and between the two sets of underlined bases in Figure 3.
- 47. The nucleotide of claim 37 wherein the segment is homologous to the DNA sequence including and between the two sets of underlined bases in Figure 3.
- 48. An isolated nucleotide or protein structure including a segment substantially corresponding to a unique portion of a mouse *mutL* homolog gene or gene product.
- 49. The structure of claim 48 wherein the segment substantially corresponds to a unique portion of a mammalian MLH1 or PMS1 gene or protein.
- 50. Purified antibodies binding specifically to a MutL homolog protein.
- 51. The antibodies of claim 50 wherein the antibodies are monoclonal antibodies.
- 52. The antibodies of claim 50 wherein the MutL homolog protein is a human protein.

- 53. The antibodies of claim 52 wherein the protein is hMLH1 or hPMS1.
- 54. The antibodies of claim 50 wherein the MutL homolog protein is a mouse protein.
- 55. The antibodies of claim 54 wherein the protein is mMLH1 or mPMS1.

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Guide for the isolation and characterization of mammalian PMSI and MLH1 genes.

- Step 1 Design of degenerate oligonucleotide pools for PCR.
- Step 2 Reverse transcription and PCR on poly A+ selected mRNA isolated from human cells.
- Step 3 Cloning and sequencing of PCR generated fragments; identification of two gene fragments representing human *PSM1* and *MLH1*.
- Step 4 Isolation of complete human and mouse PMS1 and MLH1 cDNA clones using the PCR fragments as probes.
- Step 5 Isolation of human and mouse, PMS1 and MLH1 genomic clones.
- Step 6 Chromosome positional mapping of the human and mouse, *PMSI* and *MLH1* genes by fluorescence *in situ* hybridization.
- Step 7 Using genomic and cDNA sequences to identify mutations in *PMS1* and *MLH1* genes from HNPCC Families.
- Step 8 Design targeting vectors to disrupt mouse *PMS1* and *MLH1* genes in ES cells; study mice deficient in mismatch repair.

Figure 1
SUBSTITUTE SHEET (RULE 26)

2/24 1 10 20
HPIQVLPPQLANQIAGEVVERPASVVK - SEQ. ID NO: 1 Mut I. MSHIIELPEMLANCIAAGEVIERPASVCK - SEQ. ID NO: 2 HexB HFHHIENLLIETEKRCKOKEORYIPVKYLFSHTOIHOIHDIDVHRITSGOVITDLTTAUK - SEQ. ID NO: 3 Pmel 30 40 60 ELVENSLDAGATRVDIDIERGCAKLIRIRDNGCGIKKEELALALARHATSKIASLDDLEA MutL ELVENAIDAGSSQIIIEIEEAGLKKVQITDNGHGIAHDEVELALRRHATSKIKNQADLFR <u>ELVDNSIDANANQIEIIFKDYGLESIECSDNGDGIDPSHYEFLALKHYTSKIAKFQDVAK</u> 90 100 80 70 110 120 110 IISLGFRGEALASISSVSRLTLTSRTAEQAEAWQAYAEGRDHDVTVKPAAHPVGTTLEVL MutL IRTLGFRGEALPSIASVSVLTLLTAVDGASHGTKLVARGGEVE.EVIPATSPVGTKVCVE VOTLGFRGEALSSLCGIAKLSVITTTSPPKADKELYDHVGHIT.SKTTTSRNKGTTVLVS 140 150 190 170 DLFYNTFARRK. FMRTEKTEFNHIDEIIRRIALARFDVTLNISHNGKLVRQYRAVAKDCQ DLFFNTPARLK.YHKSQQAELSHIIDIVHRLGLAHPEISFSLISDGKENTR...TAGTGQ QLFHNLPVRQKEFSKTFKRQFTKCLTVIQGYAIINAAIKFSVWNITPKGKKHLILSTWRN 240 250 KERRLGAICGTPFLEOALAIEWOHGDLTLRGWVADPNHTTTALTELQYCYVNGRHHRDRL LRQAIAGIYGLVSAKKHIEIENSDLDFEISGFVSLPELTRANRNYISL.FINGRYIKNFL SSHRKN.ISSVFGAGGHRGELEVOLVLDLNPFKNRHLGKYTDDPDFLDLDYKIRVKGYIS 260 270 280 INHAIRQACEDKLGA......DQQPAFVLYLEIDPHQVDV MutL LNRAILDGFGSKLMV..... .GRFPLAVIHIHIDPYLADV HaxB QNSFGCGRNSKDRQFIYVNKRPVEYSTLLKCCNEVYKTFNN «QFPAVFLNLELPHSLIDV Pms1 310 320 330 340 350 320 330 340 NVHPAKHEVRFHQSRLVHDFIXQGVLSVLQQQTETALPLEEIAPAPRHVQEHRIAAGRNH **NVHPTKQEVRISKEKELMTLVSEAIANSLKEQTLIPDALENLAKSTVRNREKVEQTILPL** NVTPDKRVILLMREAVID.IFKTTLSDYYNROELALPKRMCSQSEQQAQKRLKTEVFDD 390 460 480 470 SFPELEFFGQHHGTYLFA....QGRDGLYIIDQHAAQERVKYEEYRESIGNVDQSQQQLL HexB DFKKMEVVGQFNLGFIIVTRKVDNKSDLFIVDQHASDEKYNFETLQAVTVF...KSQKLI 710 720 730 740 520 530 VPYIFEFPADDALRLKERMPLLEEVGVFLAEYGENQFILREHPIWHAEEEIESGIYEHCD IPOPVELSVIDELVVLONLPVFEKNGFKLKIDEEEEFGSRVKLLSLPTSKQTLFDLGDFN 780 790 760 770 800 580 590 600 Hexb HLLLTKEVSIKKYRAELA.....IMMSCKRSIKANHRIDDHSARQLLYQLSQCDNPY ELIHLIKEDGGLRRDNIRCSKIRSHFAMRACRSSINIGKPLNKKTMTRVVHNLSELDKPW 830 850 860 840 .870 629 NCPHGRPVLVHFT HexB Figure 2 NCPHGRPTMRHLM

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```
60 — SEQ. ID NO: 4
 CTTGGCTCTTCTGGCGCCAAAATGTCGTTCGTGGCAGGGGTTATTCGGCGGCTGGACGAG
                                                                        -SEO. ID NO: 5
 acagtggtgaaccgcatcgcggcggggaagttatccagcggccagctaatgctatc<u>aaa</u>
 <u>GAGATGATTGAGAAC</u>TGTTTAGATGCAAAATCCACAAGTATTCAAGTGATTGTTAAAGAG
                   LDAKSTSI
 GGAGGCCTGAAGTTGATTCAGATCCAAGACAATGGCACCGGGATCAGGAAAGAAGATCTG
G G L K L I Q I Q D N G T G I R K E D L
GATATTGTATGTGAAAGGTTCACTACTAGTAAACTGCAGTCCTTTGAGGATTTAGCCAGT
D I V C E R F T T S K L Q S F E D L A S
actattacaacgaaaacagctgatggaaagtgtgcatacagagcaagtt<mark>actcagatgga</mark>
AAACTGAAAGCCCCTCCTAAACCATGTGCTGGCAATCAAGGGACCCAGATCACGGTGGAG
GACCTTTTTACAACATAGCCACGAGGAGAAAAGCTTTAAAAAATCCAAGTGAAGAATAT
D L F Y N I A T R R K A L K N P S E E Y
GGGAAAATTTTGGAAGTTGTTGGCAGGTATTCAGTACACAATGCAGGCATTAGTTTCTCA
G K I L E V V G R Y S V H N A G I S F S
GTTAAAAAACAAGGAGAGACAGTAGCTGATGTTAGGACACTACCCAATGCCTCAACCGTG
GACAATATTCGCTCCATCTTTGGAAATGCTGTTAGTCGAGAACTGATAGAAATTGGATGT
GAGGATAAAACCCTAGCCTTCAAAATGAATGGTTACATATCCAATGCAAACTACTCAGTG
E D K T L A F K M N G Y I S N A N Y S V AAGAAGTGCATCTTCTTCATCATCACCATCGTCTGGTAGAATCAACTTCCTTGAGA
CTCAGTTTAGAAATCAGTCCCCAGAATGTGGATGTTAATGTGCACCCCACAAAGCATGAA
                     O N
                                     N
GTTCACTTCCTGCACGAGGAGCATCCTGGAGCGGGTGCAGCAGCACATCGAGAGCAAG
                                                                              Human MLH1 cDNA
CTCCTGGGCTCCAATTCCTCCAGGATGTACTTCACCCAGACTTTGCTACCAGGACTTGCT
                                                                              Nucleotide and
GGCCCCTCTGGGGAGATGGTTAAATCCACAACAAGTCTGACCTCGTCTTCTACTTCTGGA
                                                                              Protein Sequence
AGTAGTGATAAGGTCTATGCCCACCAGATGGTTCGTACAGATTCCCGGGAACAGAAGCTT
GAGGATAAGACAGATATTTCTAGTGGCAGGCTAGGCAGCAAGATGAGGAGATGCTTGAA
                                                                1320
CTCCCAGCCCTGCTGAAGTGGCTGCCAAAAATCAGAGCTTGGAGGGGGATACAACAAAG
L P A P A E V A A K N Q S L E G D T T K
GGGACTTCAGAAATGTCAGGAGAGGAGGACCTACTTCCAGCAACCCCAGAAAGAGACAC
G T S E M S E K B G B T T C
G T S E M S E K R G P T S S N P R K R E CGGGAAGATTCTGATGTGGAAATGGTGGAAGATGATTCCCGAAAGGAAATGACTGCAGCT
R E D S D V E M V E D D S R K E M T A A
TGTACCCCCGGAGAAGC CATTAACCTCACTAGTGTTTTGAGTCTCCAGGAAGAAATT
T P R R R I I N L T S V L S L Q E E I ANTGAGCAGGGACATG GGTCTCCGGGGGAGATGTTGCATAACCACTCCTTCGTGGGCTGT N E Q G H E V L R E M I V N GCCCTTCGTGGGCTGT
GTGAATCCTCAGTGGGCCTTGGCACAGCATCAAACCAAGTTATACCTTCTCAACACCACC
AAGCTTAGTGAAGAACTGTTCTACCAGATACTCATTTATGATTTTGCCAATTTTGGTGTT
CTCAGGTTATCGGAGCCAGCACCGCTCTTTGACCTTGCCATGCTTGCCTTAGATAGTCCA
                                                                 1800
                                                                                  Figure 3
Gagagtggctggacagaggaagatggtcccaaagaaggacttgctgaatacattgttgag
TTTCTGAAGAAGAAGGCTGAGATGCTTGCAGACTATTTCTCTTTGGAAATTGATGAGGAA
GGGAACCTGATTGGATTACCCCTTCTGATTGACAACTATGTGCCCCCTTTGGAGGGACTG
GAAAGCCTCAGTAAAGAATGCGCTATGTTCTATTCCATCCGGAAGCAGTACATATCTGAG
GAGTCGACCCTCTCAGGCCAGCAGAGTGAAGTGCCTGGCTCCATTCCAAACTCCTGGAAG
E S T L S G Q Q S E V P G S I P N S W K
TGGACTGTGGAACACATTGTCTATAAAGCCTTGCGCTCACACATTCTGCCTCCTAAACAT
F T E D G N I L Q L A N L P D L Y K V F GAGAGGTGTTAAATATGGTTATTTATGCACTGTGGGATGTTCTTCTTCTCTGTATTC
2400
AATAAATAGATGTGTCTTAACATA
```

#1: 18442 to 19109 (-21 to 116)

TGGCTGGATGCTAAGCTACAGCTGAAGGAAGAACGTGAGCACGaggcactgagt gattggcTGAAGGCACTTCCGTTGAGCATCTAGACGTTTCcttggctcttctggc gccaaaatgtcgttcqtggcaggggttattcggcggctggacgagagagtggtga accgcatcgcggcgggggaagttatccagcggccagctaatgctatcaaagagat gattgagaactgGTACGGAGGGAGTCGAGCCGGgctcacttaagggctacqaCTT AACGGGCCGCGTCACTCAATGGCGCGGACACGCCTCTTTCCCCGGGCAGAGGCAT GTACAGCGCATGCCCACAACGGCGGAGGCCGCGGGTTCCCTACGTGCCATAAGC CTTCTCCTTTTC

SEQ. ID NO: 6

SEQ. ID NO: 25

#2: 19689 to 19688 (117 to 207)

AAACACGTTAATGAGGCACTATTGTTTGTATTTGGAGTTTGTTATCATTGCTTGG
CTCATATTAAaatatgtacattaqaqtaqttqCAGACTGATAAATTATTTTCTGT
TTGATTTGCCAGtttagatgcaaaatccacaagtattcaagtgattgttaaagag
gqaggcctgaagttgattcagatccaagacaatggcaccgggatcaggGTAAGTA
AAACCTCAAAGTAGCAGGATGTTTGTGCGCTTCATGGAAgagtcaggacctttct
ctqTTCTGGAAACTAGGCTTTTGCAGATGGGATTTTTTCACTGAAAAATTCAACA
CCAACAATAAATATTTATTGAGTACCTATTATTTGCGGGGCACTGTTCAGGGGAT
GTGTCAGT

SEQ. ID

SEQ. ID NO: 26

#3: 19687 to 19786 (208 to 306)

SEQ. ID NO: 8

SEQ. ID NO: 27

#4 18492 to 18421 (307 to 380)

TGGAAGCAGCAGNCAGATaacctttccctttggtgaggTGACAGTGGGTGACCCA GCAGTGAGTTTTTCTTTCAGTCTATTTTCTTTCTTCCTTAGGctttggccagca taagccatgtggctcatgttactactaccaacgaaaacagctgatggaaagtgtgc atacagGTATAGTGCTGACTTCTTTTACTCATATATATTCATTCTGAAATGTATT TTGGgcctaggtctcagagtaatcCTGTCTCAACACCAGTGTTATCTTTNNNGGC AGAGATCTTGAGTACG

SEQ. ID

NO: 9

SEQ. ID NO: 28

Figure 4A - 1

#5: 18313 to 18179 (381 to 453)

TTGATATgatttctcttttcccttqqqATTAGTATCTATCTCTCTACTGGATA
TTAATTTGTTATATTTTCTCATTAGagcaagttactcagatggaaactgaaagc
cctcctaaaccatgtgctggcaatcaagggacccagatcacgGTAAGAATGGTA
CATGGGAGAgtaaattgttgaagctttgtttqTATAAATATTGGAATAAAAATA
AAATTGCTTCTAAGTTTTCAGGGTAATAATAAAATGAATTTGCACTAGTTAATGG
AGGTCCCAAGATATCCTCTAAGCAAGATAAATGACTATTGGCTTTTNNTGGCATG
GCAGCCTG

SEQ. ID NO: 10

SEQ. ID NO: 29

#6: 18318 to 18317 (454 to 545)

GCTTTTGCCAGGACCATCTTggqttttattttcaaqtacttctatqAATTTACAA GAAAAATCAATCTTCTGTTCAGgtggaggaccttttttacaacatagccacqagg agaaaagctttaaaaaatccaagtgaagaatatgggaaaattttggaagttgttg gcagGTACAGTCCAAAATCTGGGAGTGGGTCTCTGAGATTTGTCATCAAAGTAAT GTGTTCTAGTgctcatacattgaacagttgctgagcTAGATGGTGAAAAGTAAAA

SEQ. ID NO: 11

SEQ. ID NO: 30

#7: 19009 to 19135 (546 TO 588)

SEQ. ID

SEQ. ID NO: 31

#8: 18197 to 18924 (589 TO 677)

ATGTTTCAGTctcaqccatgaqacaataaatccTTGTGTCTTCTGCTGTTTGTTT
ATCAGcaaggaqaqacagtagctgatgttaggacactacccaatgcctcaaccgt
ggacaatattcgctccatctttggaaatgctgttagtcgGTATGTCGATAACCTA
TATAAAAAAATCTTTTACATTTATTATCTTGGTTTATCATT<u>ccatcacattattt
gggaacc</u>TTTCAAGATATTATGTGTGTTAAGAGTTTGCTTTAGTCAAATACACAG
GCTTGTTTTATGCTTCAGATTTGTTAATGGAGTTCTTATTTCACGTAATCAACAC
TTTCTAGGTGTATGTAATCTCCTAGATTCTGTGGCGTGAATCATGTGTTCT

SEQ. ID

NO: 13

SEQ. ID NO: 32

Figure 4A - 2

#9: 18765 to 18198 (678 TO 790)

SEQ. ID NO: 14

NO: 12

SEQ. ID NO: 33

#10: 18305 to 18306 (791 TO 884)

ATAGTGGGCTGGAAAGTGGCCACAGGTAAAGGTGCACCTTTCTTCCTGGGGATGT GATGTGCATATCACTACAGAAATGTCTTTCCTGAGGTGATGT<u>catgactttqtq</u> <u>qaatqtacacc</u>TGTGACCTCACCCCTCAGGACAGTTTTGAACTGGTTGCTTTCTT TTTATTGTTTAGatcqtctqgtagaatcaacttccttgaqaaagccatagaaac agtqtatqcagcctatttgccaalaacacacacccattcctgtacctcagGTAA TGTAGCACCAAACTCCTCAACCAAGACTCACAAGGAA<u>caqatqttctatcaqqctctcct</u>TTTGAAAGAGATGAGCATGCTAATAGTACAATCAGAGTGAATCCCATAC ACCACTGGCAAAAGGATGTTCTGTCCCTTCTTACAGGTACAAGGCACAG

SEQ. ID

NO: 15

SEQ. ID NO: 34

#11: 18182 to 19041 (885 TO 1038)

CTTACGCAAAGCTACACAGCTCTTAAGTAGCAGTGCCAATATTTGAACACACTCA
GACTCGAGCCTGAGGTTTTGACCACTGTGTCATCTGGCCTCAAATCTTCTGGCCA
CCACATACACCATATGTgqqctttttctccccctccCACTATCTAAGGTAATTGT
TCTCTCTTATTTTCCTGACAGtttagaaatcagtcccagaatgtggatgttaat
gtgcagcccacaaagcatgaagttpacttcctgcacgaggagagacatcctggagc
gqgtgcagcacatcgagagcaagctcctgggctccaattcctccaggatgta
cttcaccaggTCAGGGCGCTTCTCATCCAGCTACTTCTCTGGGGCCTTTGAAAT
GTGCCCGGCCAGAcgtgagagcccagatttTTGCTGTTATTTAGGAACTTTTTTT
GAAGTATTACCTGGATAG

SEQ. ID NO: 16

SEQ. ID NO: 35

Figure 4A - 3

WO 95/16793 PCT/US94/14746

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#12: 18579 to 18178 (1039 TO 1409)

SEQ. ID NO: 36

The splice acceptor site is believed to have 21 T's.

#13: 18420* to 18443 (1410 TO 1558) .

CTGTGCTCCAGCACAGGTCATCCAGCTCTGTAGACCAGCGCAGAGAAGTTGCTTG
CTCCCAAAtgcaacccacaaatttggcTAAGTTTAAAAACAAGAATAATAATGA
TCTGCACTTCCTTTTCTTCATTGCAGaaagagacatcgggaagattctgatgtg
aaatggtggaagatqattcccqaaaggaaatgactgcagcttgtaccccccggag
aaggatcattaacctcactagtgttttgagtctccaggaagaaattaatgagcag
ggacatgaggGTACGTAAACGCTGTGGCCTGCCTGGGATGCATAGGGCCTCAACT
GCCAAggttttggaaatggagaaagCAGTCATGTTGTCAGAGTGGCACTACAGTT
TTGATGGGCAAGCTCCTCTTTCCTTTACTAACCCACAATAGCATCAGCTTAAAGAC
AATTTTTGATTGGGAGAAAAAGGGAGAAAATAATCTCTG

SEQ. ID NO: 37

#14: 19028 TO 18897 (1559 TO 1667)

SEQ. ID NO: 38

Figure 4A - 4

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SEQ. ID NO: 17

SEQ. ID NO: 18

SEQ. ID NO: 19

#15: 19025 to 18575 (1668 TO 1731)

SEQ. ID NO: 20

SEQ. ID NO: 39

-19 of splice acceptor site is A in some people. Others are heterozygous for A and G. GTCACTTC or CTCGCTTC (Polymorphism).

#16: 18184 to 18314 (1732 TO 1896)

CATTTATGGTTTCTCACCTGCCATTCTGATAGTGGATTCTTGGGAATTCAGGCTT
catttqqatqctccqttaaaqcTTGCTCCTTCATGTTCTTGCTTCTTCCTAGgag
ccaqcaccqctctttqaccttqccatqcttqccttagatagtccaqaqagtqqct
gqacaqaqqaaqatqqtcccaaaqaaqqacttqctqaatacattqttqaqttcct
gaaqaaqaaqqctqaqatqcttqcaqactattctcttttqqaaattgatqaqGTG
TGACAGCCATTCTTATACTTCTGTTGTATTCTCcaaataaaatttccaqccqqqt
qCATTGGCTCA

SEQ. ID

SEQ. ID NO: 40

#17: 18429 to 18315 (1897 TO 1989)

CAGATAGGAGGCACAAGGCCTGggaaaggcactggagaaatggqATTTGTTTAAA CTATGACAGCATTATTTCTTGTTCCCTTGTCCTTTTTCCTGCAAGCAGgaaggga acctgattggattaccccttctgattgacaactatgtgccccctttggagggact gcctatcttcattcttcgactagccactgagGTCAGTGATCAAGCAGATACTAAG CATTT<u>cgqtacatgcatqtqtqctgagg</u>qAAAGGGCAAA

SEQ. ID NO: 22

SEQ. ID NO: 41

#18: 18444 to 18581 (1990 TO 2103)

CTATATCTTCCCAGCAATATTCACAGTCCGTTTACAGTTTTAACGCCTAAAGTAT CACATTTCGTTTTTTAGCTT<u>taaqtaqtctqtqatctcq</u>TTTAGAATGAGAATG TTTAAATTCGTACCTATTTTGAGGTATTGAATTTCTTTGGACCAGqtqaattqgg acqaaqaaaaqgaatqttttqaaaqcctcaqtaaaqaatqcqctatqttctattccatccqqqaqcaqtacatatctqaqqaqtcqacctctcaqqccaqcaqGTACAG TGGTGATGCACACTGGCACCCCAGGACTAqqacaqqacctcatacatCTTAGGAG ATGAAACTTG

SEQ. ID NO: 42

Figure 4A - 5
SUBSTITUTE SHEET (RULE 26)

SEQ. ID NO: 23

#19: 18638 to 18637 (2104 TO 2271). 2463 is end of cDNA.

AATCCTCTTGTGTTCAGGCCTGTGGATCCCTGAGAGGCTAGCCCACAAGATCCAC
TTCAAAAGCCCTAGATAACACCAAGTCTTTCCAGACCCAGTGCACATCCCATCAG
CCAGgacaccaqtqtatqttqqGATGCAAACAGGGAGGCTTATGACATCTAATGT
GTTTTCCAGagtgaagtgcctqgctccattccaaactcctqgaagtqgactqtq
aacacattqtctataaagccttqcqctcacacattctqcctcctaaacattcac
aqaaqatqqaaatatcctqcagcttqctaacctqcctgatctatacaaagtcttt
gagaqqtqttaaatatqqttatttatqcactqtqqqatqttcttcttctctq
tatccqatacaaaqtqttqtatcaaaqtqtqatatacaaagtgtaccaacataa
qtqttqqtaqcacttaaqacttatacttqccttctqatagtattcctttatacac
aqtqqqattqattataaataaataqatqtqtcttaacataATTTCTTATTTAATTT
TATTATGTATATA

SEQ. ID NO: 24

SEQ. ID NO: 43

Figure 4A - 6

HMLH1 EXON AMPLIFICATION PRIMERS

First Stage Amplification Primer	SEQ. ID NO:	Second Stage Amplification Primer	SEQ. ID NO:
Exon 1		•	
N-18442- S'aggcactgaggtgattgge C-19109- S'tcgtagcccttaagtgagc	45	N-19295- 5'igtaaaacgacggccagtcactgaggtgattggctgaa C-19446- *5'tagcccttaagtgagcccg	83
Exon 2			
N-19689- 5'aatatgtacattagagtagttg C-19688- 5'cagagaaaggtcctgactc	46	N-18685- 5'tgtaaaacgacggccagttacattagagtagttgcaga C-19067- *5'aggtcctgactcttccatg	85
Exon 3			
N-19687- 5'agagattiggaaaatgagtaac C-19786- 5'acaatgtcatcacaggagg	48 49	N-18687- 5'tgtaaaacgacggccagtttggaaaatgagtaacatgatt C-19068- *5'tgtcatcacaggaggatat	88
Exon 4			
N-18492- 5'aacctttccctttggtgagg C-18421- 5'gattactctgagacctagge	50 51	N-19294- 5'tgtaaaacgacggccagtctttccctttggtgaggtga C-19077- *5'tactctgagacctaggccca	68 80 80
Exon 5			
N-18313- 5'gattttetetttteeeettggg C-18179- 5'caaacaaagetteaacaatttae	53	N-19301- 5'tgtaaaacgacggccagttctcttttcccttgggattag C-19046- *5'acaaagcttcaacaatttactct	91

Figure 4B - Page 1

First Stage Amplification Primer	SEQ. ID NO:	Second Stage Amplification Primer	SEQ. ID NO:
Exon 6		• ,	
N-18318- 5'gggttttattttcaagtacttctatg C-18317- 5'gctcagcaactgttcaatgtatgagc	54 55	N-19711- 5'tgtaaaacgacggccagtgttttattttcaagtacttctatgaatt C-19079- *5'cagcaactgttcaatgtatgagcact	93
Exon 7			
N-19009- 5'ctagtgtgtgtttttggc C-19135- 5'cataaccttatctccacc	56 57	N-19293- 5'tgtaaaacgacggccagtgtgtgtttttggcaac C-19435- *5'aaccttatctccaccagc	98
Exon 8			
N-18197- 5'ctcagccatgagacaataaatcc C-18924- 5'ggttcccaaataatgtgatgg	58 59	N-19329- 5'tgtaaaacgacggccagtagccatgagacaataaatccttg C-19450- *5'tcccaaataatgtgatggaatg	97
Exon 9		·	
N-18765- 5'caaaagcttcagaatctc C-18198- 5'ctgtgggtgtttcctgtgagtgg	60 61	N-19608- 5'-tgtaaaacgacggccagtaagcttcagaatctctttt C-19449- *5'-tgggtgtttcctgtgagtggatt	99
Exon 10			
N-18305- 5'catgactttgtgtggaatgtacacc C-18306-5'gaggagagcctgatagaacatctg	62 63	N-19297- 5'tgtaaaacgacggccagtactttgtgtgaatgtacacctgtg C-19081- *5'gagagcctgatagaacatctgttg	101

Figure 4B - Page 2

Figure 4B - Page 3

First Stage Amplification Primer	SEQ. ID NO:	Second Stage Amplification Primer	SEQ. ID NO:	
Exon 11				
N-18182- 5'gggctttttctcccctcc C-19041- 5'aaaatctgggctctcacg	65	N-19486- 5'tgtaaaacgacggccagtctttttctcccctcccacta C-19455- *5'tctgggctctcacgtct	103	
Exon 12 (See note at end)				
N-18579- 5'aattatacctcatactagc C-18178- 5'gttttattacagaataaaggagg	99	N-20546- *5'cttattctgagtctctcc C-20002- 5'tgtaaaacgacggccagtgtttgctcagaggctgc	105 106	
		N-19829- *5'gatggttcgtacagattccg C-19385- 5'tgtaaaacgacggccagtttattacagaataaaggaggtag	107	12
Exon 13				/24
N-18420- 5'tgcaacccacaaaatttggc C-18443- 5'etttetecatttecaaaacc	69	N-19300- 5'tgtaaaacgacggccagtaacccacaaaatttggctaag C-19078- *5'tctccatttccaaaaccttg	109	
Exon 14				
N-19028- 5'tggtgtctctagttctgg C-18897- 5'cattgttgtagtagctctgc	71 72	N-19456- *5'tgtctctagttctggtgc C-19472- 5'tgtaaaacgacggccagttgttgtagtagctctgcttg	111	
Exon 15				
N-19025- 5'cccatttgtcccaactgg C-18575- 5'cggtcagttgaaatgtcag	73	N-19697- *5'atttgtcccaactggttgta C-19466- 5'tgtaaaacgacggccagttcagttgaaatgtcagaagtg	113 114	

	First Stage Amplification Primer	SEQ. ID NO:	Second Stage Amplification Primer	SEQ. ID NO:
	Exon 16			
	N-18184- 5' cattiggatgeteegttaaage C-18314- 5' caceeggetggaaattttattig	75 76	N-19269- 5'tgtaaaacgacggccagt C-19047- *5'ccggctggaaattttatttggag	115 116
	Exon 17			
9	N-18429- 5'ggaaaggcactggagaaatggg C-18315-5'ccctccagcacacatgcatgtaccg	77	N-19298- 5'tgtaaaacgacggccagtaggcactggagaaatgggatttg C-19080- *5'tccagcacacatgcatgtaccgaaat	117
SHRST	Exon 18			
ITIITE QU	N-18444- S'taagtagtetgtgateteeg C-18581- S'atgtatgaggteetgtee	79	N-19436- *5'gtagtctgtgatctccgttt C-19471- 5'tgtaaaacgacggccagttatgaggtcctgtcctag	119 120
EET /D	Exon 19			
ili E acı	N-18638- S'gacaccagtgtatgttgg C-18637- S'gagaaagaagaacacatcc	81 82	N-19447- *5'accagtgtatgttgggatg C-19330- 5'tgtaaaacgacggccagtgaaagaagaacacatcccaca	121

All sequence reads 5' to 3'. Primer identification numbers are listed before each primer sequence. N indicates the primer on the 5' side of the exon. * indicates that the 5' nucleotide is biotinylated.

Figure 4B - Page 4

296 380 479 571 581 663 676 100 97 200 197 MSFVACVĮRKLIDETVVNRIAAGBVIORBANIKEMIENCIDIKSIISIOVIVKEGGIKLIDIODNGISIRKEDIDIVČERFTYSKI.GSFEDIJASIISTYGFR MSLR---LIKALDASVVNKIAAGBIIISBVNIKEMENSIDINAIMIDIIVKEGGIBVIDITDNGSGINMADIPIILČERFYYSKI.GKFEDISCIICTYGFB CEALASISHVAHVTITTRTADGKCAMBASYSDGRILKAFPKRÇAGNGSTQTTVEDLIFYNTATRKATIKNPSEENGKITEVVGRYSVANAGTSFSVAGGET CEALASISHVARVTVTKVKEDRCAMBVSVAEGBALESPKBVAGKDSTTUTVEDLIFNIPSBLRAIRSHNDENSKIIDVVGRYALHSKUJGESGAGFGDS ETSHONVDVNVHPTRHENHEDESTLERVQQHIESRFLGSNSSRNYFT-----QTFLEGLAGPSGENVKST-----TFLFSSSTSGSSCFVVA VIOBANDVNVHPTRESTREJSQDETIEKIANQLHAETSAIDTSFTFRASSISTNRPESTLEFNDTIESDRNRKSLRQAQVVENGYTANSQLRKAFRQE VADVRTIGNASTVONINIRSIGNAVSRELIJEI---GGEDKTGAFKANGYISVANYSVKOGI-FLIFTNHELVESTSGAKAIETVAAAYLPRATHFIKNISL NYSLSVKJSYTVODRIGTVONKSVASNIJTFHISKVEDIAJE-SVIGKVONINFISKOGIJSLIFETINRUVTCDILIBRALNSVJSVVLPRGFREJIVIGI HOMMTIDSRECKILDAFILOPLIGKPLSSOPQAIVTEDKTDISSGRARCTOFFALIETIPAPAEVAAKNOSLFICDTTKGTSEMSEKRGPTSSAFFRHR-EFISCK NKIMFIDASOABITSFIJSS-1900FNFEGSSTKROLSEPKVTNVSHSDEABALJTLN-------FISEOPRDANTINDND--LKDCJEKAKOKLCTJYNJ ENVEDDSRKEMTAA-----CTRRRRI-INLFBVLSDOHEINEQSHEMDRANSFVGGANPOMALA--DHOTKUMDLMTTKLSEELFYQILIYDFANH PSIADDERNALPISKDGYIRVBKEBVNVNLTGIKKUMBKVDDSIHREUTDIFANLMYNGVNDEERRUJAIDHDUKUHUUDDGSVCYELFYQIGLIDFANH fylefysepaplefylamlafingeresepspektiglaeytigefilmmanterigleriden ett ett mynderigleriden ett ett en fynderische Krintystinystotynytig-bedeintbaskok----offskindnsminenngibundiondiksyntikselulukoligsyngeresiyng Nambeffregfeslskfigdarfa--sirkoviseestlgcosevpgsipnswkatvæfivykalrshifffraffriedgnilolanlpdlykvferg Neweggegldgilffigiblegippovpkvdtldaslgedekaqfinrkehissllehylfpcikrrflaffrejilm--vveianlpdlykvferg HUMAN 'FAST /EAST YEAST HUFTAN HUMAN HUMAN YEAST YEAST HUMAN TEAST HUMAN HUMAN HUMAN

SEQ. ID NOS

and 12

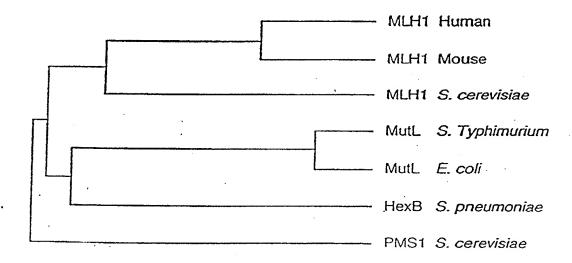


Figure 6

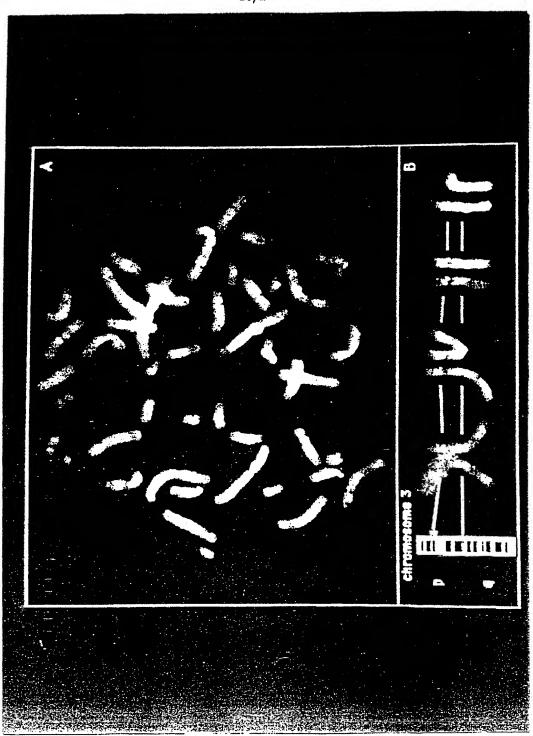


Figure 7
SUBSTITUTE SHEET (RULE 26)

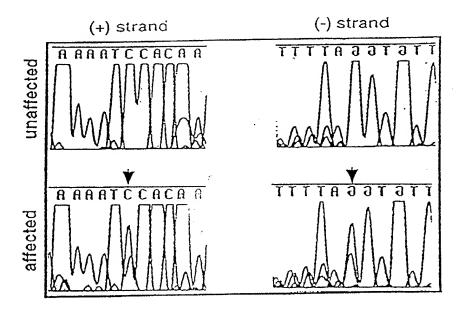


Figure 8

SEO. ID NO

VNRIAAGEVIQRPANAIKEMIENCLDAKFTSIQVIVKEGGLKLIQIQDNGTGIRKEDLDIVCER VNRIAAGEVIQRPANAIKEMIENCLDAKSTSIQVIVKEGGLKLIQIQDNGTGIRKEDLDIVCER affected normal human MLH1 human MLH1

ANQIAAGEVIERPASVCKELVENAIDAGSSQIIIEIEEAGLKKVQITDNGHGIAHDEVELALRR ANQIAAGEVVERPASVVKELVENSLDAGATRIDIDIERGGAKLIRIRDNGCGIKKDELALARA ANQIAAGEVVERPASVVKELVENSLDAGATRVDIDIERGGAKLIRIRDNGCGIKKEELALALARPANAIKEMIENCLDAKSTNIQVVVKEGGLKLIQIQDNGTGIRKEDLDIVCER VNKIAAGEIIISPVNALKEMMENSIDANATMIDILVKEGGIKVLQITDNGSGINKADLPILCER VHRITSGQVITDLTTAVKELVDNSIDANANQIEIIFKDYGLESIECSDNGDGIDPSNYEFLALK HexB Muth Muth PMS1 S. cerevisiae N. cerevisiae F. E. coli mouse typhimurium pneumoniae

126 127 128 129 130

Figure 9

human PMSI nucleotide sequence. The putative start (ATG) and stop (TGA) codons are underlined.

	80	240	320	480	560	640	720	800	880	960	1130	1200	1280	1360	1440	1520	1600	1680	1760	1840	1920	2000	2080	2160	2240	2320	2400	2480	2560	2640	2687	
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SEQ. ID NO: 132

662 691

Alignment Report of PMS1, using Clustal method with PAM250 residue weight table. Wednesday, January 26, 1994 4:51 PM

Wednesday, January 26, 1994 4:51 PM	PRSI_HUMAN M-ERABSSSTBPARATKPIDRKSPHOTOSGOVILSISTAVKELVENGTODACATWIDLKIKDYGVITIBVSDNGGEERNF 80	PHSI_HUMAN BGÜTÜKHHTSKIJOERADILTGVERFRGEALSSIZALSDVIISTCHASARVGTRÜMEDHAGKÜIÇKÜPYERPGTIVSQOLASTLEVRHKERORNIRKE 180	PMSI_HUMAN YARMVQVLHANGIISRGIRVGCINQLGGGRRQPVVCNGGGSFGIRENIGSVFGQRQLQSN—IPFN—CNFR-CNFR-SDSVCEENGLSCSDALHNUFN—1	PHSI_HUMAN SOCTHOVGESTGEDERGEFFERENDERVORLVNEVHEWIRFOWENTENDSECVOTWVTPDKOGILLIQEEKLLLAVIKTGIIGMEDSDVNKOMS 373	PMS1_HUMAN QOPLIDVBGNLIR-MHAADLEKPMYEKODOSPSLAFIGEEKKDVGISRLREAFISLRHTTENFPHSPKTPEPRRSPLGOKRGMLSGSTSGAISDKGVTRSQK 472
	PRSI_YEAST MFHHIBNLLIBTEBRCKQKEQRYIPVKYLFSWTQIHQINDIDVHRITISGOVITTITTAVKELVINGIDANANQIBIIFKDYGLESIEGSDNGGIDPSYY 100	PHSI_YEAST BEUALKHYTSKIJAKBGÜVAKVIGILGERGEALSSIZGIAKUSVIÜTTSPERAD-KÜEYÜMVGHÜTSKUTTSENKETIVLUSOLAHULEVRIKERROR 199	PMSI_YEAST FTECLTNIQGNALINDAJIKFGVANITTPKGKRALILSJMRNSGNRKANISSVFGAGGWRGLEEVDLMLDLINDFRORMLGRYTDD—PDFLDLDMKIRVKGYNJ 297	PHSI_YEAST SONSEGCENGERDROBIYVANKEBVEYSTLLKCCNEVIKTENNVOFBANFLANELPMSLIDVAVTPDKOVILLIGAGEAVIDIFKTOSDYVNRQEDALP 395	PMS1_YEAST_KRMCSOSBOOACHTLKTEVFDDRSTTHESDNEAYHJARSESNGENHAMENSTTGVIDSNGTELTSVMDGNYTNVTDVIGGECEVSVDSSVAJJDEGN 492

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PHSI_HUMAN EÜRKEJSHTWERENTIGETITKLAREDIFTVDOHAPDEKYNFEM GOHTMEGGRIJAPGTLAUTAANBANLIENDEJHRKWGFDFVIDEN 75	PMSI_HUMAN APVTERAKUISLPTSKNWTGGPGDVTELJFMLSDSFGVMGREGRVKOVFASRACHKGVMIGTALNTSFYKKLITHMGEMGHPWNCPHGRPTMRHIAN 85
PHSI_YEAST YÜTLTVSKAUHKRAHVVGOFNIGELIVTRUVDNKSDIFTVDOHASDEKYNFEMIGKLATREGKLIJIPGFVFUSVIDELVVLAUIPVUHRKUNGHKKIDHE 79	PMSI_YEAST BEFGSGVKULSLPTSKOTUBDLGDFNELJHLIKEDGGLRRUNTBGGKIRSMEJMRACHSGIMIGKFLNKKTEVTRVVHNLSELDKPWNCPHGRPTMRHIME 89

855 891 862 904

758 791

> PMS1_HUMAN L---GVISQN PMS1_YEAST IRDMSSFISKDYEI

Decoration 'Decoration #1': Box residues that match the consensus named 'Consensus #1' exactly.

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Partial nucleotide sequence of mouse MLH1 cDNA. The putative stop (TAA) codon is underlined
                                                                                 GAGCAAGTTA
                                              TTCAAGTGGT
                                                           GATATTGTGT
                                                                       TGGTGAGCAT
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30. ID NO: 135

694

SEO, ID NO: (Note: reading frames of both are pieced together to include those with strong similarity to yeast MLH1, not based on similarity with Comparison of the predicted amino acid sequences for mMLH1 and hMLH1 proteins. Vertical lines indicate amino acid identities.

each other)

GFR 73 136 111 GFR 100	GET 173 111 GET 200	NQP 272 13P 300	22/ 000 7407
house 1	74 GEHLASISHVAHVTITTKTADGKCAYRASYSDGKLQAPPKPCAGNQGTLITVEDLFYNIITRRKALKNPSEEYGKILEVVGRYSIHNSGISISVKKQGET 173 11	174 VSDVRTLPNATTVDNIRSIFGNAVSRELIEVGCEDKTLAFKANGYISNAKYSVKKCIFLLFINHRLVESAALRKAIETVYAAYLPK-THTHSCTSVZNQP 272 	273 SERDVNVHPTKTEVHFLHEESILQRVQQHIESKLLGSNSSRMVFHPDLASRTCWASGEAARPTTGVASSSTSGSGDKVYAYQMSRTDSRDQKLDAFLQFV 372
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rriinltsvlslqeeiserchetlreilrnhsfvgcvnpqwalaqhqtklyllnttklseelfyqiliydfanfgvlrlsepaplfdlamlaztvlkva 572 SKPLSSQPQA--IVTEDKTDISSGRARQQDEEMLELPAPAEVAAKNQSLEGDTTKGTSEMSEKRGPTSS--NPRKRHREDSDVEMVEDDSRKEMTAACTP 496 RRRIINLTSVLSLQEEINEQGHEVLREMLHNHSFVGCVNPQWALAQHQTKLYLLNTTKLSEELFYQILIYDFANFGVLRLSEPAPLFDLAMLA--LDSPE 401 473

SSLVPSQPQDPRPVRGARTEGSPERATREDEEMLALPAPAEAAESENLERESLMETSDAAQKAAPTSSPGSSRKSHREDSDVEMVENASGKEMTAACYP

22/24

472

GORTTAR-RRACRVHCRVSEEKRDACRLFSVRSMRREPDZ-----LLFZZQLCATFGGTAYLHSSTGHZGELGEEKECFESLSKECAMFYSIRKQYILEE 573 497

SGWTEEDGPKEGLAEYIVEFLKKKAEMLADYFSLEIDEEGNLIGLPLLIDNYVPPLEGLPIFILRLATEVNWDEEKECFESLSKECAMFYSIRKQYISEE STLSGQQSDMPGSTSKPWKWTVEHIIYKAFRSHLLPPKHFTEDGNVLQLANLPDLYKVFERC 728

STLSGQQSEVPGSIPNSWKWTVEHIVYKALRSHILPPKHFTEDGNILQLANLPDLYKVFERC 756 695

667

Figure 13

373

Figure 14

SEQ. ID NO: 137

862

FMLSDSPGVMCRPSRVRQMFASRACRKSVMIGTALNASEMKKLITHMGEMDHPWNCPHGRPTMRHVANLDVISQN

FMLSDSPGVMCRPSRVKQMFASRACRKSVMIGTALNTSEMKKLITHMGEMGHPWNCPHGRPTMRHIANLGVISQN

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INTERNATIONAL SEARCH REPORT

cernational application No. PCT/US94/14746

1	ASSIFICATION OF SUBJECT MATTER		
	:Please See Extra Sheet. :Please See Extra Sheet.		
According	to International Patent Classification (IPC) or to both	national classification and IPC	
	LDS SEARCHED		
	documentation searched (classification system followed	d by classification symbols)	
U.S. :	Please See Extra Sheet.		
Documenta	tion searched other than minimum documentation to th	e extent that such documents are included	in the fields searched
	data base consulted during the international search (name ee Extra Sheet.	ame of data base and, where practicable	, search terms used)
C. DOC	UMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where a	ppropriate, of the relevant passages	Relevant to claim No.
A	JOURNAL OF MOLECULAR BI ISSUED 1986, GRANGER-SCHNA OF N-ACETOXY-N-2-ACETYLAN FRAMESHIFT MUTATION SPECTR DEFICIENT ESCHERICHIA COLI S' U", PAGES 499-507, SEE ENTIRE	RR ET AL., "SPECIFICITY MINOFLUORENE-INDUCED UM IN MISMATCH REPAIR TRAINS MUTH, L, S AND	1-55
Y	JOURNAL OF BACTERIOLOGY, VOISSUED OCTOBER 1989, FOUNDED OCTOBER 1989, FOUNDED OF SEQUENCE OF PNEUMONIAE HEXB MISMATCH ROF HEXB TO MUTL OF SALMONE TO PMS1 OF SACCHAROMYCE 5332-5338, SEE ESPECIALLY TO DISCUSSION AT PAGE 5336, SEC PARAGRAPH.	26,27, 36-45, 47-55	
X Furth	er documents are listed in the continuation of Box C	See patent family annex.	
Spe	cial categories of cited documents:	"T" later document published after the inte	
'A' doc to b	nument defining the general state of the art which is not considered be of particular relevance	date and not in conflict with the applic principle or theory underlying the inv	
"L" doc	lier document published on or after the international filing date ument which may throw doubts on priority claim(s) or which is	"X" document of particular relevance; the considered novel or cannot be conside when the document is taken alone	
cite	d to establish the publication date of another citation or other cial reason (as specified)	"Y" document of particular relevance; the	
O° doc	nument referring to an oral disclosure, use, exhibition or other	considered to involve an inventive combined with one or more other suc being obvious to a person skilled in the	h documents, such combination
'P" doc	ument published prior to the international filing date but later than priority date claimed	*&* document member of the same patent	
	actual completion of the international search	Date of mailing of the international sea	arch report
29 MARC	H 1995	10APR1995	
Commission Box PCT	nailing address of the ISA/US ner of Patents and Trademarks	Authorized officer Outona	h Freise Pe
Washington Facsimile No	, D.C. 20231 n. (703) 305-3230	Telephone No. (703)308-0196	

Form PCT/ISA/210 (second sheet)(July 1992)★

INTERNATIONAL SEARCH REPORT

ernational application No. PCT/US94/14746

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	JOURNAL OF BACTERIOLOGY, VOLUME 171, NUMBER 10, ISSUED OCTOBER 1989, MANKOVICH ET AL., "NUCLEOTIDE SEQUENCE OF THE SALMONELLA TYPHIMURIUM MUTL GENE REQUIRED FOR MISMATCH REPAIR: HOMOLOGY OF MUTL TO HEXB OF STREPTOCOCCUS PNEUMONIAE AND TO PMS1 OF THE YEAST SACCHAROMYCES CEREVISIAE", PAGES 5325-5331, SEE ESPECIALLY THE ABSTRACT AND THE DISCUSSION SECTIONS.	26, 27, 36-45, 47-55
Y	GENETICS, VOLUME 110, ISSUED AUGUST 1985, WILLIAMSON ET AL, "MEIOTIC GENE CONVERSION MUTANTS IN SACCHAROMYCES CEREVISIAE: I. ISOLATION AND CHARACTERIZATION OF PMS1-1 AND PMS1-2", PAGES 609-646, SEE THE ENTIRE DISCLOSURE.	1-55
	NATURE, VOLUME 365, ISSUED 16 SEPTEMBER 1993, STRAND ET AL., "DESTABILIZATION OF TRACTS OF SIMPLE REPETITIVE DNA IN YEAST BY MUTATIONS AFFECTING DNA MISMATCH REPAIR", PAGES 274-276, SEE ENTIRE DISCLOSURE.	26, 27, 36-45, 47-55
	JOURNAL OF BACTERIOLOGY, VOLUME 171, NUMBER 10, ISSUED OCTOBER 1989, KRAMER ET AL., "CLONING AND NUCLEOTIDE SEQUENCE OF DNA MISMATCH REPAIR GENE PMS1 FROM SACCHAROMYCES CEREVISIAE: HOMOLOGY OF PMS1 TO PROCARYOTIC MUTL AND HEXB", PAGES 5339-5346, SEE ESPECIALLY THE ABSTRACT AND DISCUSSION SECTIONS.	26,27, 36-45, 47-55

INTERNATIONAL SEARCH REPORT

aternational application No. PCT/US94/14746

A. CLASSIFICATION OF SUBJECT MATTER: IPC (6):

C12Q 1/68; C07H 21/00,21/02,21/04; C12P 19/34; C07K 13/00

A. CLASSIFICATION OF SUBJECT MATTER: US CL :

435/6,91.2; 530/350,387,1; 536/23.1,24.3,24.31.24.33

B. FIELDS SEARCHED
Minimum documentation searched
Classification System: U.S.

435/6,69.3,91.1,91.2,810; 530/350,387.1,388.1; 536/23.1,24.3,24.31,24.33; 935/77,78

B. FIELDS SEARCHED Electronic data bases consulted (Name of data base and where practicable terms used):

APS, CAS ONLINE, MEDLINE, BIOTECH ABS, WPI, BIOSIS search terms: cancer,mlh1,mlh2,pms1,mutl.pmlh2,pmutl.ppms1,mismatch,repair

No of the second